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### ► To cite this version:

C. Rouquie, S. Liu, Murielle Rabiller-Baudry, A. Riaublanc, M. Frappart, et al.. Electrokinetic leakage as a tool to probe internal fouling in MF and UF membranes. *Journal of Membrane Science*, 2020, 599, pp.117707. 10.1016/j.memsci.2019.117707 . hal-02498315

**HAL Id: hal-02498315**

**<https://univ-rennes.hal.science/hal-02498315v1>**

Submitted on 27 Mar 2020

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# Electrokinetic leakage as a tool to probe internal fouling in MF and UF membranes

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## Abstract

Tangential electrokinetic measurements are widely used to characterize membrane fouling as the membrane zeta potential is partly governed by the presence of foulant materials on its surface. However, in the case of porous materials as micro- (MF) and ultrafiltration (UF) membranes, a part of the streaming current flows through the porosity of the membrane during measurements. This electrokinetic leakage, is directly impacted by the presence of foulant materials inside the membrane porosity. Hence, this paper investigates for the first time the possibility of using electrokinetic leakage as a probe for detecting internal fouling, taking lipid fouling as example. Firstly, a lab-scale methodology combining “upside-down” fouling experiments with electrokinetic measurements demonstrated that the intensity of the electrokinetic leakage was related to the presence of internal fouling. Secondly, the concept was applied to the pilot-scale MF and UF of an oil-in-water emulsion

23 under various transmembrane pressures (TMP). A significant impact of the TMP on the internal  
24 fouling of a MF PES membrane was highlighted, whereas almost no impact of the TMP was noticed in  
25 the case of an UF PAN membrane. The developed methodology using the quantification of the  
26 electrokinetic leakage phenomenon allows distinguishing the contributions of internal and external  
27 (surface) fouling. These findings offer new application of tangential electrokinetic measurements to  
28 gain more insight into the characterization of membrane fouling.

29

30 **Highlights:**

- 31 • New characterization of internal fouling by tangential electrokinetic measurement
- 32 • Internal and external fouling contributions to membrane electrokinetic properties
- 33 • Impact of the operating transmembrane pressure on internal fouling occurrence

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35 **Keywords:** Membrane fouling; electrokinetics; streaming current; microfiltration; ultrafiltration

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# 46 1 Introduction

47 Despite the many advantages and applications of membrane filtration, performances keep being  
48 hampered by membrane fouling due to deposit and/or adsorption of compounds on the membrane  
49 surface and/or inside the membrane porosity. In the light of this, there is an important need of  
50 characterization methods in order to strengthen membrane fouling comprehension and thus improve  
51 fouling control during large-scale filtrations.

52 Many strategies have been applied so far to characterize membrane fouling, starting with  
53 conventional flux/pressure monitoring often combined with prediction or comprehension models  
54 (resistance-in-series, pore blocking, inertial lift models, etc.). These widely used approaches are  
55 powerful tools allowing describing membrane fouling through the estimation of various parameters:  
56 resistance due to membrane fouling, reversibility of membrane fouling, main fouling mechanisms,  
57 filtration cake thickness and porosity, etc. However, their findings are based on estimation models  
58 themselves based on macroscopic scale parameters (flux and pressure) and can be far from the  
59 microscopic reality occurring around the membrane surface [1].

60 In view of the foregoing, advanced techniques allowing fouling characterization directly on the  
61 membrane surface have been gaining more and more interest these past years, as they can provide  
62 precise information about fouling composition, concentration and location on the membrane. The  
63 major techniques used for membrane surface characterization have been reviewed by Johnson et al.  
64 [2]. It includes spectroscopic techniques such as ATR-FTIR, Raman and XPS spectroscopy, mainly  
65 used to investigate the chemical nature of foulant compounds, their location and even their  
66 concentration on the membrane surface [3–5]. Imaging techniques (AFM, SEM, TEM, etc.) are also of  
67 great interest for membrane fouling characterization as they can bring information about fouling  
68 layer thickness and distribution [1]. The study of the modification of the membrane wetting  
69 properties (contact angle) and electrokinetic behavior (electrophoresis, streaming and  
70 sedimentation potential measurements) induced by the presence of foulant materials on its surface  
71 bring useful information as well [2].

72 However, most of these techniques mainly focus on the membrane surface characterization. Few  
73 studies deal with the characterization of internal fouling, even though this latter is often irreversible  
74 and causes severe loss of performances and cleaning overcost. Yeo et al. [6] managed to monitor

75 external and internal organic fouling deposition on and in a hollow fiber membrane using phase-  
76 contact XMI, while 3D optical coherence tomography was successfully used by Trinh et al. [7] to  
77 characterize external and internal fouling during microfiltration of oil-in-water emulsions. Similarly,  
78 electrochemical impedance spectroscopy has been recently used to identify the main fouling  
79 mechanism (external fouling layer or irreversible internal fouling) [8]. Apart from these anecdotal  
80 applications, there still is a lack of analytical strategies for internal fouling investigation. Considering  
81 the well-known impact of fouling on the membrane charge, the investigation of surface and porous  
82 structures zeta potentials may bring useful information for external and internal fouling  
83 characterization.

84 Determining the zeta potential of a membrane provides insight into its surface electrical properties  
85 in a given physicochemical environment. It is therefore of great interest when investigating problems  
86 of practical relevance such as membrane fouling [9, 10] or ageing [11, 12].

87 The zeta potential of membranes can be inferred from electrokinetic techniques such as streaming  
88 potential and streaming current. From the experimental point of view, the easiest way to perform the  
89 electrokinetic characterization of porous membranes is to implement through-pore streaming  
90 potential measurements (also known as transversal mode) [13–15]. However, the multilayer  
91 structure of commercial membranes used in pressure-driven processes (micro-, ultra- and nano-  
92 filtration as well as reverse osmosis) makes the determination of the skin layer properties quite tricky  
93 [16–18]. In order to overcome the difficulty inherent in the analysis of through-pore measurements,  
94 an alternative measuring method, known as tangential mode and based on the application of the  
95 pressure gradient along the membrane skin layer (and not through the membrane thickness) has  
96 been proposed and has become the most widely used technique in membrane science [19–22].

97 When considering tangential electrokinetic measurements, it has been argued that streaming current  
98 should be preferred over streaming potential since this latter is likely to be impacted by the extra  
99 contribution of the underlying support layer(s) to the overall electrical conductance [23, 24].  
100 Tangential streaming current has been shown to be a reliable technique to highlight the presence of  
101 thin coating layers onto the surface of some commercial nanofiltration and reverse osmosis  
102 membranes [25]. However, complications have been pointed out when tangential streaming current  
103 measurement is applied to porous materials like micro- and ultra-filtration membranes due to the  
104 occurrence of a non-negligible streaming current through the membrane porosity[26–28]. This

105 parasitic contribution to the experimental signal, hereinafter referred as electrokinetic leakage, can  
106 be taken into account by applying a protocol consisting in a series of measurements performed by  
107 changing the distance between the two membranes samples required for tangential measurements  
108 [26–28]. Such a procedure has been successfully applied by Szymczyk *et al.* (2013) to give evidence  
109 for the presence of 4-benzyltriphenylphosphonium groups within pores of an ultrafiltration  
110 membrane after its functionalization [28].

111 In this work, we show for the first time that the electrokinetic leakage phenomenon can be used to  
112 gain insight into membrane fouling. Notably, we show that electrokinetic leakage can be used as a  
113 probe to highlight the occurrence of internal fouling within the porous structure of micro- and ultra-  
114 filtration membranes. The methodology followed in this work also enables distinction between  
115 fouling onto the membrane surface and within the membrane pores.

