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Red pepper peptide coatings control *Staphylococcus epidermidis* adhesion and biofilm formation

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Medical devices (indwelling) have greatly improved healthcare. Nevertheless, infections related to the use of these apparatuses continue to be a major clinical concern. Biofilms form on surfaces after bacterial adhesion, and they function as bacterial reservoirs and as resistance and tolerance factors against antibiotics and the host immune response. Technological strategies to control biofilms and bacterial adhesion, such as the use of surface coatings, are being explored more frequently, and natural peptides may promote their development. In this study, we purified and identified antibiofilm peptides from *Capsicum baccatum* (red pepper) using chromatography-tandem mass spectrometry, MALDI-MS, MS/MS and bioinformatics. These peptides strongly controlled biofilm formation by *Staphylococcus epidermidis*, the most prevalent pathogen in device-related infections, without any antibiotic activity. Furthermore, natural peptide-coated surfaces displayed effective antiadhesive properties and showed no cytotoxic effects against different representative human cell lines. Finally, we determined the lead peptide predicted by Mascot and identified CSP37, which may be useful as a prime structure for the design of new antibiofilm agents. Together, these results shed light on natural *Capsicum* peptides as a possible antiadhesive coat to prevent medical device colonization.

Key words: biofilm, antibiofilm, antiadhesive, capsicum, peptide, coating

Introduction

Medical devices (indwelling) have made great contributions to health and quality of life evolution.^{1,2} However, infections related to the use of these medical devices and to widespread antibiotic misuse have led to a lethal health crisis with the increasing appearance of tolerant and resistant bacteria that do

not respond to conventional antibiotics.³⁻⁵ Associated with this phenomenon is the ability of microorganisms to form biofilms that confer evolutionary advantages that are phenotypically beneficial to increased survival and symbiotic relationships.⁶ A biofilm is a complex matrix composed of assembled extracellular polymeric substances (EPSs) enwrapping microorganisms. This structure allows bacterial surface fixation on catheters, implants, probes, prostheses or damaged tissues and protects bacteria against antibiotics and host immune defenses.⁷⁻¹¹

Biofilm formation is considered a significant virulence factor and is present in 80% of difficult-to-treat human infections, such as endocarditis, osteomyelitis, urethritis, and periodontitis, strongly impacting patient outcomes and health system costs.¹²⁻¹⁶

In this context, *Staphylococcus epidermidis* is an opportunistic pathogen that is highly associated with medical device-related infections.¹⁷⁻²² Current discussions suggest that this bacterium should be classified as pathogenic because disease-causing *S. epidermidis* strains represent a pathogenic subpopulation that has acquired genetic elements and related phenotypes that promote infection.²³ Furthermore, the ability to form biofilms on diverse devices or surfaces is the major virulence factor of *S. epidermidis*.²⁴ For instance, biofilm-forming *S. epidermidis* strains have been reported to include many strains that are resistant to ciprofloxacin and sulfamethoxazole, as well as to aminoglycosides.²⁵

Some technological strategies for the development of biomaterials that have been proposed include functionalized or coated antiadhesive surfaces and intrinsically antibacterial or nanostructured materials.²⁶⁻²⁸ However, among these

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Electronic Supplementary Information (ESI) available: (S1) Photos of *C. baccatum* pepper fruits, seeds and flowers; (S2) Graph with a set of the main bioactive fractions screened for antibiofilm and antibacterial activities; (S3) Scanning electron microscopy (SEM) images of the coatings. See DOI: xxxxxxxx/x0xx00000x

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strategies, the use of natural products to control bacterial adhesion and biofilm formation is rare.²⁹

Peppers from the genus *Capsicum* comprise more than 200 varieties, and their fruits are broadly diverse in relation to size, shape, flavor and sensory heat.³⁰ *Capsicum* species are native to the tropical and humid zones of Central and South America and include peppers of important economic value.³¹⁻³³ They are commonly consumed as spice or food as well as used as remedies in Indian, Native American and Chinese traditional medicine to treat skin rashes, dog/snake bites and wounds.³⁴ *Capsicum* peppers display several biological activities, such as anti-inflammatory, hypoglycemic and hypocholesterolemic, though its pharmacological potential and chemical composition have been underexplored.³⁵⁻⁴² Recently, we demonstrated that an aqueous extract of *C. baccatum* seeds decreased *S. epidermidis* biofilm formation by 80%, indicating that this red pepper is a potential source of antibiofilm and antiadhesive compounds.⁴³ In the present study, we purified and identified antibiofilm and antiadhesive peptides from *C. baccatum* using chromatography-tandem mass spectrometry, MALDI-MS/MS and bioinformatics tools. We demonstrated the ability of *Capsicum* storage peptides (CSP) to prevent biofilm formation by methicillin-resistant *S. epidermidis* without antibiotic mechanisms. CSP cytotoxic profiles were assessed *in vitro*. Surface CSP coatings were developed, and their effects were examined and characterized. These results may corroborate the development of new natural strategies in the war against medical device-related infections.

Material and Methods

Chemicals

All reagents, which were analytical or HPLC grade, were purchased from Sigma Chemical Company™. All solvents were purchased from Vetec™ AG (Rio de Janeiro, Brazil). The matrices 2,5-dihydroxybenzoic acid (DHB) and sinapinic acid (SA) were purchased from Bruker Daltonics. Synthetic peptides suitable for use in cell culture were purchased from China Peptides with >95% purity.

Plant material and extraction

C. baccatum var. *pendulum* (Willd.) Eshbaugh (Solanaceae) fruits were obtained from a cultivated area in Turuçú, Rio Grande do Sul (RS), Brazil (Figure S1, Supplementary Material). A voucher specimen (ICN 181469) was identified and deposited at the Herbarium of Universidade Federal of Rio Grande (UFRGS, Brazil). The seeds and extracts were obtained and prepared according to Gomes Von Borowski *et al.*⁴³ The most active extract was lyophilized to obtain the residual aqueous extract from seeds (RAqS).

Purification of residual aqueous extract from seeds (RAqS)

RAqS (700 mg) were solubilized in ultrapure water and subjected to column chromatography (3 cm diameter) using Sephadex™ LH-20 (40 g). Water was used as the eluent, and fourteen fractions of 10 ml each were collected (F1-F14). These

purified fractions were screened for antibiofilm and antibacterial activities against *S. epidermidis* at 4.0, 2.0, 1.0, 0.5 and 0.25 mg/mL. The selected active fraction, which presented 1.4% yield, was submitted to characterization by LC-DAD-ESI-MS and MALDI-MS/MS.

Bacterial strain and culture conditions

Staphylococcus epidermidis (ATCC 35984) was cultivated on Muller Hinton agar at 37°C overnight. The inoculum concentration was adjusted to an optical density at 600 nm equivalent to 10⁸ CFU/ml.

Sample preparation for microbiological assays

Samples were freshly dissolved in ultrapure water and filtered through a 0.2 µm pore size membrane before each assay. The red pepper active fraction was tested at 2.0 mg/ml and the synthetic peptide (CSP37) at 0.2, 0.4 and 0.9 mM.

Antibiofilm and antibacterial assays

To evaluate biofilm formation, an adapted protocol from Trentin *et al.*²⁹ was applied using 0.4% crystal violet staining and standard sterile 96-well polystyrene flat-bottom microtiter plates. Finally, the absorbance (570 nm) was measured (Spectramax™ M2e Multimode Microplate Reader, Molecular Devices, Sunnyvale, CA, USA). Bacterial growth was monitored by absorbance at 600 nm. Biofilm formation in the presence of the water control was considered to represent 100% biofilm formation.

Kinetics of bacterial growth

Bacterial growth kinetics were evaluated by monitoring the optical density (600 nm) at different timepoints. Samples were monitored after 0, 1, 3, 6, 12, and 24 h of incubation (37°C). The red pepper active fraction (2.0 mg/ml) was evaluated, and rifampicin (8 µg/ml) was used as a death control. The results were expressed as OD_{600nm} per incubation time.

Coatings preparation and characterization

Solutions of the red pepper active fraction were prepared using 70% methanol and filtered through a 0.2 µm pore size membrane. Permanox™ slides (30 x 25 mm) were coated once or three times with the active fraction solution at either 2.0 mg/ml or 10.0 mg/ml. Methanol was used to prepare nontreated control slides. In a Laurell model WS-650MZ-23NPP/LITE spin coater, 300 µl of the active fraction solution was deposited on each Permanox™ slide. It was accelerated for 5 s up to an angular velocity of 500 rpm and subsequently accelerated to 4000 rpm for 40 s. To remove the remaining solvent and to promote film annealing, slides were heat-treated at 40°C in a stove for 4 h. After the coating process, slides were sterilized using UV light for 20 min, cut to 10 x 25 mm and used in antibiofilm assays and scanning electron microscopy evaluation. Permanox™ samples were characterized before and after coating by water contact angle (WCA), which was measured using the sessile drop technique and Milli-Q water.

