1	The LC3B FRET biosensor monitors the modes of action of
2	ATG4B during autophagy in living cells
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6 Supplementary figures



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Figure S1. The G120A LC3B biosensor is not sensitive to autophagy induction and/or
lysosomal inhibition, it does not colocalize with LAMP2, and the expression of the WT
biosensor does not perturb the abundance of the endogenous LC3B protein. (A) Representative
fluorescence images of U2OS expressing the G120A biosensor and stained for endogenous

LAMP2. To investigate the changes in Aqua-LC3B puncta numbers and their colocalization 12 with LAMP2, cells were treated with: DMSO (6 h), BafA₁ (6 h, 100 nM), torin1 (3 h, 250 nM), 13 torin1 (3 h, 250 nM) + BafA₁ (6 h, 100 nM), HBSS (1 h), HBSS (1 h) + BafA₁ (6 h, 100 nM). 14 Scale bar: 9 µm. (B) Representative western blotting images and corresponding quantifications 15 (C) of total lysates from U2OS cells expressing the empty vector or the WT LC3B biosensor 16 and treated as indicated. Loading control: Actin. n = 3 independent experiments. ns (not 17 significant) as determined by two-way ANOVA with two-stage step-up method of Benjamini, 18 Krieger and Yekutieli's multiple comparison test to control the false discovery rate in (C). 19



Figure S2. The WT or G120A donors do not respond to ATG4B silencing. (A) Representative fluorescence and Δ Lifetime images of U2OS cells co-expressing the WT or G120A donor with control or ATG4B-specific siRNAs, and analyzed by FRET-FLIM. Squares on the top images of WT or G120A donor panels illustrate the location of the enlarged images. Dotted lines on

25 the enlarged images illustrate where the line analysis was performed. Pseudocolor scale: pixelby-pixel ΔLifetime. Scale bars: overviews, 40 μm; enlarged, 6 μm. (B) Mean ΔLifetime 26 analysis of U2OS cells co-expressing the WT or G120A donor with control or ATG4B siRNA. 27 Representative western blotting images (C) and corresponding quantifications (D) of total 28 lysates from U2OS cells co-expressing the WT or G120A biosensor with control or ATG4B-29 specific siRNAs. IB1 and IB2 correspond to the same lysates blotted for overexpressed (IB1) 30 or endogenous (IB2) LC3B forms. Loading control: Actin. n = 3 independent experiments. (E) 31 Line and (F) number of Aqua-LC3B-II puncta analyses of U2OS cells co-expressing the WT 32 or G120A donor with control or ATG4B siRNA. n = 10 cells per condition from one 33 representative experiment (of three) in (B) and (F). *P < 0.05, ****P < 0.0001, ns (not 34 significant) as determined by two-way ANOVA with Tukey's multiple comparison test in (B) 35 and (F), and with two-stage step-up method of Benjamini, Krieger and Yekutieli's multiple 36 comparison test to control the false discovery rate in (**D**). 37



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Figure S3. No FRET is detected when donor-bound LC3B and acceptor-bound LC3B are coexpressed. (A) Representative fluorescence and Δ Lifetime images of U2OS cells co-expressing the WT or G120A biosensor, or donor (Aquamarine-proLC3B) + acceptor (proLC3B-

tdLanYFP) constructs with control or ATG4B-specific siRNAs, and analyzed by FRET-FLIM. 42 Squares on the top images of WT or G120A biosensor, or donor + acceptor panels illustrate 43 the location of the enlarged images. Dotted lines on the enlarged images illustrate where the 44 line analysis was performed. Pseudocolor scale: pixel-by-pixel Δ Lifetime. Scale bars: 45 overviews, 40 μ m; enlarged, 6 μ m. Mean Δ Lifetime (**B**), number of high Δ Lifetime pixels (**C**), 46 line (**D**), and histogram (**E**) analyses of U2OS cells co-expressing the WT or G120A biosensor, 47 or donor + acceptor constructs with control or ATG4B-specific siRNAs in (**B**), (**D**), and (**E**), 48 and the WT donor, biosensor or donor + acceptor constructs with control or ATG4B siRNA in 49 (C). Vertical dotted lines on each histogram depict the mode value in (E). n = 10 cells per 50 condition from one representative experiment (of three) in (**B**), (**C**), and (**E**). *P < 0.05, **P <51 0.01, ****P < 0.0001, ns (not significant) as determined by two-way ANOVA with Tukey's 52 multiple comparison test in (B) and (C). 53



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Figure S4. The LC3B biosensor and endogenous LC3B proteins cannot be primed in cells
lacking ATG4B and the ectopic expression of active ATG4B does not alter LC3B donor-only
Δlifetime variations in *ATG4B* SKO cells. (A) Representative western blotting images of total

58 lysates from HeLa control, ATG4B SKO, ATG4A ATG4B DKO, ATG4A ATG4B ATG4C TKO cells expressing the WT or G120A biosensor. IB1 and IB2 correspond to the same lysates 59 blotted for overexpressed (IB1) or endogenous (IB2) LC3B forms. Loading control: Actin. n =60 3 independent experiments. Mean Δ Lifetime (**B**) and number of high Δ Lifetime pixels (**C**) 61 analyses of control and ATG4B SKO HeLa cells co-expressing the WT donor with an empty 62 vector, or with vectors expressing WT ATG4B, ATG4B^{C74S} or ATG4B^{W142A}. (**D**) Histogram 63 analysis of HeLa control cells co-expressing the WT biosensor with an empty vector, or with 64 vectors expressing WT ATG4B, ATG4B^{C74S} or ATG4B^{W142A}. Vertical dotted lines on each 65 histogram depict the mode value. n = 10 cells per condition from one representative experiment 66 (of three) in (**B**), (**C**) and (**D**). (**E**) Representative western blotting images and corresponding 67 ATG4B:Actin quantification of total lysates from HeLa control and ATG4B SKO cells co-68 expressing the WT biosensor with an empty vector, or with vectors expressing WT ATG4B, 69 ATG4B^{C74S} or ATG4B^{W142A}. IB1 and IB2 correspond to the same lysates blotted for 70 71 overexpressed (IB1) or endogenous (IB2) LC3B forms. Short and long ATG4B exposures are shown. Loading control: Actin. n = 3 independent experiments. ns (not significant) as 72 determined by two-way ANOVA with Tukey's multiple comparison test in (B) and (C), and 73 with two-stage step-up method of Benjamini, Krieger and Yekutieli's multiple comparison test 74 to control the false discovery rate in (E). 75