## 116 **2 Theoretical background**

117 Tangential streaming current measurement consists in applying a pressure gradient along a channel  
118 formed by two identical membrane samples facing each other and immersed in an electrolyte  
119 solution. While the pressure gradient is applied along the membrane skin layers, the solution is forced  
120 to move tangentially to the charged surfaces, pulling the excess of mobile ions within the electrical  
121 double layers towards the low-pressure side. It results in an electrical current, known as the  
122 streaming current ( $I_s$ ), flowing between the membrane surfaces.

123 The standard theory implicitly assumes that the channel through which tangential streaming current  
124 is measured has impermeable walls. If this condition is fulfilled and the distance between the surfaces  
125 of the two membrane samples ( $h_{ch}$ ) is much larger than the Debye length of the measuring solution,  
126 the zeta potential ( $\zeta$ ) can be inferred from the streaming current by means of the well-known  
127 Smoluchowski equation:

$$128 \quad I_s = - \frac{Wh_{ch}\epsilon_0\epsilon_r\Delta P}{\eta L} \zeta \quad (1)$$

129 where  $W$  and  $L$  are the channel width and length, respectively,  $\epsilon_0$  the vacuum permittivity ( $8.854 \times 10^{-12}$   
130  $\text{F m}^{-1}$ ),  $\epsilon_r$  and  $\eta$  the dielectric constant and the dynamic viscosity of the electrolyte solution,  
131 respectively, and  $\Delta P$  is the pressure difference applied between the channel ends.

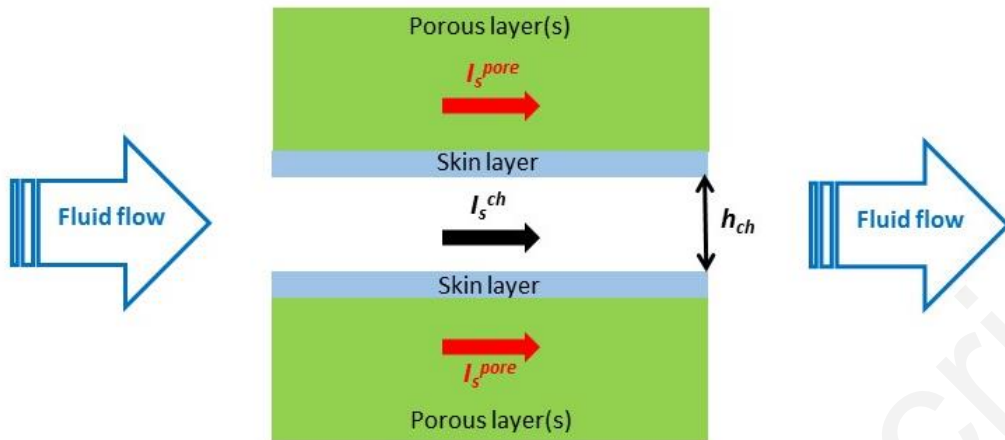
132 Although Eq. (1) is reliable for dense materials, it may break down when applied to ion-permeable  
 133 materials such as micro- and ultra-filtration membranes, especially if the electrokinetic cell has been  
 134 designed in such a way that it leaves the membrane support layer(s) exposed to the hydrodynamic  
 135 flow during streaming current measurements [26, 28]. The reason is that a non-negligible streaming  
 136 current is likely to flow through the membrane porous structure, which is filled with the measuring  
 137 solution. This additional streaming current that we shall refer to as “electrokinetic leakage” is not  
 138 accounted for in Eq. (1), which implicitly assumes that the experimental streaming current only flows  
 139 along the membrane surfaces. For a given membrane, the occurrence of electrokinetic leakage can be  
 140 easily confirmed or invalidated by measuring the streaming current for (at least) two different values  
 141 of  $h_{ch}$ . Indeed, in the case of electrokinetic leakage, the (apparent) zeta potential that would be  
 142 obtained by means of Eq. (1) would become dependent upon the distance between the membrane  
 143 samples.

144 Eq. (2) has been proposed by Yaroshchuk and Luxbacher [26] to account for the contribution of  
 145 electrokinetic leakage (see schematic description given in Fig. 1):

$$146 \quad I_s^{tot} = I_s^{ch} + 2I_s^{pore} = - \left( \frac{Wh_{ch}\epsilon_0\epsilon_r\Delta P}{\eta L} \zeta_{surf} + \frac{2Wh_{mb}^{eff}\epsilon_0\epsilon_r\Delta P}{\eta L} \zeta_{pore} \right) \quad (2)$$

147 Where  $I_s^{tot}$ ,  $I_s^{ch}$  and  $I_s^{pore}$  are the total streaming current (i.e. the current measured experimentally),  
 148 the streaming current flowing through the channel (i.e. between the membrane surfaces) and the  
 149 electrokinetic leakage occurring within a single membrane, respectively,  $\zeta_{surf}$  and  $\zeta_{pore}$  are the zeta  
 150 potentials of the membrane surface and inside the membrane porous body, respectively, and  $h_{mb}^{eff}$  the  
 151 effective height where the electrokinetic leakage takes place in a single membrane (it includes the  
 152 membrane thickness, porosity and tortuosity).

153



154  
 155 **Figure 1: Schematic representation of the streaming current distribution when tangential**  
 156 **electrokinetic measurements are carried out with porous membranes. The streaming current does not**  
 157 **flow only between the membrane surfaces but also through the membranes porous body. The**  
 158 **experimental streaming current is then equal to  $I_s^{ch} + 2I_s^{pore}$ .**

159  
 160 It is worth mentioning that, unlike Eqs. (1) and (2), the softwares associated with the current  
 161 commercial electrokinetic analyzers use an arbitrary convention stating that the measured streaming  
 162 current and the zeta potential(s) are of the same sign. This convention will be used in the rest of this  
 163 manuscript.

164 According to Eq. (2), the correct value of the zeta potential of the membrane surface can be  
 165 determined by carrying out a series of streaming current measurements with various channel heights  
 166 ( $h_{ch}$ ). Indeed,  $\zeta_{surf}$  can be obtained from the slope of  $I_s^{tot}$  vs.  $h_{ch}$  while the total electrokinetic leakage  
 167 ( $2I_s^{pore}$ ) is given by the y-intercept (the streaming current that would be measured if the two  
 168 membrane surfaces were brought into contact, i.e.  $h_{ch} = 0$ ). Eq. (2) also shows that the zeta potential  
 169 inside the membrane porosity can be determined from the electrokinetic leakage provided if  $h_{mb}^{eff}$  is  
 170 known, which can be achieved by means of additional electric conductance measurements for various  
 171  $h_{ch}$ . Indeed, considering the system described in Fig. 1 the following expression for the cell  
 172 conductance ( $G_{cell}$ ) can be derived [26,28]:



173 
$$G_{cell} = \frac{W}{L} (h_{ch} \lambda_0 + 2h_{mb}^{eff} \lambda_{mb}) \quad (3)$$

174 Where  $\lambda_0$  and  $\lambda_{mb}$  are the electric conductivities of the measuring solution in the channel between the  
175 membrane surfaces and inside the membrane pores, respectively.