Drop images were captured and analyzed by a Theta Lite optical tensiometer (Attension). The reported WCAs are the means of five measurements performed on different areas of each sample surface.

Scanning electron microscopy (SEM)

Biofilms of *S. epidermidis* were cultured as described above. One group received a piece of noncoated Permanox™ slide (as a nontreated and no solvent control), while others received a piece of red pepper active fraction-coated slide (once or three times at 2.0 mg/ml or 10.0 mg/ml) or methanol-coated slide (as a solvent control). Noncoated controls were incubated with TSB broth containing the active fraction and bacteria, and coated samples were incubated with only TSB broth and bacteria (24 h at 37°C). Then, the slides were washed with sterile 0.9% NaCl and fixed with 2.5% glutaraldehyde (prepared in 100 mM cacodylate buffer, pH 7.2). Afterwards, they were washed with 100 mM cacodylate buffer (pH 7.2), dehydrated with increasing concentrations of acetone, dried with CO₂ as the critical point (Bal-Tec CPD 030 critical point dryer, Balzers, Liechtenstein) and placed on a mounting base. Finally, they were coated with carbon and examined under a scanning electron microscope (model JSM-6060, JEOL) at 10 kV.

Hemolysis test

Hemolysis assays were performed with human venous blood from healthy donors as previously described by Rocha *et al.*⁴⁴ All consents, procedures and documents were approved by the Ethics Committee of Universidade Federal do Rio Grande do Sul (UFRGS) (number 666.655/2014). The red pepper active fraction was tested at 2.0 mg/ml. A solution of 1% Triton™ was used to produce 100% hemolysis (reference, positive control), and phosphate buffered saline (PBS) was used as a negative control (no hemolysis). The test sample results were statistically compared with those of the positive and negative controls.

Lymphocyte cytotoxicity test and LDH release determination

Lymphocytes were obtained from heparin-treated human venous blood from healthy donors and isolated as previously described by Waetchter *et al.*⁴⁵ Briefly, the lymphocytes were separated by Histopaque™ gradient centrifugation, cultured at 100,000 cells/well in a 24-well plate in RPMI 1640 medium containing 10% fetal bovine serum (FBS) (Gibco BRL, Grand Island, NY), and treated with either the red pepper active fraction at 2 mg/ml, 1% Triton™ solution (cytotoxic control) or RPMI medium (nontoxic control) for 24 h, all in quintuplicate. Cell morphology was verified by optical microscopy, and viable cell counting was performed with a FACSVerser™ flow cytometer equipped with a 488 nm blue laser and a flow sensor (BD Biosciences, San Jose, CA, EUA). BD FacSuite™ software (BD Biosciences) was used for data analysis. All consents, procedures and documents were approved by the Ethics Committee of UFRGS (number 666.655/2014). To monitor lactate dehydrogenase (LDH) release due to lysis, the supernatants of the treated cells were assessed for LDH activity using the LDH enzymatic assay kit available from Laborclin™

(São José do Rio Preto, São Paulo, Brazil). The results are expressed as percentages, and the Triton treatment was considered to result in 100% LDH release.

High-throughput multiparameter cell image analysis

High Content Screening (HCS) and High Content Analysis (HCA) were performed with a robotic platform (ImpACcell, BIOSIT, Université de Rennes 1) using 7 representative human cell lines: HuH7 (liver), CaCo-2 (colon), MDA (breast), HCT116 (colon), PC3 (prostate), NCI-H727 (lung) and MCF7 (skin). The number of normal cells is presented as the residual cell percentage (%) compared to the DMSO control average, and the synthetic peptide (CSP37) was tested at 25 µM.^{46,47} The platform is equipped with an Olympus upright microscope (Spot NB camera and Simple PCI software, Compix), a Zeiss AxioImager M1 upright microscope (Marzhauser, Zeiss NB camera and AxioVision software), an ArrayScan VTI high-content imager (Thermo/Cellomics), Hamilton STARlet and Nimbus liquid handling systems, and a Scienion liquid spotter.

Liquid chromatography-tandem mass spectrometry analysis

Analyses were performed on a Shimadzu UFLC-20AD system coupled to a diode array detector (DAD) and an ESI-ion trap mass spectrometer (LC-DAD-ESI-MS) (Amazon SL, Bruker Daltonics). A Luna C18 column (5 µm, 4,6 mm x 250 mm, Phenomenex) was used for chromatography analyses. The mobile phase consisted of acetonitrile (solvent B) and water (solvent A), each containing 0.1% formic acid. The flow rate was 0.6 ml/min, and the injection volume was 20 µl. The elution profile was the following: 2% B (0 to 3 min), 2 to 20% B (3 to 22 min), 20 to 30% B (22 to 30 min), 30 to 100% B (30 to 40 min), and 100% B (40 to 45 min). The LC flow was split 3:7. The MS conditions were the following: the capillary voltage was 3500 V, nebulizer gas (nitrogen) pressure was set to 50 psi, dry gas flow was 9 l/min, and dry temperature was 300°C. MS data were acquired in negative and positive ion modes. The samples were analyzed over a mass range (*m/z*) of 0-1200.

MALDI-MS/MS analyses

UltrafleXtreme MALDI-TOF/TOF equipment (BrukerDaltonics, Bremen, Germany) was used for the analyses. For analyses of the fractions, the matrices 2,5-dihydroxybenzoic acid (DHB) and sinapinic acid (SA) were evaluated, and better results were obtained with DHB, which was used for all protein analyses. MS analysis: the samples were analyzed by reflector (700 to 5000 Da and 4 to 20 kDa) and linear modes (4-20, 20-50 and 50-100 kDa). The samples were dissolved in acetonitrile (ACN):H₂O (20:80, v/v) with 0.1% trifluoroacetic acid and the DHB matrix (20 mg/ml) in ACN:H₂O (30:70, v/v) with 0.1% trifluoroacetic acid. The sample solution and the matrix were mixed in equal amounts and spotted onto a ground stainless steel MALDI target (1 µl). The experimental conditions for MS analyses were as follows: pulsed ion extraction of 360 ns, laser frequency of 1000 Hz, and positive ion mode. In all, 5000 shots were averaged to record each mass spectrum. For the MS/MS analyses, the red pepper active fraction was resolved by

electrophoresis gel and the obtained spots (Sp1, Sp2 and Sp3) were digested with trypsin. For these analyses, a saturated solution of α -cyano-4-hydroxycinnamic acid (CCA) prepared in ACN:H₂O (30:70, v/v) with 0.1% trifluoroacetic acid was used. The samples were dissolved in ACN:H₂O (30:70, v/v) with 0.1% trifluoroacetic acid. The sample solution and the matrix were mixed in equal amounts and spotted onto a ground stainless steel MALDI target (1 μ l). The peptide fractions were analyzed by MS and MS/MS. For MS analyses, the experimental conditions used were as follows: pulsed ion extraction of 100 ns, laser frequency of 1000 Hz, reflectron mode, and positive ion mode. In all, 600 shots were averaged to record each mass spectrum. The selected ions were accelerated to 19 kV on the LIFT cell for MS/MS analyses. External calibrations were performed using peptide calibration standards (angiotensin II and I, substance P, bombesin, ACTH clip 1-17, ACTH clip 18-39 and somatostatin) and protein standards I (insulin, ubiquitin I, cytochrome C and myoglobin) and II (trypsinogen, protein A and bovine serum albumin (BSA)) from Bruker Daltonics.

Database identification and CSP37 prediction

Database identification: MS/MS spectra were analyzed and converted to mfg files using Mascot Distiller (MATRIX SCIENCES, London, UK). Protein identification was performed by database searches with a local Mascot Server v 2.3 (MATRIX SCIENCES, London, UK) against the proteome sequence of *Capsicum annum*.⁴⁸ The parameters used for identification allowed a maximum of one missed tyrosine cleavage, the carbamidomethylation of cysteine, and the possible oxidation of methionine. The peptide MS/MS tolerance was 0.25 Da, and the significance threshold was set at $p < 0.05$. The sequences identified by MASCOT as significant hits were annotated using the NCBI BLAST program and the NCBI Conserved Domain Database. CSP37 prediction: since the genome/proteome of *C. baccatum* is not fully sequenced, we used an alternative strategy to identify the principal peptides predicted by Mascot, which led to the identification of CSP37. Proteins containing the predicted peptides were identified by using P-BLAST to search a database of annotated *Capsicum* sequences. This led to the identification of two proteins (XP_016559134.1, 2S albumin-like [*Capsicum annum*] and XP_016567465.1, 2S sulfur-rich seed storage protein 2-like [*Capsicum annum*]) with very strong sequence identity. We then realigned the two proteins and the Mascot peptides using Clustal Omega. We discovered that the two peptides were in fact two consecutive segments (separated by a single residue) of a single longer peptide (CSP37), whose mass indeed corresponded to the most intense peak of the MS spectrum.