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Figure S5. The number of Aqua-LC3B-II puncta and the mean Δ Lifetime values do not correlate. Correlation analyses of Aqua-LC3B-II puncta and the mean Δ Lifetime values of HeLa cells expressing the WT biosensor and treated with DMSO (6 h), MG132 (6 h, 1 μ M), tioconazole (6 h, 4 μ M), LV-320 (6 h, 120 μ M), FMK-9a (6 h, 10 μ M), NSC 185058 (6 h, 100

- 81 μ M), or Z-L-Phe chloromethyl ketone (6 h, 3 μ M). n = 10 cells per condition from one
- 82 representative experiment (of three).





Figure S6. Treatment with FMK-9a, NSC and ZPCK results in the accumulation of the unprimed biosensor and FRET-FLIM data analyzed by using sensitive methods reveal the

86 mode of action of tioconazole and LV-320. (A) Representative western blotting and corresponding Aqua-proLC3B-tdLanYFP:Actin quantification of total lysates of HeLa cells 87 expressing the WT or G120A biosensor and treated with DMSO (6 h), or expressing the WT 88 89 biosensor and treated with tioconazole (6 h, 4 µM), LV-320 (6 h, 120 µM), FMK-9a (6 h, 10 μ M), NSC 185058 (6 h, 100 μ M), or Z-L-Phe chloromethyl ketone (6 h, 3 μ M). IB1 and IB2 90 91 correspond to the same lysates blotted for overexpressed (IB1) or endogenous (IB2) LC3B forms. Aqua-proLC3B-tdLanYFP:Actin quantification is performed by normalizing the 92 density of Aqua-proLC3B-tdLanYFP band in IB1 to that of the respective loading control in 93 IB2. Loading control: Actin. n = 3 independent experiments. *P < 0.05, ****P < 0.0001, ns 94 (not significant) as determined by one-way ANOVA with two-stage step-up method of 95 Benjamini, Krieger and Yekutieli's multiple comparison test to control the false discovery rate. 96 97 (B) Representative fluorescence and Δ Lifetime images of HeLa cells expressing the WT or G120A biosensor, treated with DMSO (6 h), and analyzed by FRET-FLIM. Representative 98 fluorescence and ALifetime images of HeLa cells expressing the WT biosensor and treated 99 100 with the following compounds: tioconazole (6 h, 4 μ M) (C) and LV-320 (6 h, 120 μ M) (D). Squares on the top images of WT or G120A biosensor panels illustrate the location of the 101 enlarged images. Dotted lines on the enlarged images illustrate where the line analysis was 102 performed. Pseudocolor scale: pixel-by-pixel \(\Delta\)Lifetime. Scale bars: overviews, 40 \(\mu\)m; 103 enlarged, 6 μ m. Number of high Δ Lifetime pixels analysis of HeLa cells expressing the WT 104 105 donor or biosensor and treated with tioconazole (6 h, 4 μ M) (C) or LV-320 (6 h, 120 μ M) (D). Line, and histogram analyses of HeLa cells expressing the WT or G120A biosensor and treated 106 with tioconazole (6 h, 4 μ M) (C) or LV-320 (6 h, 120 μ M) (D). n = 10 cells per condition from 107 one representative experiment (of three) in (C, D). ns (not significant) as determined by two-108 way ANOVA with Tukey's multiple comparison test in (C, D). 109



Figure S7. The LC3B biosensor reveals the mode of action of MG132 in cells. (A and B)
Representative fluorescence and ΔLifetime images of HeLa cells expressing the WT or G120A

113 biosensor, treated with DMSO (6 h) or MG132 (6 h, 1 µM), and analyzed by FRET-FLIM. Pseudocolor scale: pixel-by-pixel Δ Lifetime. Scale bars: overviews, 40 µm; enlarged, 6 µm. 114 Mean Δ Lifetime (**B**), number of Aqua-LC3B-II puncta (**B**), histogram (**C**), and line (**D**) 115 analyses of HeLa cells expressing the WT or G120A biosensor and treated with DMSO (6 h) 116 or MG132 (6 h, 1 μM). Number of high ΔLifetime pixels analysis (**B**) of HeLa cells expressing 117 the WT donor or biosensor and treated with DMSO (6 h) or MG132 (6 h, 1 µM). Vertical 118 dotted lines on each histogram depict the mode value in (C). n = 10 cells per condition from 119 one representative experiment (of three) in (B) and (C). (E) Representative western blotting of 120 total lysates of HeLa cells expressing the WT or G120A biosensor and treated with DMSO (6 121 h), or expressing the WT biosensor and treated with MG132 (6 h, 1 µM). IB1 and IB2 122 correspond to the same lysates blotted for overexpressed (IB1) or endogenous (IB2) LC3B 123 forms. Loading control: Actin. n = 3 independent experiments. **P < 0.01, ****P < 0.0001, 124 ns (not significant) as determined by two-way ANOVA with Tukey's multiple comparison test 125 in (**B**). 126

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