176

## 177 **3 Materials and methods**

### 178 **3.1 Membranes**

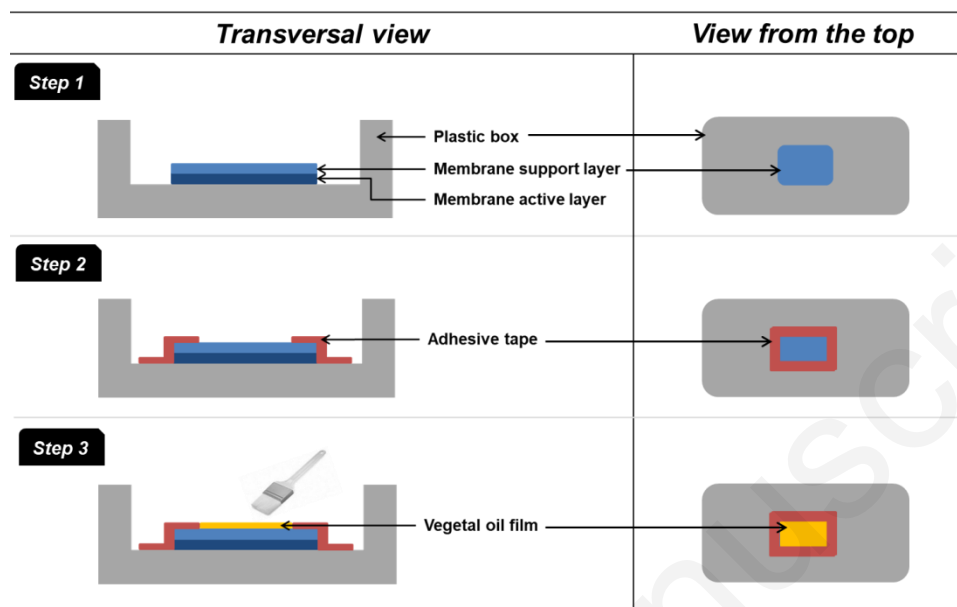
179 Two commercial flat-sheet membranes were used in this work: a microfiltration (MF)  
180 polyethersulfone (PES) membrane with an average pore diameter of 0.1  $\mu\text{m}$  (Koch, USA) and an  
181 ultrafiltration (UF) polyacrylonitrile (PAN) membrane with a molecular weight cut-off (MWCO) of  
182 500kDa (Orelis, France). The following protocol was followed to remove membrane preservatives  
183 prior use: sonication first in a 50 v/v % water-ethanol mixture for 10 min followed by sonication in  
184 deionized water (DI) water for 10 min (this last step was repeated twice).

185

### 186 **3.2 Membrane fouling**

#### 187 **3.2.1 Lab-scale fouling experiments**

188 “Upside down” fouling experiments were performed in order to provide the proof of concept of the  
189 methodology we propose in this work, which is based on the variation of the electrokinetic leakage  
190 occurring in the porous structures of membranes as a result of internal fouling. The protocol is  
191 schematically described in Fig. 2. The membrane coupons were fixed at the bottom of a plastic box  
192 using adhesive tape, the membrane skin layer facing the bottom of the box (steps 1 and 2 in Fig. 2).  
193 Then, commercial sunflower oil was poured onto the surface of the membrane support and  
194 homogeneously spread with a paintbrush. Oil was then let diffuse through the membrane (from the  
195 support towards the skin layer) for 4 hours at room temperature (step 3). The remaining oil on the  
196 support surface was wiped with a tissue paper prior to membrane characterization.



198

199 **Figure 2: Experimental protocol for upside down fouling of membrane samples by vegetable oil.**

200

### 201 3.2.2 Pilot-scale fouling experiments

202 Pilot-scale fouling experiments were performed using a cross-flow filtration module (Rayflow X100,  
 203 Orelis-Novasep, France) with a filtration area close to 130 cm<sup>2</sup>. The feed cross-flow circulation was  
 204 ensured by a peristaltic pump (ref 520S IP31, Watson-Marlow, USA), the cross-flow velocity being  
 205 around 0.4 m.s<sup>-1</sup>. Permeation through the membrane was ensured by applying a constant  
 206 transmembrane pressure difference (TMP) adjusted with a back-pressure valve. Two pressure  
 207 sensors placed at the module inlet and outlet on the retentate side allowed TMP monitoring and  
 208 adjustment [29]. Before measurements, the membranes were first cleaned by successive filtrations  
 209 of alkali solutions according to the following order: mixture of 0.1 g.L<sup>-1</sup> NaOH and 0.02 g.L<sup>-1</sup> NaClO  
 210 solution at 30°C (pH ~ 10-11), followed by 2 g.L<sup>-1</sup> commercial Ultrasil 110 solution (Ecolab, France)  
 211 at 45°C. Between each cleaning step, membranes were carefully rinsed in order to remove the  
 212 remaining chemicals. To this end, successive filtrations of DI water (30°C) without permeate and  
 213 retentate recirculation and under constant pressure of 0.2 bar were performed until the permeate  
 214 reached the pH of DI water. At the end of the cleaning step, membranes were compacted by DI water

215 filtration at constant TMP (2 bar) and constant temperature ( $30\pm 1^\circ\text{C}$ ) until a steady state water flux  
216 was reached ( $\pm 5\%$ ). The evolution of the permeate flux during the filtration process was measured  
217 by collecting the permeate in a beaker placed on an electronic scale (model XL1200C, Precisa,  
218 Switzerland).

219 After membrane compaction, DI water filtration pressure-stepping experiments (5 pressure steps  
220 from 0.2 to 1.0 bar) were carried out in order to determine the  $30^\circ\text{C}$  water permeability of each  
221 coupon.

222 Fouling experiments were further conducted using a 2% oil-in-water emulsion as the foulant feed (~  
223 2 liters). It was used by some of us in a previous study as a model system representative of the  
224 supernatant of a concentrated pretreated culture of *Parachlorella kessleri* microalgae, after bead  
225 milling and separation of the cell fragments by centrifugation [29,30]. The aqueous phase had a pH of  
226 7.4 and a conductivity of  $790\ \mu\text{S cm}^{-1}$  and the lipid phase consisted in a mixture of vegetable oils  
227 containing 70 wt% of neutral lipids and 30 wt% of polar lipids [29]. Fouling experiments were carried  
228 out at two TMP (0.2 and 1.0 bar) so as to get different levels of fouling. Experiments were stopped  
229 when a volume reduction ratio of 2 was achieved and the membranes were further rinsed by two  
230 successive DI water filtrations under a TMP of 0.2 bar for 15 minutes in order to remove the reversible  
231 part of fouling.

232

### 233 **3.3 Membrane characterization: Tangential streaming current**

234 Electrokinetic measurements were performed with a Surpass electrokinetic analyzer (Anton Paar  
235 GmbH) equipped with an adjustable-gap cell requiring two membrane samples (each one  $2 \times 1\ \text{cm}$ )  
236 [28]. For all experiments, the membranes were positioned in the adjustable-gap cell with their skin  
237 layers facing each other (as shown schematically in Fig. 1). Experiments were performed at  $T=22\pm 2$   
238  $^\circ\text{C}$  with 500 mL of a 0.001 M KCl solution, the pH of which was adjusted in the range 4.50 – 5.00 with  
239 a 0.05 M HCl solution and kept constant within  $\pm 0.05$  throughout the course of the experiment. Prior  
240 to measurements, the solution was circulated through the channel for *ca.* 2 h to allow the sample  
241 equilibration. After equilibration, the streaming current was measured and recorded for increasing  
242 pressure differences ( $\Delta P$ ) up to 300 mbar. Measurements were repeated by progressively decreasing