Statistical analysis

The results are expressed as the mean \pm standard deviation. Biofilm and growth data were analyzed using Student's t-test, and p-values <0.05 were considered statistically significant. Cytotoxicity data were analyzed using ANOVA followed by Tukey's post hoc test, and p-values <0.05 were considered statistically significant.

Results

Bioguided fractionation and identification of antibiofilm peptides from *C. baccatum*

Previously, we obtained a protein extract from *C. baccatum* seeds (RAqS) with potential antibiofilm activity.⁴³ Herein, further purification steps were performed in order to obtain the bioactive fraction from RAqS responsible for antibiofilm activity. After chromatographic collection of 14 fractions, the ninth fraction was determined to be the one containing the biological activity (Figure S2). Once identified, this antibiofilm active fraction was subjected to chemical elucidation. Briefly, MALDI-MS analyses did not show ions up to m/z 4000, but some ions were observed in the range of 4 to 20 kDa (Figure 1), characteristic of peptides abundant in *Capsicum*.^{49,50} The m/z 4305 ion showed the highest relative ion intensity in the mass spectrum of the red pepper active fraction, and less intense ions were observed at m/z 4220, 4364, 9372 and 9415. No ions were observed in the mass range 20-100 kDa, moreover, LC-DAD-ESI-MS analysis did not reveal secondary metabolites of low molecular weight. Thus, the red pepper active fraction was subjected to gel electrophoresis, and the obtained spots (Sp1, Sp2 and Sp3) were analyzed by MALDI-MS and MS/MS to identify the protein precursors. All spots were found to contain peptides belonging to the same protein family, the 2S sulfur-rich seed storage protein (2SS protein) family, according to the NCBI CDD and BLAST searches using the proteome of *C. annum* as the database (the *C. baccatum* proteome is unknown) (Figure 1, Table 1). Therefore, the red pepper active fraction is composed of peptides derived from a 2SS protein (*C. baccatum*) and will be referred to herein as "*Capsicum* storage peptides (CSP)".

Figure 1

CSP control methicillin-resistant *S. epidermidis* biofilm formation

Staphylococcus epidermidis biofilm formation was completely prevented in the presence of CSP solution (2.0 mg/mL) (Figure 2A). Remarkably, CSP did not influence bacterial growth at the same concentration (Figure 2A). In addition, bacterial growth kinetics confirmed the absence of any antibiotic effect of CSP (Figure 2B). Although biofilm formation was prevented, the planktonic cells remained viable, and the colonies were phenotypically normal.

Figure 2

Red pepper peptide coatings strongly prevent *S. epidermidis* adhesion

To verify the antiadhesive capability of CSP, we applied the spin-coat technique to coat Permanox™ slides with CSP (herein called "red pepper coat"), exposed the slides to *S. epidermidis* cultures and analyzed them by scanning electron microscopy (SEM) after 24 h of exposure (Figure 3A-J). Red pepper-coated surfaces prevented bacterial adhesion, aggregation and accumulation (Figure 3C-J). In addition, the noncoated control that received 2.0 mg/mL soluble CSP in the culture medium did

not display any adherent bacteria (Figure S8, Supplementary Material). Compared to controls, bacterial adhesion and aggregation profiles decreased on slides that had been coated once with 2.0 mg/ml CSP (Figure 3C-D). Slides that had been coated three times with 2.0 mg/ml CSP show only a few individual cells, although extracellular matrix was not detected (Figure 3G-H). Likewise, slides coated with a higher CSP concentration (10.0 mg/ml) displayed a complete absence of bacteria and extracellular matrix (Figure 3E-F, I-J). Control slides (vehicle-coated and noncoated slides without CSP solution) were not able to prevent the bacterial adhesion, aggregation and accumulation that facilitate biofilm formation (Figures 3A-B and S3, Supplementary Material).

Red pepper peptide coatings create a hydrophilic surface on coated material

To evaluate the hydrophilicity and the quality of red pepper coatings produced by spin coating (Figure 3k), we used the water contact angle (WCA) image and measurement analysis. Red pepper coatings displayed hydrophilic characteristics in a concentration-dependent manner compared to controls. The surfaces singly and triply coated with 2.0 mg/mL CSP displayed WCAs of 75° and 81°, respectively. At 10.0 mg/mL, the WCA decreased to 26° and 18° for singly and triply coated surfaces, respectively. However, noncoated and methanol-coated surfaces exhibited hydrophobic characteristics, displaying WCAs of 87° and 92°, respectively.

Figure 3

CSP37 is the lead antibiofilm peptide from the CSP fraction

We postulated that the peptide detected with higher intensity in the CSP active fraction (m/z 4289, Figure 1) would mediate CSP bioactivity. Thus, we performed sequence and structural predictions of this major peptide of CSP. For this purpose, we used the CSP MS/MS data and sequence alignment analysis to determine its sequence: RSCQQIQQAQQLSSCQQYLKQRVQSEEGEDQISQRE (European Patent n°EP19305205) (Figure 4A-C). The predicted peptide was named "*Capsicum* storage peptide 37 (CSP37)" and was subsequently synthesized and experimentally tested. CSP37 prevented *S. epidermidis* biofilm formation independently of growth inhibition, equivalently to CSP (Figure 4D). Their bioactivities (Antibiofilm, non-antibiotic and non-cytotoxic) support the correlation between them (Figure 4D, Figure 2A and Figure 5).

Figure 4

Red pepper peptides are not cytotoxic

To evaluate CSP cytotoxicity, human erythrocytes and lymphocytes were used as model cells. CSP at 2.0 mg/ml displayed a slight hemolytic activity (11% hemolysis), preserving the erythrocytes integrity similarly to the PBS control (Figure 5B). Equally, CSP did not present cytotoxic effects against

lymphocytes after 24 h of exposure (7.7% decrease in cell counts) (Figures 5A, C-E). Furthermore, the LDH assay indicated that CSP did not affect lymphocyte membrane integrity (LDH release by Triton treatment was considered 100% for quantification, the normal control showed 17.3%, and CSP 14.7%). Likewise, high content analysis (HCA) showed no CSP37 cytotoxicity against 7 different representative human cell lines (Figure 5F).

Figure 5

Discussion

Biofilm-forming *S. epidermidis* is an important pathogen worldwide and is closely linked with medical device- and immunosuppression-associated infections.⁵¹⁻⁵³ The present study demonstrates that natural *Capsicum* storage peptides (CSP) exhibit potent antibiofilm activity against methicillin-resistant *S. epidermidis* by a mechanism that does not involve cell killing. The study presents the identification of CSP37, the major peptide of CSP and a source of its antibiofilm activity. Strikingly, the results of our coating experiments support the antiadhesive applicability of these peptides.

Plants are constantly exposed to a large variety of pathogenic microorganisms, and they produce natural compounds as resistance factors, such as constitutive peptides and proteins.⁵⁴⁻⁵⁶ Peptides are a very common chemical component of *Capsicum* seeds.^{50,57} In this context, some studies have suggested that plant-derived peptide fractions exhibit antiadhesive properties.^{43,58-60} Recent literature reviews have shown that peptides and peptidomimetics from different sources have been investigated for their prominent antibiofilm activity.⁶¹⁻⁶⁵

There are no previous studies demonstrating a relationship between 2S sulfur-rich seed storage (2SS) proteins or peptides and antibiofilm activity. To the best of our knowledge, this is the first scientific report using *Capsicum* natural peptides as antibiofilm and antiadhesive agents, herein called *red pepper coat*. The purification of *C. baccatum* extract allowed the chemical identification of the active fraction able to prevent *S. epidermidis* biofilm formation. This antibiofilm fraction is composed of peptides from a *Capsicum* 2SS protein, herein named *Capsicum* storage peptides (CSP). Hence, the reconstitution of the major MS peak of the CSP spectrum allowed the identification of its main peptide (CSP37, European Patent n° EP19305205), consisting of a 37 amino acid sequence that is also present in the 2SS protein described in the *Capsicum* genus.

The NCBI database describes the 2SS protein and the 2S albumin-like protein as members of the Alpha-Amylase Inhibitors (AAIs) and Seed Storage (SS) Protein subfamilies. They are present in the seeds of a variety of plants and are widely distributed in dicot seeds, and they play an important role in plant natural defenses (NCBI accession: XP_016567465.1, CAA01774.1, NP_001295683.1, NP_001078429.1). Despite differences in their subunit structure and synthesis, all are compact globular proteins with conserved cysteine residues.⁶⁶

Notably, both CSPs and CSP37 strongly decrease *S. epidermidis* biofilm formation without antibiotic effects. These results establish that the antibiofilm activity of CSP and CSP37 utilizes a mechanism of action that is independent of cell killing and of antibiotic mechanisms. Accordingly, nonantibiotic strategies have emerged as alternatives to combat bacterial resistance and tolerance, such as the targeting of extracellular bacterial virulence factors. This strategy is very important because it is less likely to impact bacteria gene machinery, and it may include antibiofilm agents to reduce bacteria adhesion.^{15,67-71} In this sense, the prototype red pepper coat (CSP spin-coated slides) prevented bacterial adhesion, aggregation, accumulation and consequently biofilm establishment. This activity seems to be concentration dependent.