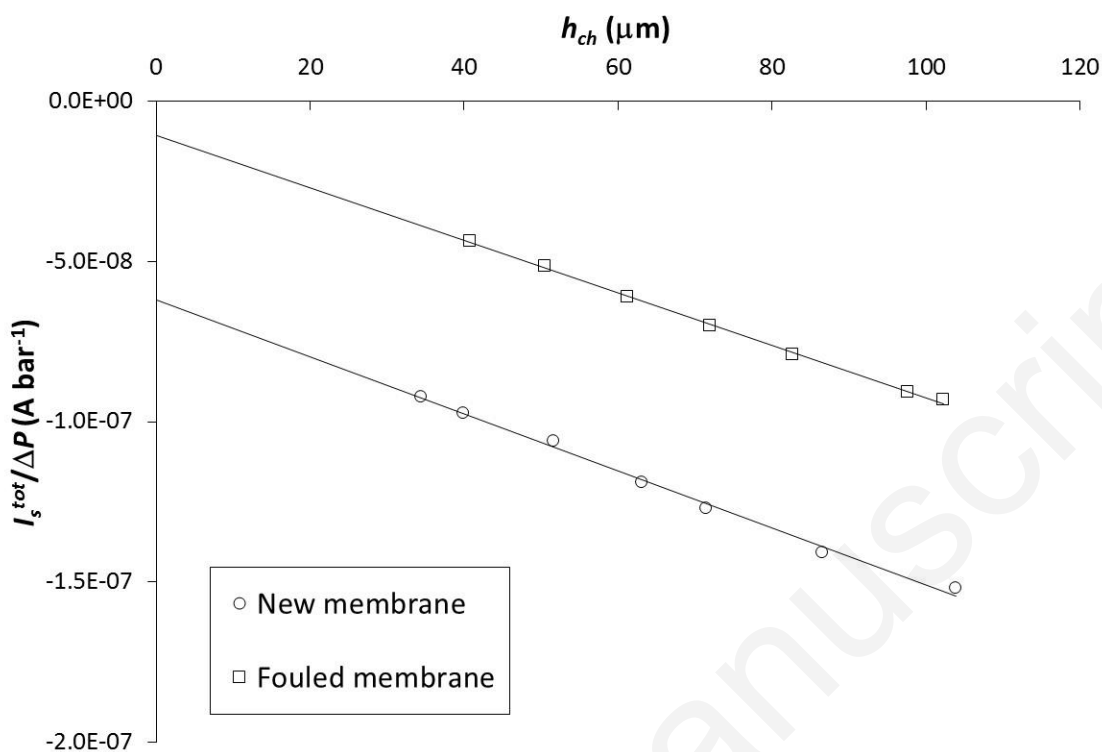
243 the distance between the membrane samples ( $h_{ch}$ ) from  $\sim 100 \mu\text{m}$  to  $\sim 40 \mu\text{m}$  by means of the  
244 micrometric screws of the adjustable-gap cell.

245 For some experiments, the electrical conductance of the adjustable-gap cell was also measured (for  
246 various values of  $h_{ch}$ ) in order to evaluate the effective height where the electrokinetic leakage takes  
247 place in the membrane ( $h_{mb}^{eff}$ ). For these additional measurements, a more concentrated KCl solution  
248 (0.1 M) was used in order to minimize the impact of the electrical double layers on the electrical  
249 conductivity inside the membrane pores [28].

## 250 **4 Results**

### 251 **4.1 Proof of concept**

252 In a preliminary work, the UF PAN membrane was first fouled according to the “upside down” fouling  
253 protocol described in section 3.2.1 and schematically shown in Fig. 2. Next, two membrane samples  
254 were positioned in the adjustable-gap cell of the electrokinetic analyzer with their skin layers facing  
255 each other (as shown schematically in Fig. 1). For all experiments, a linear variation was observed  
256 between the applied pressure and the resulting streaming current, making it possible to  
257 unambiguously define the streaming current coefficient as the slope of the  $I_s^{tot}$  vs  $\Delta P$  plots. Fig. 3 shows  
258 the variation of the streaming current coefficient as function of the distance between the membrane  
259 samples in the adjustable-gap cell ( $h_{ch}$ ). As expected from Eq. (2), a linear variation was observed for  
260 both new and fouled membrane samples.



261

262 **Figure 3: Streaming current coefficient of the pristine and fouled PAN membranes as function of the**  
 263 **distance between the samples; measuring solution: 0.001 M KCl solution at pH 4.50  $\pm$ 0.05.**

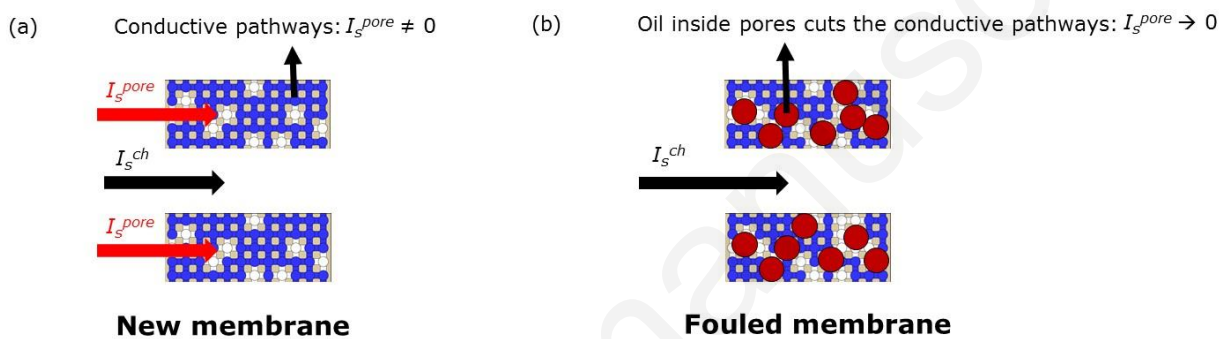
264

265 Extrapolating the streaming current coefficient of the pristine membrane down to  $h_{ch} = 0$  (i.e.  
 266 membrane surfaces in contact with no longer a channel in between) gives evidence of the  
 267 electrokinetic leakage phenomenon since the y-intercept differs from zero, as shown by Eq. (2). It was  
 268 found to be  $-6.2 \times 10^{-8}$  A.bar $^{-1}$ , which represents 40% of the experimental signal ( $I_s^{tot}/\Delta P$ ) measured by  
 269 setting the distance between the samples at 100  $\mu\text{m}$  and up to 65% of the experimental signal if  $h_{ch}$  is  
 270 set at 40  $\mu\text{m}$ . In the Supplementary Information of this manuscript, the reader can find additional  
 271 measurements obtained with a track-etched membrane (Fig. S1). As expected, no electrokinetic  
 272 leakage occurred since track-etched membranes have non-interconnected pores, thus preventing any  
 273 tangential flow through the porous structure.

274 Interestingly, an almost six-fold decrease in the magnitude of the electrokinetic leakage was obtained  
 275 after letting the support layer of the PAN membrane in contact with sunflower oil for four hours as  
 276 described in section 3.2.1. These results can be explained as follows. With the pristine membranes,

277 the electrokinetic leakage phenomenon can take place because the membrane pores are filled with  
 278 the electrolyte solution required for electrokinetic measurements, which creates ion-conductive  
 279 pathways through which a part of the streaming current,  $2I_s^{pore}$  in Eq. (2), can flow (see Fig. 4a); the  
 280 magnitude of the electrokinetic leakage would increase with the membrane thickness, porosity,  
 281 hydrophilicity and pore surface charge. Because of the hydrophobic character of oil, these conductive  
 282 pathways are progressively clogged as oil penetrates into the membrane pores (Fig. 4b), which results  
 283 in a weaker electrokinetic leakage as reported in Fig. 3.

284



285

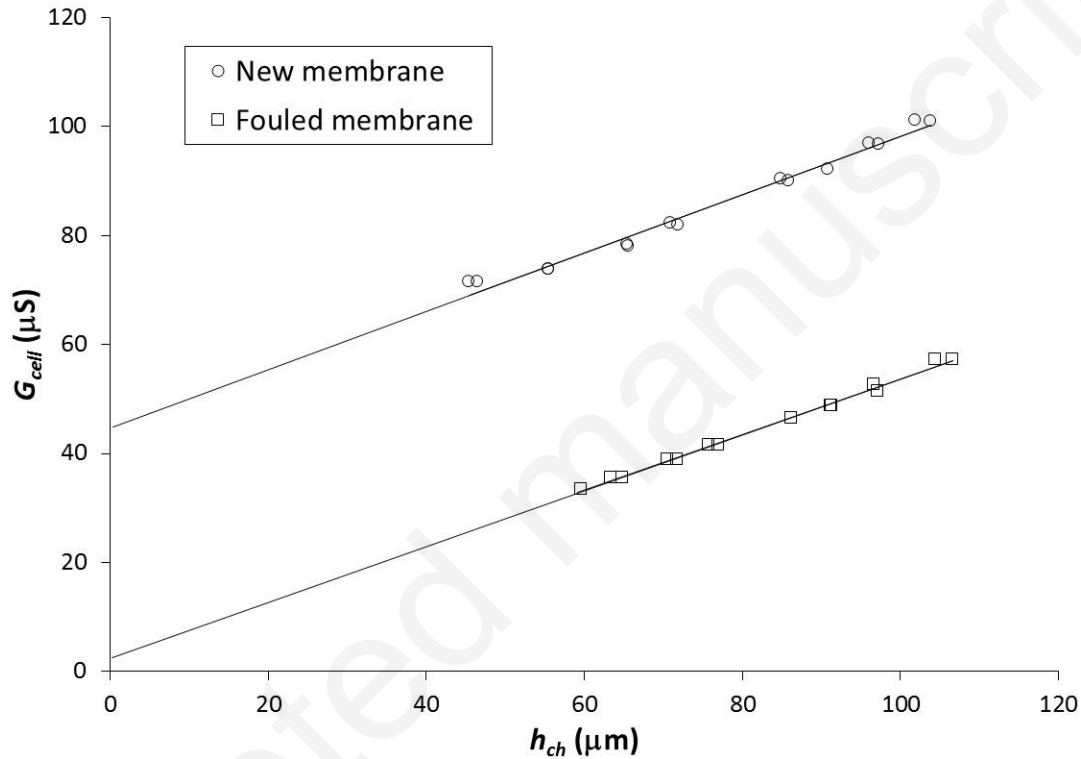
286 **Figure 4: Schematic representation of (a) open conductive pathways allowing electrokinetic leakage**  
 287 **through the membrane porosity and (b) clogged conductive pathways leading to electrokinetic**  
 288 **leakage disappearance.**

289 It is worth noting in Fig. 3 that the same slope was obtained in the electrokinetic response of both  
 290 PAN membrane coupons. According to Eq. (2), this results means that the zeta potential of the skin  
 291 layer surface ( $\zeta_{surf}$ ) was not impacted after membrane fouling. Otherwise stated, it means that oil  
 292 entered the membrane pores from the support layer side but did not cross the membrane skin layer.

293 These results show that the proposed methodology makes it possible to separate the contribution of  
 294 surface and internal fouling.

295 As stated in section 2, the effective height where the electrokinetic leakage takes place in a single  
 296 membrane ( $h_{mb}^{eff}$ ) can be estimated by means of additional electric conductance measurements. Fig.  
 297 5 shows the variation of the cell conductance ( $G_{cell}$ ) as function of  $h_{ch}$ . As predicted by Eq. (3), a linear  
 298 variation of  $G_{cell}$  with  $h_{ch}$  was obtained for both new and fouled PAN membranes. A concentrated

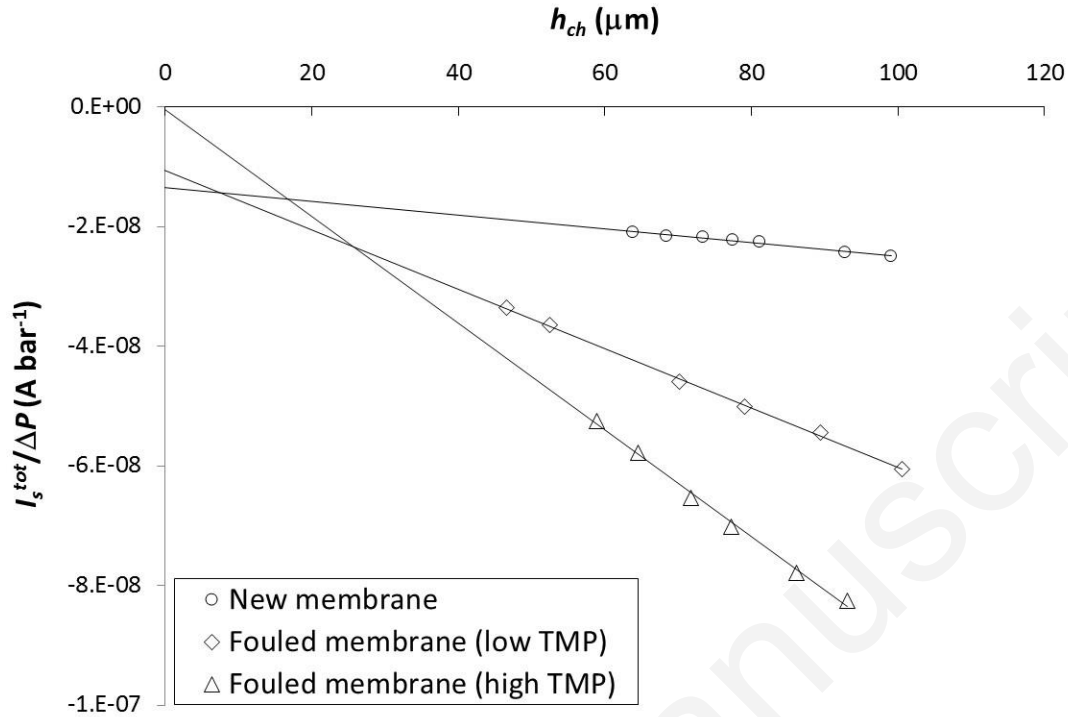
299 solution (0.1 M KCl) was used to minimize the impact of the electrical double layers on the electrical  
 300 conductivity inside the membrane pores and to reasonably assume  $\lambda_{mb} \approx \lambda_0$ [28]. The  $h_{mb}^{eff}$  values for  
 301 both membranes were inferred from the y-intercept according to Eq. (3). For the pristine membrane,  
 302  $h_{mb}^{eff}$  was found to be about 40  $\mu\text{m}$  while it fell down to 2  $\mu\text{m}$  for the fouled membrane, thus confirming  
 303 that the presence of oil inside the membrane pores cut the conduction pathways responsible for the  
 304 electrokinetic leakage phenomenon.



305  
 306 **Figure 5: Cell conductance measured with the PAN membranes as a function of the distance between**  
 307 **the samples; measuring solution: 0.1 M KCl solution at pH 4.50  $\pm$  0.05.**

308  
 309 **4.2 Application to oil-in-water emulsion filtration**

310 Pilot-scale filtration experiments were then conducted on the 2% oil-in-water emulsion using MF PES  
 311 and UF PAN membranes. Fig. 6 shows the variation of the streaming current coefficient of the MF PES  
 312 membrane as function of the distance between the membrane samples in the adjustable-gap cell ( $h_{ch}$ ).



313

314 **Figure 6: Streaming current coefficient of the pristine and fouled PES membranes as function of the**  
 315 **distance between the samples; measuring solution: 0.001 M KCl solution at pH 5.00 ±0.05.**

316

317 Different electrokinetic responses were obtained for the pristine membrane and the membranes  
 318 fouled by filtering the 2% oil-in-water emulsion described in section 3.2.2.