The antiadhesive effect of red pepper coatings may be due to multiple causes, including electrostatic interactions and steric hindrance. Repulsion forces may act between the anionic Gram-positive bacterial surface and biofilm matrix and the electronegative field of the coated surface (Figure 6). The film produced by CSP coating may also give rise to steric hindrance that blocks bacterium-substratum interactions.²⁹ Moreover, CSP are mainly composed of polar amino acids, leading to important hydrophilic characteristics. Supported by angle contact analysis, red pepper-coated surfaces presented increased hydrophilicity on the entire coated surface compared to controls (hydrophobic material), showing that CSP occupy and alter the hydrophobic characteristics of Permanox™. Bacterial attachment is initially dependent on their hydrophobic and electrostatic physicochemical characteristics as well as on the surfaces involved; thus, the hydrophilic character of red pepper coat contributes to a reduction in these interactions.^{29,72,73}

Figure 6

In addition, both activities remain, even after 24 h under the incubation conditions. The spin-coating technique anchors substances on surfaces and is appropriate for the study of these coatings during transient or short-term exposure, which is sufficient for establishing proof of concept that natural products may act against pathogenic bacterial adhesion, although stronger interactions such as covalent attachment are required to extend coating lifetime for long-term studies.

Importantly, CSP was not cytotoxic to human erythrocytes and lymphocytes. CSP37, the proposed major peptide of CSP, showed no cytotoxicity against 7 representative human cell lines. Furthermore, previous studies with *C. baccatum* corroborate this favorable toxicological profile. For example, RAqS, the precursor of CSP, was not toxic to *Galleria mellonella* larvae⁴³ which presents similarities to the innate immune response of mammals like humoral and cellular components, and other extracts from *C. baccatum* displayed a good level of pharmacological safety during 60 days of oral administration in mice.³⁷ These results suggest that red pepper coatings do not affect either eukaryotic or prokaryotic membranes.^{74,75}

Conclusions

In conclusion, this study shows that red pepper peptide coatings are a promising antibiofilm and antiadhesive alternative to prevent methicillin-resistant *S. epidermidis* colonization. Importantly, components of red pepper coatings are not cytotoxic and act by a nonantibiotic mechanism of action. In addition, CSP37 may facilitate research for innovative molecular designs as a prime structure. Importantly, this study sheds light on the use of natural products and their derivatives, which could bring innovative solutions for materials science. The identification of novel peptides aims to overcome the lack of therapies against biofilm infections and to contribute to the development of anti-infective medical devices.

Conflicts of interest

R.G.V.B, S.C.B.G, A.R.Z, A.J.M and R.G. are co-authors of a patent register of CSP37. Application number EP19305205.7 at European Patent Office. Specific aspect of manuscript covered in patent application: its amino acid sequence and bioactivity.

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References

1. Lim K, Chua RR, Bow H, Tambyah PA, Hadinoto K, Leong SS. Development of a catheter functionalized by a polydopamine peptide coating with antimicrobial and antibiofilm properties. *Acta Biomater* 2015;15:127-38.
2. Shah AR, Goyal RK. Current status of the regulation for medical devices. *Indian J Pharm Sci* 2008;70(6):695-700.
3. Levin-Reisman I, Ronin I, Gefen O, Braniss I, Shores N, Balaban NQ. Antibiotic tolerance facilitates the evolution of resistance. *Science* 2017;355(6327):826-830.

4. Heilmann C, Ziebuhr W, Becker K. Are coagulase-negative staphylococci virulent? *Clin Microbiol Infect* 2018.
5. Meneguetti MG, Ciol MA, Bellissimo-Rodrigues F, Auxiliadora-Martins M, Gaspar GG, Canini SRMD, Basile-Filho A, Laus AM. Long-term prevention of catheter-associated urinary tract infections among critically ill patients through the implementation of an educational program and a daily checklist for maintenance of indwelling urinary catheters: A quasi-experimental study. *Medicine (Baltimore)* 2019;98(8):e14417.
6. Olsen I. Biofilm-specific antibiotic tolerance and resistance. *Eur J Clin Microbiol Infect Dis* 2015;34(5):877-86.
7. Davies D. Understanding biofilm resistance to antibacterial agents. *Nat Rev Drug Discov* 2003;2(2):114-22.
8. Romling U, Balsalobre C. Biofilm infections, their resilience to therapy and innovative treatment strategies. *Journal of Internal Medicine* 2012;272(6):541-561.
9. Haussler S, Fuqua C. Biofilms 2012: New Discoveries and Significant Wrinkles in a Dynamic Field. *Journal of Bacteriology* 2013;195(13):2947-2958.
10. Hall CW, Mah TF. Molecular mechanisms of biofilm-based antibiotic resistance and tolerance in pathogenic bacteria. *FEMS Microbiol Rev* 2017;41(3):276-301.
11. Post V, Harris LG, Morgenstern M, Mageiros L, Hitchings MD, Méric G, Pascoe B, Sheppard SK, Richards RG, Moriarty TF. Comparative Genomics Study of Staphylococcus epidermidis Isolates from Orthopedic-Device-Related Infections Correlated with Patient Outcome. *J Clin Microbiol* 2017;55(10):3089-3103.
12. Arciola CR, Campoccia D, Speziale P, Montanaro L, Costerton JW. Biofilm formation in Staphylococcus implant infections. A review of molecular mechanisms and implications for biofilm-resistant materials. *Biomaterials* 2012;33(26):5967-5982.
13. Lim K, Chua RRY, Saravanan R, Basu A, Mishra B, Tambyah PA, Ho B, Leong SSJ. Immobilization Studies of an Engineered Arginine-Tryptophan Rich Peptide on a Silicone Surface with Antimicrobial and Antibiofilm Activity (vol 5, pg 6412, 2013). *Acs Applied Materials & Interfaces* 2013;5(17):8821-8821.
14. Madeo J, Frieri M. Bacterial biofilms and chronic rhinosinusitis. *Allergy and Asthma Proceedings* 2013;34(4):335-341.
15. Travier L, Rendueles O, Ferrieres L, Herry J-M, Ghigo J-M. Escherichia coli Resistance to Nonbiocidal Antibiofilm Polysaccharides Is Rare and Mediated by Multiple Mutations Leading to Surface Physicochemical Modifications. *Antimicrobial Agents and Chemotherapy* 2013;57(8):3960-3968.
16. Macedo AJ, Abraham W-R. Can Infectious Biofilm be Controlled by Blocking Bacterial Communication? *Medicinal Chemistry* 2009;5(6):517-528.
17. Chusri S, Sompetch K, Mukdee S, Jansrisewangwong S, Srichai T, Maneenoon K, Limsuwan S, Voravuthikunchai SP. Inhibition of Staphylococcus epidermidis Biofilm Formation by Traditional Thai Herbal Recipes Used for Wound Treatment. *Evidence-Based Complementary and Alternative Medicine* 2012.
18. Laverty G, Gorman SP, Gilmore BF. Biomolecular mechanisms of staphylococcal biofilm formation. *Future Microbiology* 2013;8(4):509-524.
19. Chessa D, Ganau G, Spiga L, Bulla A, Mazzarello V, Campus GV, Rubino S. Staphylococcus aureus and Staphylococcus epidermidis Virulence Strains as Causative Agents of Persistent Infections in Breast Implants. *PLoS One* 2016;11(1):e0146668.
20. Vassallo M, Genillier PL, Dunais B, Kaphan R, Saudes L, Duval Y, Rolland F, Jullien V, Weiss N, Blanchouin E and others. Short-course daptomycin lock and systemic therapy for catheter-related bloodstream infections: a retrospective cohort study in cancer patients with surgically implanted devices. *J Chemother* 2017;29(4):232-237.
21. Oliveira WF, Silva PMS, Silva RCS, Silva GMM, Machado G, Coelho LCBB, Correia MTS. Staphylococcus aureus and Staphylococcus epidermidis infections on implants. *J Hosp Infect* 2018;98(2):111-117.
22. Lee JYH, Monk IR, Gonçalves da Silva A, Seemann T, Chua KYL, Kearns A, Hill R, Woodford N, Bartels MD, Strommenger B and others. Global spread of three multidrug-resistant lineages of Staphylococcus epidermidis. *Nat Microbiol* 2018.
23. Méric G, Mageiros L, Pensar J, Laabei M, Yahara K, Pascoe B, Kittiwat N, Tadee P, Post V, Lambie S and others. Disease-associated genotypes of the commensal skin bacterium Staphylococcus epidermidis. *Nat Commun* 2018;9(1):5034.
24. Otto M. Staphylococcal biofilms. *Bacterial Biofilms* 2008;322:207-228.
25. Li B, Webster TJ. Bacteria antibiotic resistance: New challenges and opportunities for implant-associated orthopedic infections. *J Orthop Res* 2018;36(1):22-32.
26. Campoccia D, Montanaro L, Arciola CR. A review of the biomaterials technologies for infection-resistant surfaces. *Biomaterials* 2013;34(34):8533-54.
27. Ashton NN, Allyn G, Porter ST, Haussener TJ, Sebahar PR, Looper RE, Williams DL. In vitro testing of a first-in-class tri-alkylmorpholine-biaryl antibiotic in an anti-biofilm silicone coating. *Acta Biomater* 2019.
28. Gao Q, Yu M, Su Y, Xie M, Zhao X, Li P, Ma PX. Rationally designed dual functional block copolymers for bottlebrush-like coatings: In vitro and in vivo antimicrobial, antibiofilm, and antifouling properties. *Acta Biomater* 2017;51:112-124.
29. Trentin DS, Silva DB, Frasson AP, Rzhapishvskaya O, da Silva MV, Pulcini ED, James G, Soares GV, Tasca T, Ramstedt M and others. Natural Green Coating Inhibits Adhesion of Clinically Important Bacteria. *Scientific Reports* 2015;5.
30. Qin C, Yu C, Shen Y, Fang X, Chen L, Min J, Cheng J, Zhao S, Xu M, Luo Y and others. Whole-genome sequencing of cultivated and wild peppers provides insights into Capsicum domestication and specialization. *Proc Natl Acad Sci U S A* 2014;111(14):5135-40.
31. Govindarajan VS. CAPSICUM - PRODUCTION, TECHNOLOGY, CHEMISTRY, AND QUALITY .3. CHEMISTRY OF THE COLOR, AROMA, AND PUNGENCY STIMULI. *Crc Critical Reviews in Food Science and Nutrition* 1986;24(3):245-355.
32. Menichini F, Tundis R, Bonesi M, Loizzo MR, Conforti F, Statti G, De Cindio B, Houghton PJ, Menichini F. The influence of fruit ripening on the phytochemical content and biological activity of Capsicum chinense Jacq. cv Habanero. *Food Chemistry* 2009;114(2):553-560.