319 The slope of the electrokinetic response was found to increase with fouling, being even steeper after  
 320 filtration at high TMP (1 bar). Since the slope of the  $I_s^{tot}/\Delta P$  vs  $h_{ch}$  plot is proportional to  $\zeta_{surf}$  (see Eq.  
 321 (2)), it means that (i) the zeta potential of the foulant was higher than that of the bare membrane  
 322 surface and (ii) the coverage of the PES membrane surface by the oil droplets (surface fouling)  
 323 increased by applying a higher TMP. The values of  $\zeta_{surf}$  can be straightforwardly deduced from Eq.  
 324 (2) and are collected in Table 1.

325 A slight decrease in the electrokinetic leakage is observed after filtration of the emulsion at low TMP  
 326 (0.2 bar). On the other hand, the electrokinetic leakage almost vanished after filtration at 1 bar (it was  
 327 divided by a factor of 34 compared with the pristine membrane). These results indicate that a higher



328 TMP favored the penetration of the oil droplets into the porous structure of the 0.1  $\mu\text{m}$  PES  
329 membrane.

330 **Table 1: Surface zeta potentials ( $\zeta_{surf}$ ) of the various membranes inferred from Eq. (2).**

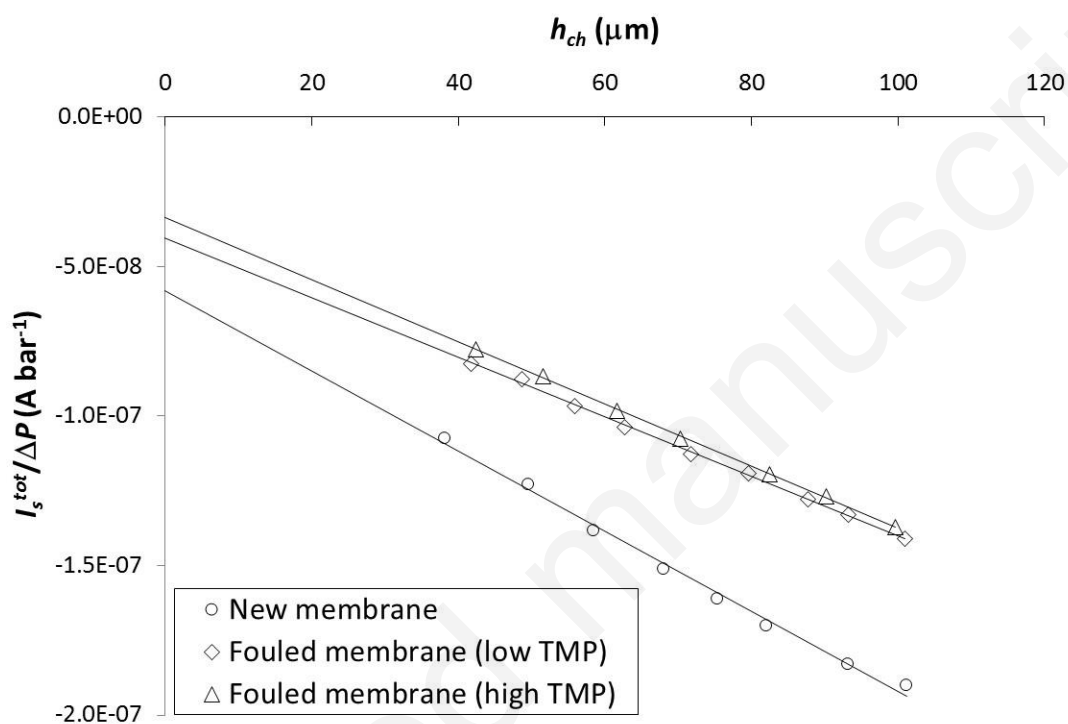
Membrane	Membrane surface zeta potential (mV)		
	New	Fouled (low TMP)	Fouled (high TMP)
PES 0.1 $\mu\text{m}$	$-2.9 \pm 0.9$	$-13.0 \pm 0.6$	$-23.1 \pm 1.4$
PAN 500 kDa	$-35.1 \pm 1.0$	$-26.1 \pm 2.9$	$-26.8 \pm 2.3$

331  
332 It can be noted that the standard Smoluchowski equation (Eq. (1)) could be used with confidence to  
333 compute the surface zeta potential of the PES membrane fouled at high TMP since the electrokinetic  
334 leakage phenomenon was suppressed under these operating conditions: the surface zeta potential  
335 determined from Eq. (1) with  $h_{ch} = 100 \mu\text{m}$  and from Eq. (2) are  $-23.2 \text{ mV}$  and  $-23.1 \text{ mV}$ , respectively.  
336 However it would overestimate the surface zeta potential of the new PES membrane and the  
337 membrane fouled at low TMP by 117 % and 21 %, respectively (still considering data for  $h_{ch} = 100$   
338  $\mu\text{m}$  in Eq. (1)).

339 A different behavior was observed with the 500 kDa UF PAN membrane. Indeed, as shown in Fig. 7,  
340 the impact of the TMP on both the external and internal fouling was marginal since the electrokinetic  
341 response (slope and y-intercept) of membranes fouled at low and high TMP were found to be very  
342 close.

343 The surface of the PAN membrane was found to be more negatively charged than the PES membrane  
344 (see Table 1). The slope of the electrokinetic response (and thus the surface zeta potential) was found  
345 to slightly decrease after fouling, unlike what was observed with the PES membrane for which a strong  
346 increase was reported (Fig. 6). It means that the foulant species from the emulsion were less  
347 negatively charged than the pristine PAN membrane. The surface zeta potentials of both the new and  
348 fouled PAN membranes are given in Table 1.

349 The PAN membrane appeared to be less prone to internal fouling by the oil-in-water emulsion than  
 350 the PES membrane since the electrokinetic leakage occurring in the PAN and PES membranes  
 351 decreased by a factor of 1.7 and 34, respectively, after fouling at high TMP. It results from to the more  
 352 open porous structure of the MF PES membrane compared with the UF PAN membrane along with  
 353 the more hydrophobic character of PES [10, 11] favoring interactions with the oil droplets.



354  
 355 **Figure 7: Streaming current coefficient of the pristine and fouled PAN membranes as function of the**  
 356 **distance between the samples; measuring solution: 0.001 M KCl solution at pH 5.00 ±0.05.**

357  
 358 Interestingly,  $\zeta_{surf}$  for the fouled PAN membranes and the PES membrane fouled at high TMP were  
 359 found to be very close: -26.1, -26.8 and -23.1 mV for the PAN membrane fouled at low TMP, the PAN  
 360 membrane fouled at high TMP and the PES membrane fouled at high TMP, respectively. Since the bare  
 361 PAN and PES membranes had substantially different  $\zeta_{surf}$  (-35.1 and -2.9 mV, respectively), this  
 362 finding suggests that (i) the surface of both membranes was fully covered by foulant species from the  
 363 emulsion and (ii) these foulant species develop an electrokinetic charge density in the

364 physicochemical environment considered in this study (0.001 M KCl at pH 5), which is characterized  
365 by a zeta potential in the range from  $\sim -20$  mV to  $\sim -30$  mV.

366 Additional electrophoretic light scattering measurements were performed with the emulsion diluted  
367 1000 times in a 0.001 M KCl solution at pH 5. The zeta potential of the emulsion droplets was found  
368 to be  $-25.7 \pm 1.1$  mV, which confirms that the PES membrane surface was fully covered after filtration  
369 at high TMP while that of the PAN membrane was covered even at low TMP.

370

## 371 **5 Conclusion**

372 The electrokinetic leakage, which is the part of the streaming current that flows through the porosity  
373 of a membrane when tangential electrokinetic measurements are carried out, is a parasitic  
374 phenomenon that makes more tedious the accurate determination of the surface zeta potential.  
375 Nonetheless, since this parasitic contribution is sensitive to local changes inside the membrane  
376 porosity it is likely to provide useful information when investigating problems of practical relevance.  
377 In this study, we showed for the first time that electrokinetic leakage can serve as a probe for  
378 detecting internal fouling in MF and UF membranes, taking lipidic fouling as example. Notably, it was  
379 used to highlight the substantial impact of the TMP on the internal fouling of a MF PES membrane  
380 caused by filtration of an oil-in-water emulsion. On the other hand, almost no impact of the TMP was  
381 noticed in the case of a UF PAN membrane with narrower pores. Moreover, the experimental protocol  
382 implemented here to quantify the electrokinetic leakage phenomenon allowed to distinguish between  
383 the contribution of internal fouling and that of external (surface) fouling.

384 This paper offers new prospects for tangential electrokinetic measurements by providing an  
385 innovative methodology for the characterization of internal fouling. These findings could find useful  
386 application in MF and UF processes for which internal fouling is often responsible of severe  
387 productivity decrease and laborious cleaning procedures. Moreover, in view of the vast volumes of  
388 oily wastewater produced in various industrial fields (microalgae, food and beverages, oil and gas,  
389 metal processing, etc.), the characterization of membrane fouling during oil-in-water emulsions  
390 treatment is a key challenge for the development of cost-effective and eco-friendly processing  
391 involving membranes filtration.

392 Interestingly, the methodology proposed in this work is not restricted to membrane fouling studies  
393 and it could be applied straightforwardly to other topics of membrane science including membrane  
394 functionalization and membrane degradation (physical or chemical ageing). A limitation of this  
395 method is that it requires varying the spacing between the membranes, which makes it difficult to  
396 apply it on-line with existing membrane modules.

## 397 **6 Acknowledgements**

398 Cédric Brossard (Malvern Panalytical) is gratefully acknowledged for electrophoretic light scattering  
399 measurements. This work was supported by GIS Europôle Mer, France (Sciences et Ingénieries  
400 Marines, 2016-2017), Université de Bretagne Loire, ADEME and the Challenge Food For  
401 Tomorrow/Cap Aliment, Pays de la Loire, France.

## 402 **7 Bibliography**

- 403 [1] G. Rudolph, T. Virtanen, M. Ferrando, C. Güell, F. Lipnizki, M. Kallioinen, A review of in situ real-  
404 time monitoring techniques for membrane fouling in the biotechnology, biorefinery and food  
405 sectors, *J. Membr. Sci.* 588 (2019) 117221. doi:10.1016/j.memsci.2019.117221.
- 406 [2] D.J. Johnson, D.L. Oatley-Radcliffe, N. Hilal, State of the art review on membrane surface  
407 characterisation: Visualisation, verification and quantification of membrane properties, *Rev.*  
408 *Res. Dev. Desalination.* 434 (2018) 12–36. doi:10.1016/j.desal.2017.03.023.
- 409 [3] D. Delaunay, M. Rabiller-Baudry, J.M. Gozálvarez-Zafrilla, B. Balannec, M. Frappart, L. Paugam,  
410 Mapping of protein fouling by FTIR-ATR as experimental tool to study membrane fouling and  
411 fluid velocity profile in various geometries and validation by CFD simulation, *Euromembrane*  
412 2006. 47 (2008) 1106–1117. doi:10.1016/j.cep.2007.12.008.
- 413 [4] M. Rabiller-Baudry, L. Bégoïn, D. Delaunay, L. Paugam, B. Chaufer, A dual approach of membrane  
414 cleaning based on physico-chemistry and hydrodynamics: Application to PES membrane of  
415 dairy industry, *10th Fr. Congr. Chem. Eng.* 47 (2008) 267–275. doi:10.1016/j.cep.2007.01.026.
- 416 [5] M.J. Luján-Facundo, J.A. Mendoza-Roca, B. Cuartas-Urbe, S. Álvarez-Blanco, Evaluation of  
417 cleaning efficiency of ultrafiltration membranes fouled by BSA using FTIR-ATR as a tool, *J. Food*  
418 *Eng.* 163 (2015) 1–8. doi:10.1016/j.jfoodeng.2015.04.015.
- 419 [6] A. Yeo, P. Yang, A.G. Fane, T. White, H.O. Moser, Non-invasive observation of external and internal  
420 deposition during membrane filtration by X-ray microimaging (XMI), *J. Membr. Sci.* 250 (2005)  
421 189–193. doi:10.1016/j.memsci.2004.10.035.

- 422 [7] T.A. Trinh, W. Li, Q. Han, X. Liu, A.G. Fane, J.W. Chew, Analyzing external and internal membrane  
423 fouling by oil emulsions via 3D optical coherence tomography, *J. Membr. Sci.* 548 (2018) 632–  
424 640. doi:10.1016/j.memsci.2017.10.043.
- 425 [8] M.C. Martí-Calatayud, S. Schneider, M. Wessling, On the rejection and reversibility of fouling in  
426 ultrafiltration as assessed by hydraulic impedance spectroscopy, *J. Membr. Sci.* 564 (2018) 532–  
427 542. doi:10.1016/j.memsci.2018.07.021.
- 428 [9] J. Benavente, G. Jonsson, Effect of adsorbed protein on the hydraulic permeability, membrane  
429 and streaming potential values measured across a microporous membrane, *Colloids Surf.*  
430 *Physicochem. Eng. Asp.* 138 (1998) 255–264. doi:10.1016/S0927-7757(97)00119-2.
- 431 [10] L. Marbelia, M. Mulier, D. Vandamme, K. Muylaert, A. Szymczyk, I.F.J. Vankelecom,  
432 Polyacrylonitrile membranes for microalgae filtration: Influence of porosity, surface charge and  
433 microalgae species on membrane fouling, *Algal Res.* 19 (2016) 128–137.  
434 doi:10.1016/j.algal.2016.08.004.
- 435 [11] Y. Hanafi, A. Szymczyk, M. Rabiller-Baudry, K. Baddari, Degradation of Poly(Ether  
436 Sulfone)/Polyvinylpyrrolidone Membranes by Sodium Hypochlorite: Insight from Advanced  
437 Electrokinetic Characterizations, *Environ. Sci. Technol.* 48 (2014) 13419–13426.  
438 doi:10.1021/es5027882.
- 439 [12] Y. Hanafi, P. Loulergue, S. Ababou-Girard, C. Meriadec, M. Rabiller-Baudry, K. Baddari, A.  
440 Szymczyk, Electrokinetic analysis of PES/PVP membranes aged by sodium hypochlorite  
441 solutions at different pH, *J. Membr. Sci.* 501 (2016) 24–32. doi:10.1016/j.memsci.2015.11.041.
- 442 [13] M. Nyström, A. Pihlajamäki, N. Ehsani, Characterization of ultrafiltration membranes by  
443 simultaneous streaming potential and flux measurements, *J. Membr. Sci.* 87 (1994) 245–256.  
444 doi:10.1016/0376-7388(94)87031-4.
- 445 [14] I.H. Huisman, P. Prádanos, A. Hernández, Electrokinetic characterisation of ultrafiltration  
446 membranes by streaming potential, electroviscous effect, and salt retention, *J. Membr. Sci.* 178  
447 (2000) 55–64. doi:10.1016/S0376-7388(00)00479-8.
- 448 [15] M. Sbaï, P. Fievet, A. Szymczyk, B. Aoubiza, A. Vidonne, A. Foissy, Streaming potential,  
449 electroviscous effect, pore conductivity and membrane potential for the determination of the  
450 surface potential of a ceramic ultrafiltration membrane, *J. Membr. Sci.* 215 (2003) 1–9.  
451 doi:10.1016/S0376-7388(02)00553-7.
- 452 [16] A. Szymczyk, C. Labbez, P. Fievet, B. Aoubiza, C. Simon, Streaming potential through multilayer  
453 membranes, *AIChE J.* 47 (2001) 2349–2358. doi:10.1002/aic.690471019.
- 454 [17] A.E. Yaroshchuk, Y.P. Boiko, A.L. Makovetskiy, Filtration Potential across Membranes Containing  
455 Selective Layers, *Langmuir.* 18 (2002) 5154–5162. doi:10.1021/la025503s.
- 456 [18] P. Fievet, M. Sbaï, A. Szymczyk, Analysis of the pressure-induced potential arising across  
457 selective multilayer membranes, *J. Membr. Sci.* 264 (2005) 1–12.  
458 doi:10.1016/j.memsci.2005.04.011.