33. Kim S, Park M, Yeom SI, Kim YM, Lee JM, Lee HA, Seo E, Choi J, Cheong K, Kim KT and others. Genome sequence of the hot pepper provides insights into the evolution of pungency in *Capsicum* species. *Nat Genet* 2014;46(3):270-8.
34. Meghvansi MK, Siddiqui S, Khan MH, Gupta VK, Vairale MG, Gogoi HK, Singh L. Naga chilli: A potential source of capsaicinoids with broad-spectrum ethnopharmacological applications. *Journal of Ethnopharmacology* 2010;132(1):1-14.
35. Srinivasan M, Devipriya N, Kalpana KB, Menon VP. Lycopene: An antioxidant and radioprotector against gamma-radiation-induced cellular damages in cultured human lymphocytes. *Toxicology* 2009;262(1):43-49.
36. Zimmer AR, Leonardi B, Miron D, Schapoval E, de Oliveira JR, Gosmann G. Antioxidant and anti-inflammatory properties of *Capsicum baccatum*: From traditional use to scientific approach. *Journal of Ethnopharmacology* 2012a;139(1):228-233.
37. Zimmer AR, Leonardi B, Zimmer ER, Kalinine E, de Souza DO, Portela LV, Gosmann G. Long-Term Oral Administration of *Capsicum baccatum* Extracts Does Not Alter Behavioral, Hematological, and Metabolic Parameters in CF1 Mice. *Evid Based Complement Alternat Med* 2012b;2012:196358.
38. Ahuja KDK, Robertson IK, Geraghty DP, Ball MJ. The effect of 4-week chilli supplementation on metabolic and arterial function in humans. *European Journal of Clinical Nutrition* 2007;61(3):326-333.
39. Aizawa K, Inakuma T. Dietary capsanthin, the main carotenoid in paprika (*Capsicum annuum*), alters plasma high-density lipoprotein-cholesterol levels and hepatic gene expression in rats. *British Journal of Nutrition* 2009;102(12):1760-1766.
40. Galvez Ranilla L, Kwon Y-I, Apostolidis E, Shetty K. Phenolic compounds, antioxidant activity and in vitro inhibitory potential against key enzymes relevant for hyperglycemia and hypertension of commonly used medicinal plants, herbs and spices in Latin America. *Bioresource Technology* 2010;101(12):4676-4689.
41. Liu Y, Nair MG. Capsaicinoids in the Hottest Pepper Bhut Jolokia and its Antioxidant and Antiinflammatory Activities. *Natural Product Communications* 2010;5(1):91-94.
42. Sharma SK, Vij AS, Sharma M. Mechanisms and clinical uses of capsaicin. *Eur J Pharmacol* 2013;720(1-3):55-62.
43. Gomes Von Borowski R, Rigon Zimmer K, Franco Leonardi B, Silva Trentin D, Campos Silva R, Primon de Barros M, José Macedo A, Cristina Baggio Gnoatto S, Gosmann G, Rigon Zimmer A. Red pepper *Capsicum baccatum* : source of antiadhesive and antibiofilm compounds against nosocomial bacteria. *Ind. Crops Prod.* 2019;127:148-157.
44. Rocha T, Vieira P, Gnoatto S, Tasca T, Gosmann G. Anti-Trichomonas vaginalis activity of saponins from Quillaja, Passiflora, and Ilex species. *Parasitology Research* 2012;110(6):2551-2556.
45. Waechter F, da Silva GNS, Willig JB, de Oliveira CB, Vieira BD, Trivella DBB, Zimmer AR, Buffon A, Pilger DA, Gnoatto SCB. Design, Synthesis and Biological Evaluation of Betulinic Acid Derivatives as New Antitumor Agents for Leukemia. *Anticancer Agents Med Chem* 2017;17(13):1777-1785.
46. Richy N, Sarraf D, Maréchal X, Janmamode N, Le Guével R, Genin E, Reboud-Ravaux M, Vidal J. Structure-based design of human immuno- and constitutive proteasomes inhibitors. *Eur J Med Chem* 2018;145:570-587.
47. Rodriguez Castillo AS, Guihéneuf S, Le Guével R, Biard PF, Paquin L, Amrane A, Couvert A. Synthesis and toxicity evaluation of hydrophobic ionic liquids for volatile organic compounds biodegradation in a two-phase partitioning bioreactor. *J Hazard Mater* 2016;307:221-30.
48. Kim S, Park M, Yeom S-I, Kim Y-M, Lee JM, Lee H-A, Seo E, Choi J, Cheong K, Kim K-T and others. Genome sequence of the hot pepper provides insights into the evolution of pungency in *Capsicum* species. *Nature Genetics* 2014;46(3):270-+.
49. Lee SC, Hwang IS, Choi HW, Hwang BK. Involvement of the pepper antimicrobial protein CaAMP1 gene in broad spectrum disease resistance. *Plant Physiology* 2008;148(2):1004-1020.
50. Dias GB, Gomes VM, Pereira UZ, Ferreira Ribeiro SF, Carvalho AO, Rodrigues R, Tavares Machado OL, Sales Fernandes KV, Ferreira ATS, Perales J and others. Isolation, Characterization and Antifungal Activity of Proteinase Inhibitors from *Capsicum chinense* Jacq. Seeds. *Protein Journal* 2013;32(1):15-26.
51. Lourtet-Hascoët J, Bicart-See A, Félicé MP, Giordano G, Bonnet E. Staphylococcus lugdunensis, a serious pathogen in periprosthetic joint infections: comparison to Staphylococcus aureus and Staphylococcus epidermidis. *Int J Infect Dis* 2016;51:56-61.
52. Widerström M. Significance of Staphylococcus epidermidis in Health Care-Associated Infections, from Contaminant to Clinically Relevant Pathogen: This Is a Wake-Up Call! *J Clin Microbiol* 2016;54(7):1679-1681.
53. Gabriel EM, Fitzgibbon S, Clair J, Coffey A, O'Mahony JM. Characterisation of clinical meticillin-resistant Staphylococcus epidermidis demonstrating high levels of linezolid resistance (>256 µg/ml) resulting from transmissible and mutational mechanisms. *J Infect Chemother* 2015;21(7):547-9.
54. Castro MS, Fontes W. Plant defense and antimicrobial peptides. *Protein and Peptide Letters* 2005;12(1):13-18.
55. Silva FAC, de Sousa Oliveira M, de Souza JM, Martins PGS, Pestana-Calsa MC, Junior TC. Plant Proteomics and Peptidomics in Host-Pathogen Interactions: The Weapons Used by Each Side. *Curr Protein Pept Sci* 2017;18(4):400-410.
56. de Vries J, Evers JB, Poelman EH. Dynamic Plant-Plant-Herbivore Interactions Govern Plant Growth-Defence Integration. *Trends Plant Sci* 2017;22(4):329-337.
57. Lee YM, Wee HS, Ahn IP, Lee YH, An CS. Molecular characterization of a cDNA for a cysteine-rich antifungal protein from *Capsicum annuum*. *Journal of Plant Biology* 2004;47(4):375-382.
58. Lengsfeld C, Titgemeyer F, Faller G, Hensel A. Glycosylated compounds from okra inhibit adhesion of Helicobacter pylori to human gastric mucosa. *Journal of Agricultural and Food Chemistry* 2004;52(6):1495-1503.
59. Wittschier N, Lengsfeld C, Vorthems S, Stratmann U, Ernst JF, Verspohl EJ, Hensel A. Large molecules as anti-adhesive compounds against pathogens. *Journal of Pharmacy and Pharmacology* 2007;59(6):777-786.