- 459 [19] D. Möckel, E. Staude, M. Dal-Cin, K. Darcovich, M. Guiver, Tangential flow streaming potential  
460 measurements: Hydrodynamic cell characterization and zeta potentials of carboxylated  
461 polysulfone membranes, *J. Membr. Sci.* 145 (1998) 211–222. doi:10.1016/S0376-  
462 7388(98)00077-5.
- 463 [20] P. Fievet, M. Sbaï, A. Szymczyk, C. Magnenet, C. Labbez, A. Vidonne, A New Tangential Streaming  
464 Potential Setup for the Electrokinetic Characterization of Tubular Membranes, *Sep. Sci. Technol.*  
465 39 (2004) 2931–2949. doi:10.1081/SS-200028652.
- 466 [21] V.T. Do, C.Y. Tang, M. Reinhard, J.O. Leckie, Degradation of Polyamide Nanofiltration and Reverse  
467 Osmosis Membranes by Hypochlorite, *Environ. Sci. Technol.* 46 (2012) 852–859.  
468 doi:10.1021/es203090y.
- 469 [22] S. Liu, C. Wu, X. Hou, J. She, S. Liu, X. Lu, H. Zhang, S. Gray, Understanding the chlorination  
470 mechanism and the chlorine-induced separation performance evolution of polypiperazine-  
471 amide nanofiltration membrane, *J. Membr. Sci.* 573 (2019) 36–45.  
472 doi:10.1016/j.memsci.2018.11.071.
- 473 [23] A. Yaroshchuk, V. Ribitsch, Role of Channel Wall Conductance in the Determination of  $\zeta$ -Potential  
474 from Electrokinetic Measurements, *Langmuir.* 18 (2002) 2036–2038. doi:10.1021/la015557m.
- 475 [24] P. Fievet, M. Sbaï, A. Szymczyk, A. Vidonne, Determining the  $\zeta$ -potential of plane membranes  
476 from tangential streaming potential measurements: effect of the membrane body conductance,  
477 *J. Membr. Sci.* 226 (2003) 227–236. doi:10.1016/j.memsci.2003.09.007.
- 478 [25] E. Idil Mouhoumed, A. Szymczyk, A. Schäfer, L. Paugam, Y.H. La, Physico-chemical  
479 characterization of polyamide NF/RO membranes: Insight from streaming current  
480 measurements, *J. Membr. Sci.* 461 (2014) 130–138. doi:10.1016/j.memsci.2014.03.025.
- 481 [26] A. Yaroshchuk, T. Luxbacher, Interpretation of Electrokinetic Measurements with Porous Films:  
482 Role of Electric Conductance and Streaming Current within Porous Structure, *Langmuir.* 26  
483 (2010) 10882–10889. doi:10.1021/la100777z.
- 484 [27] S. Déon, P. Fievet, C. Osman Doubad, Tangential streaming potential/current measurements for  
485 the characterization of composite membranes, *J. Membr. Sci.* 423–424 (2012) 413–421.  
486 doi:10.1016/j.memsci.2012.08.038.
- 487 [28] A. Szymczyk, Y.I. Dirir, M. Picot, I. Nicolas, F. Barrière, Advanced electrokinetic characterization  
488 of composite porous membranes, *J. Membr. Sci.* 429 (2013) 44–51.  
489 doi:10.1016/j.memsci.2012.11.076.
- 490 [29] L. Villafaña-López, E.C. Rivera, S. Liu, E. Couallier, M. Frappart, Shear-enhanced membrane  
491 filtration of model and real microalgae extracts for lipids recovery in biorefinery context,  
492 *Bioresour. Technol.* (2019) 121539. doi:10.1016/j.biortech.2019.121539.
- 493 [30] E. Clavijo Rivera, V. Montalescot, M. Viau, D. Drouin, P. Bourseau, M. Frappart, C. Monteux, E.  
494 Couallier, Mechanical cell disruption of *Parachlorella kessleri* microalgae: Impact on lipid  
495 fraction composition, *Bioresour. Technol.* 256 (2018) 77–85.  
496 doi:10.1016/j.biortech.2018.01.148.

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## Supplementary Information

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### Electrokinetic leakage as a tool to probe internal

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### fouling in MF and UF membranes

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Number of pages: 2

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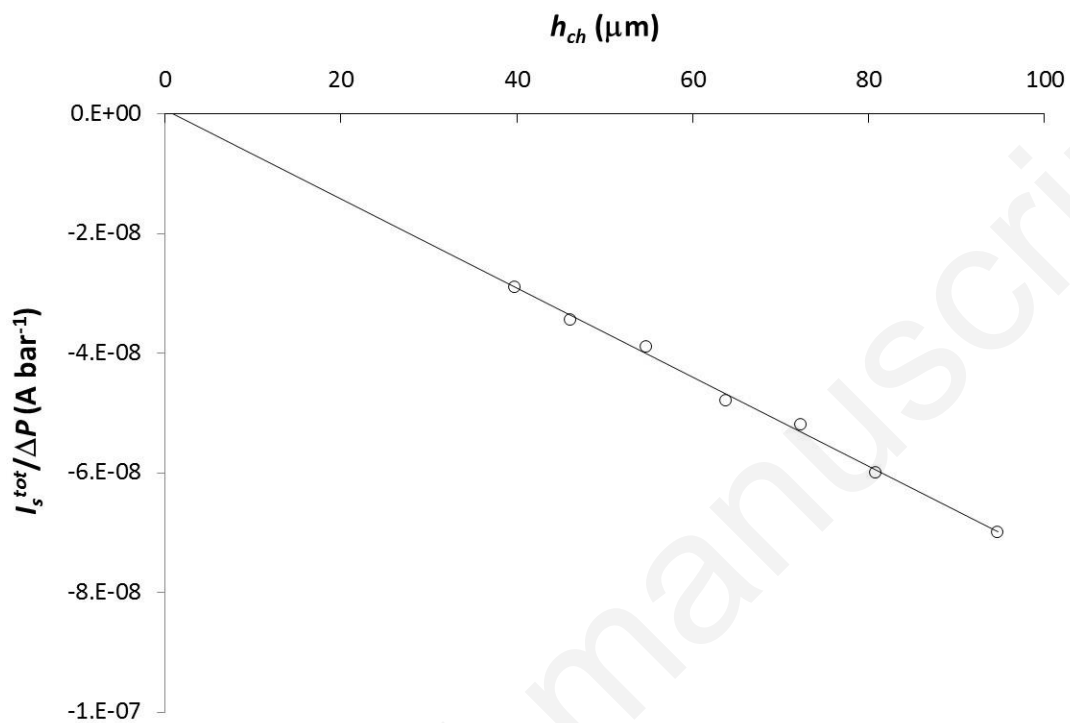
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539 **Figure S1: Streaming current coefficient of a track-etched membrane as function of the distance**  
540 **between the samples; measuring solution: 0.001 M KCl solution at pH 5.00  $\pm$  0.05.**

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