60. Bensch K, Tiralongo J, Schmidt K, Matthias A, Bone KM, Lehmann R, Tiralongo E. Investigations into the antiadhesive activity of herbal extracts against *Campylobacter jejuni*. *Phytother Res* 2011;25(8):1125-32.
61. Von Borowski RG, Macedo AJ, Gnoatto SCB. Peptides as a strategy against biofilm-forming microorganisms: Structure-activity relationship perspectives. *Eur J Pharm Sci* 2017;114:114-137.
62. Gomes Von Borowski R, Gnoatto SCB, Macedo AJ, Gillet R. Promising Antibiofilm Activity of Peptidomimetics. *Front Microbiol* 2018;9:2157.
63. Batoni G, Maisetta G, Esin S. Antimicrobial peptides and their interaction with biofilms of medically relevant bacteria. *Biochimica Et Biophysica Acta-Biomembranes* 2016;1858(5):1044-1060.
64. de la Fuente-Nunez C, Reffuveille F, Mansour SC, Reckseidler-Zenteno SL, Hernandez D, Brackman G, Coenye T, Hancock REW. D-Enantiomeric Peptides that Eradicate Wild-Type and Multidrug-Resistant Biofilms and Protect against Lethal *Pseudomonas aeruginosa* Infections (vol 22, pg 196, 2015). *Chemistry & Biology* 2015;22(9):1280-1282.
65. Pfalzgraff A, Brandenburg K, Weindl G. Antimicrobial Peptides and Their Therapeutic Potential for Bacterial Skin Infections and Wounds. *Front Pharmacol* 2018;9:281.
66. Shewry PR, Napier JA, Tatham AS. SEED STORAGE PROTEINS - STRUCTURES AND BIOSYNTHESIS. *Plant Cell* 1995;7(7):945-956.
67. Dostert M, Belanger CR, Hancock REW. Design and Assessment of Anti-Biofilm Peptides: Steps Toward Clinical Application. *J Innate Immun* 2018;1-12.
68. Sun H, Lv L, Bai Y, Yang H, Zhou H, Li C, Yang L. Nanotechnology-enabled materials for hemostatic and anti-infection treatments in orthopedic surgery. *Int J Nanomedicine* 2018;13:8325-8338.
69. Brannon JR, Hadjifrangiskou M. The arsenal of pathogens and antivirulence therapeutic strategies for disarming them. *Drug Des Devel Ther* 2016;10:1795-806.
70. Totsika M. Disarming pathogens: benefits and challenges of antimicrobials that target bacterial virulence instead of growth and viability. *Future Med Chem* 2017;9(3):267-269.
71. Silva LN, Zimmer KR, Macedo AJ, Trentin DS. Plant Natural Products Targeting Bacterial Virulence Factors. *Chem Rev* 2016;116(16):9162-236.
72. Nostro A, Scaffaro R, D'Arrigo M, Botta L, Filocamo A, Marino A, Bisignano G. Study on carvacrol and cinnamaldehyde polymeric films: mechanical properties, release kinetics and antibacterial and antibiofilm activities. *Applied Microbiology and Biotechnology* 2012;96(4):1029-1038.
73. Otto M. Staphylococcal Infections: Mechanisms of Biofilm Maturation and Detachment as Critical Determinants of Pathogenicity. *Annual Review of Medicine*, Vol 64 2013;64:175-188.
74. Atefyekta S, Pihl M, Lindsay C, Heilshorn SC, Andersson M. Antibiofilm elastin-like polypeptide coatings: functionality, stability, and selectivity. *Acta Biomater* 2019;83:245-256.
75. Stempel N, Strehmel J, Overhage J. Potential Application of Antimicrobial Peptides in the Treatment of Bacterial Biofilm Infections. *Current Pharmaceutical Design* 2015;21(1):67-84.

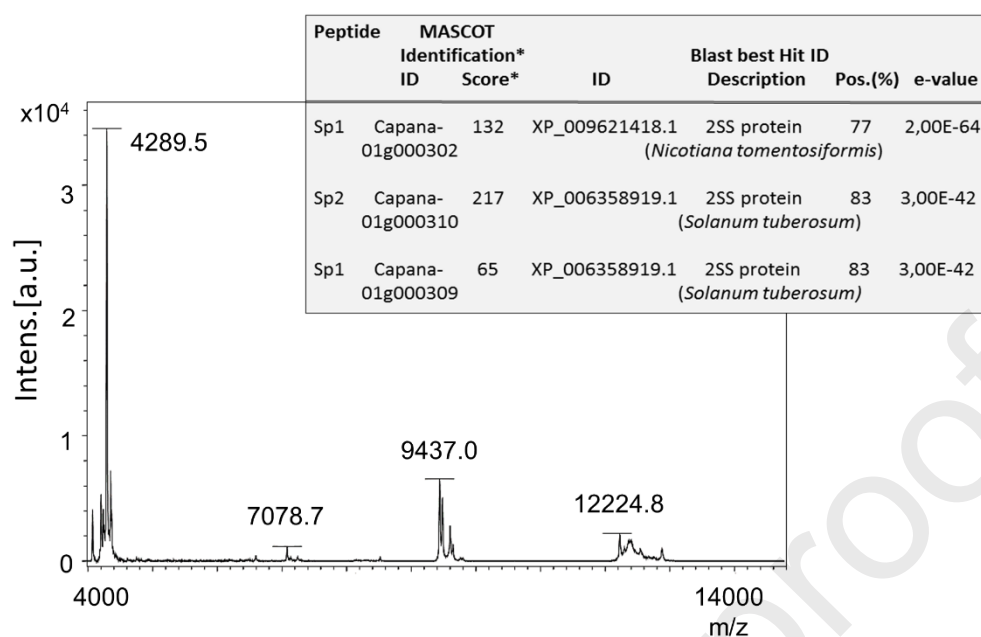


Figure 1. Identification of the peptides from *C. baccatum* active fraction. MALDI-MS mass spectra (linear, positive ion mode) of the antibiofilm active fraction peptides, range of 4,000-20,000 m/z. Table 1. (Insertion) Mascot results for CSP spots according to the NCBI CDD and BLAST searches using the proteome of *C. annuum* as the database. (2SS protein) is 2S sulfur-rich seed storage protein; *For the Capsicum proteome database in Mascot, scores > 28 indicate identity or extensive homology ($p < 0.05$).

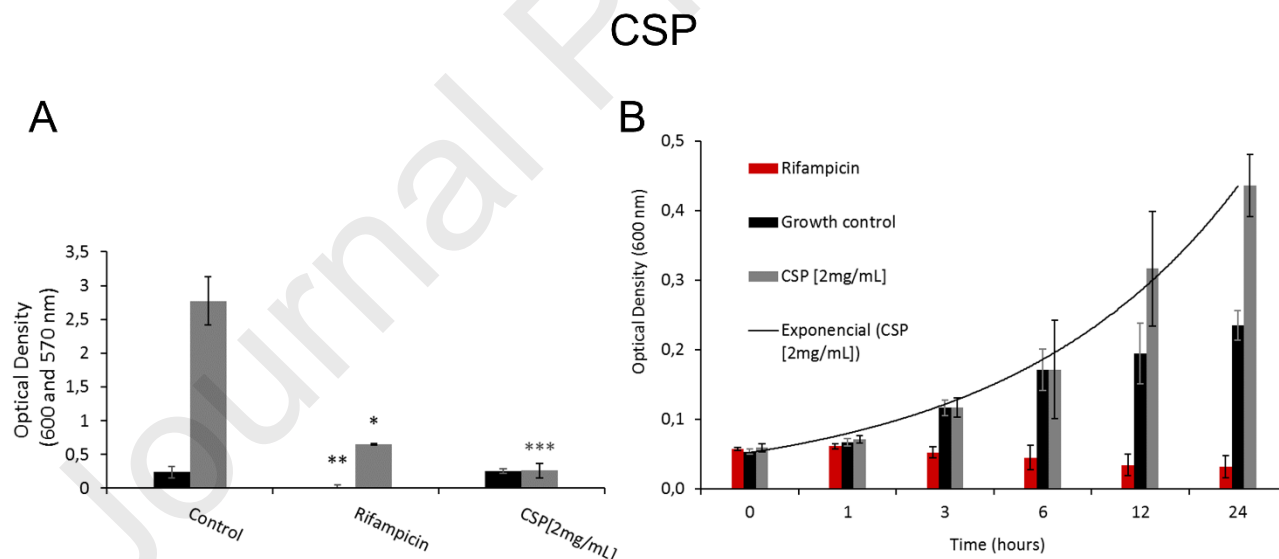


Figure 2. Bacterial biofilm formation and growth by *S. epidermidis* (ATCC 35984) exposed to natural *Capsicum* storage peptides (CSP). A) Growth (black bars) and biofilm (gray bars), both after 24 h of CSP solution exposure at 2 mg/ml. B) Bacterial growth kinetics of *S. epidermidis* exposed to CSP solution at 2 mg/ml (0, 1, 3, 6, 12 and 24 h). Untreated samples were used as growth and biofilm controls, and rifampicin (8 μ g/mL) was used as an antibiotic control. The results were monitored by optical density (OD₅₇₀ for biofilm formation and OD₆₀₀ for bacteria growth), and untreated samples were considered 100% growth and biofilm formation. (*) represents significant differences when compared to biofilm/growth control using Student's t-test; p -values < 0.05 were considered statistically significant.

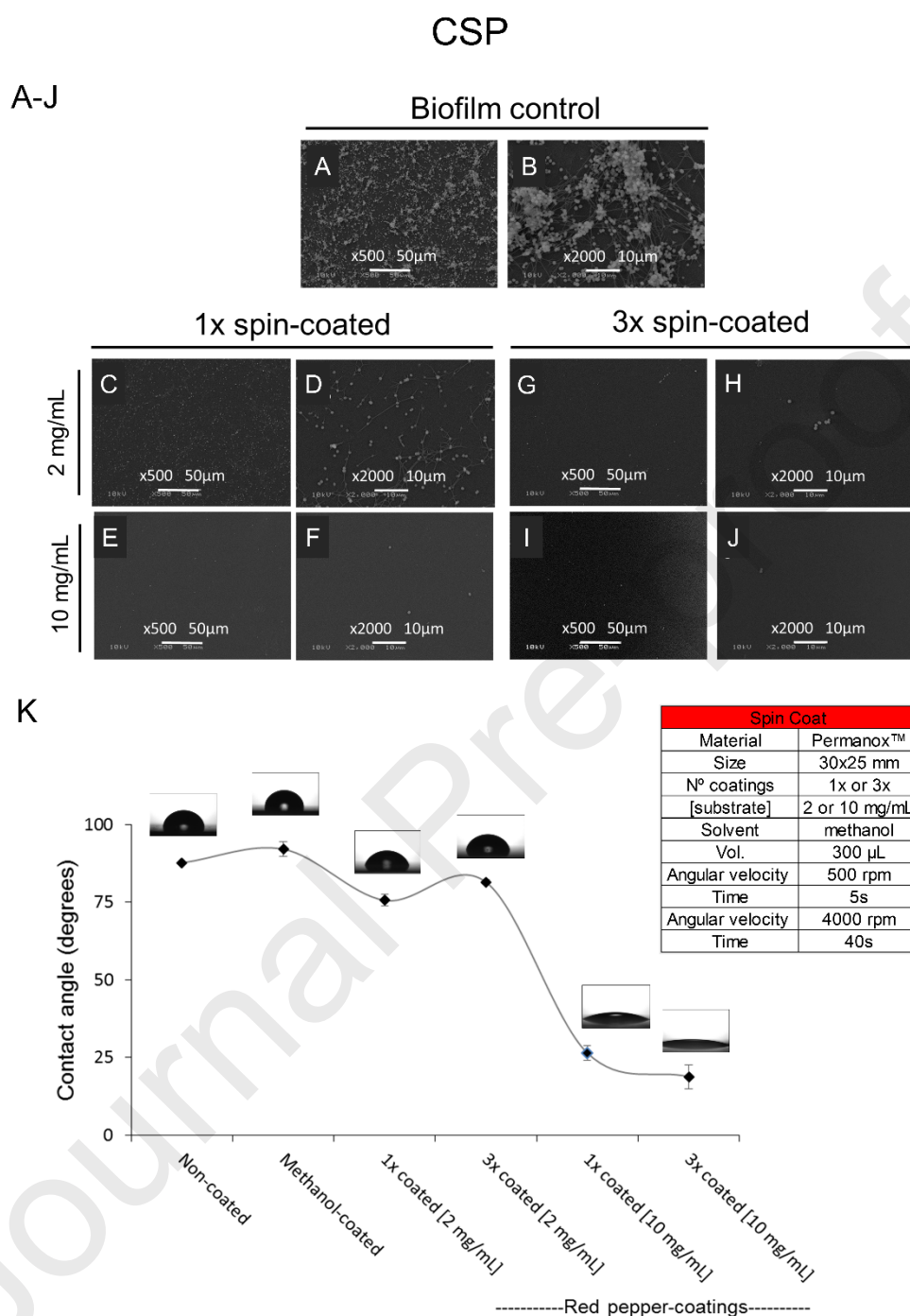


Figure 3. Scanning electron microscopy (SEM) images of Red pepper-coated slides after exposure to biofilm-forming *S. epidermidis* and the water contact angle (WCA) of Permanox™ slides before and after spin coating. A-B) Biofilm control: methanol (vehicle) was used to prepare untreated surface slides and anchored by spin coating three times. C-J) Red pepper coatings: natural capsicum storage peptides (CSP) were anchored by spin coating once or three times at 2 or 10 mg/ml. Scale bars: 500 and 2,000 x magnification. K) Graphic displaying the contact angle (degrees) results of noncoated and red pepper-coated Permanox™ slides. In the upper part, we show the photographs of the water drop and WCA, captured by a Theta Lite optical tensiometer and (Insertion) Spin Coat parameters. The WCAs are a mean of five measurements of different points of each sample.

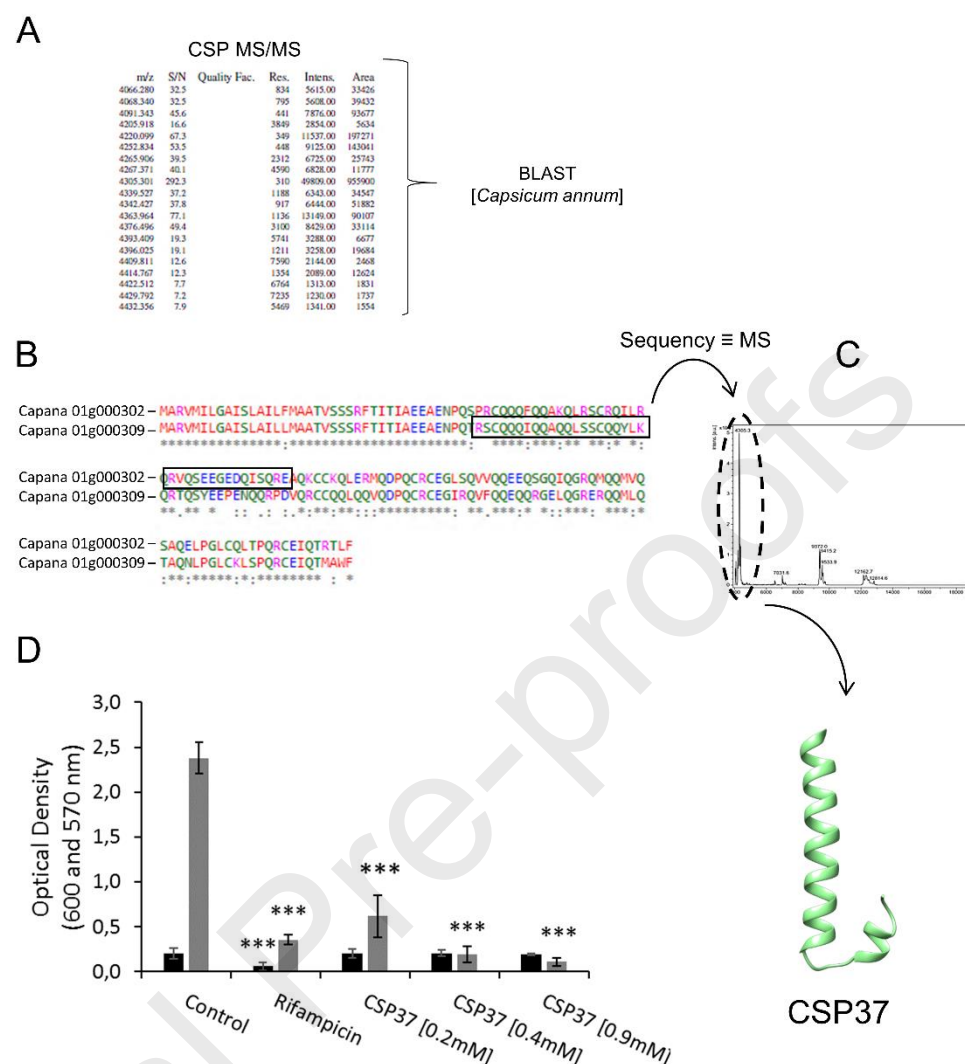


Figure 4. CSP37 prediction and activity on biofilm formation and growth by *S. epidermidis* (ATCC 35984). A) Predicted peptides by MASCOT were identified using BLAST (NCBI) and the annotated *Capsicum annuum* proteome. This lead to the identification of two separated proteins (Capana01g000302 = XP_016559134.1 and Capana01g000309 = XP_016567465.1). B) Sequence alignment of the two proteins identified with BLAST. The two MASCOT peptides are highlighted in black. The position of the two peptides in the protein sequences and the strong sequence similarity between these two proteins suggest that the two peptides can be in fact two consecutives portion of a unique protein of *Capsicum baccatum*. C) The mass of the CSP37 peptide (predicted from the union of the two precedent peptides) corresponds to the major MS pick of natural CSP fraction and, its 3D structural prediction by PEP-FOLD 3. D) Growth (black bars) and biofilm formation (gray bars) of *S. epidermidis* exposed to CSP37 (synthetic) solution at 0.23, 0.46 or 0.92 mM for 24 h. Untreated samples were used as growth and biofilm formation controls, and rifampicin (8 μ g/mL) was used as an antibiotic control. The results were monitored by optical density (OD570 for biofilm formation and OD600 for bacteria growth), and untreated samples were considered 100% growth and biofilm formation. (*) represents significant differences when compared to biofilm/growth controls using Student's t-test; p-values <0.05 were considered statistically significant.

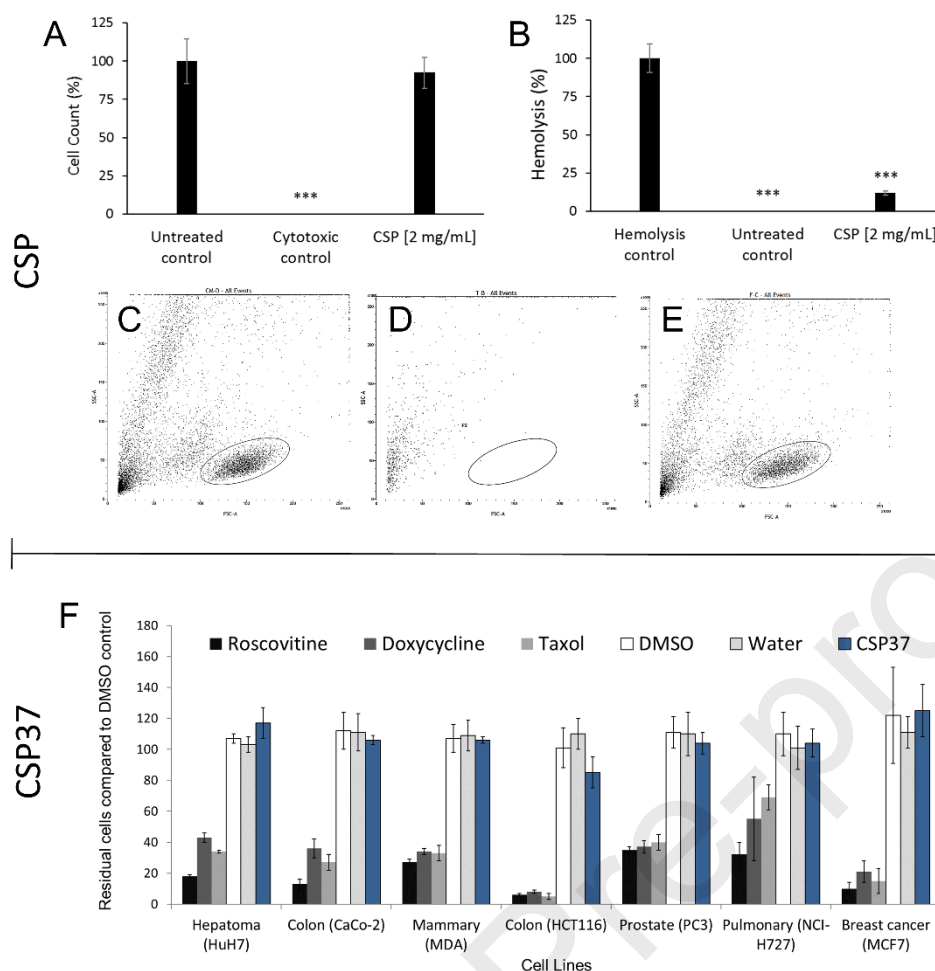


Figure 5. Cytotoxicity of natural CSP and synthetic CSP37. A) Human lymphocytes cells after 24 h of exposure to *Capsicum* storage peptides (CSP) at 2.0 mg/ml, 1% Triton solution (cytotoxic control), or RPMI 1640 medium (untreated control). B) Hemolysis test with human erythrocytes exposed to CSP at 2 mg/ml, 1% Triton solution (100% of hemolysis), or phosphate buffered saline (PBS) (untreated control as nonhemolytic control). C-E) Dot plots of lymphocytes obtained by flow cytometry. The circled areas highlight the cell populations of interest: C) untreated control, D) cytotoxic control and E) cells exposed to CSP (2.0 mg/ml). (*) Represents a significant difference when compared to reference samples using ANOVA followed by Tukey's post hoc test; p-values <0.05 were considered statistically significant. F) CSP37 cytotoxicity evaluation in representative human cell lines at 25 μ M. The number of normal cells is presented as the residual cell percentage (%) compared to the average of the control (DMSO, shown as white). The first three bars, which are black-gray, represent classic cytotoxic controls (roscovitine, doxycycline and Taxol), and the blue bar represents CSP37-exposed cells. Cells lines are described under the bars. An automated image-based cellular content analysis system (HCS/HCA) was used.

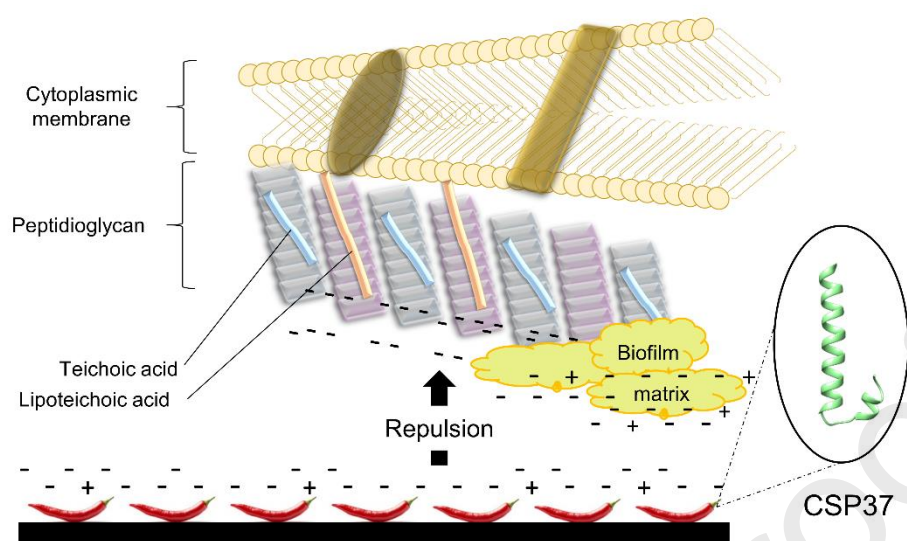
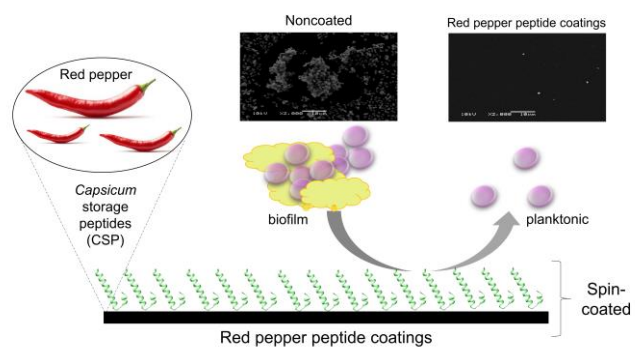


Figure 6. Proposed mechanism of anti-adhesion and of biofilm formation inhibition by red pepper coating. The effects are due to repulsive forces between the polar material surface after CSP coating and the anionic *S. epidermidis* surface and biofilm matrix.



Declaration of interests

☐ The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

☒ The authors declare the following financial interests/personal relationships which may be considered as potential competing interests:

R.G.V.B, S.C.B.G, A.R.Z, A.J.M and R.G. are co-authors of a patent register of CSP37. Application number EP19305205.7 at European Patent Office. Specific aspect of manuscript covered in patent application: its amino acid sequence and bioactivity.