



An effective acid pretreatment of agricultural biomass residues for the production of second-generation bioethanol

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An effective acid pretreatment of agricultural biomass residues for the production of second-generation bioethanol

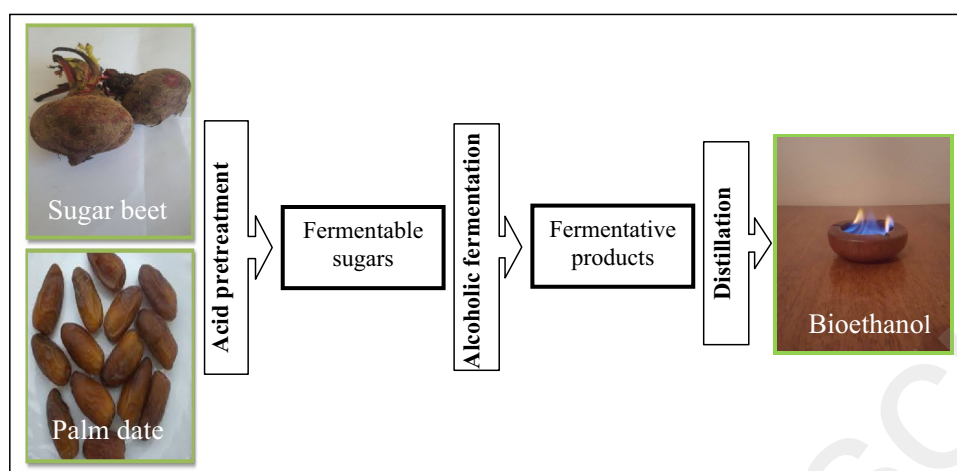
Ibtissem Fennouche¹ · Nabila Khellaf¹  · Hayet Djelal² · Abdeltif Amrane³

Abstract

Currently, the potential for energy recovery of plant biomass by biotechnological processes is a preferred solution for the use of agricultural products of low commercial value in order to produce bioenergy that is alternative to fossil fuels. The objective of this study was to obtain second-generation bioethanol by valorization of sugar beet and common dates of low quality. This involved separate hydrolysis and anaerobic fermentation process using the yeast strain *Saccharomyces cerevisiae*. Physicochemical and biochemical analyzes were carried out on beet and date substrates before and after alcoholic fermentation to determine their effect on yeast activity. The results showed that palm date was a good substrate for the microorganisms in contrast to sugar beet which required a high pretreatment in order to hydrolyze sucrose into fermentable sugars. After distillation, it was possible to recover bioethanol with a quality and a concentration depending on the substrate nature. For an initial sugar concentration of 12.0 °Brix, 74.7 g/kg DM of bioethanol (92.4 g/L) was produced by *Saccharomyces* from date syrup. In the presence of concentrated date syrup (22.2 °Brix), a lower efficiency of ethanol production was observed (78.2 g/L). It can be conclude that the diluted date substrate led to a good quality bioethanol with a high production yield.

Statement of novelty In this project, we tried to valorize agricultural residues (Algerian sugar beet and palm date of low quality) for the production of bioethanol which can substitute fossil energies. Algeria is considered as one of the major producers and exporters of palm dates in the world, and thousands of tons of dates remain unused. Thus, these agricultural by-products could be recovered instead of being released into the environment by polluting it. In order to produce bioethanol, an effective pretreatment and optimal fermentation conditions were used. This project is part of a strategy to increase the feedstock exploitation and decrease the residues while producing clean and renewable energy.

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Keywords Alcoholic fermentation · Beet juice · Bioethanol · Date syrup · Energy recovery · *Saccharomyces cerevisiae*

1 Introduction

Fossil fuels (oil and natural gas) provide fast and efficient tools of transport as well as a good source of several industrial activities. However, their reserves are limited and security of supply is problematic for many countries importing this kind of fuels. On the other hand, their contribution to environmental pollution is a major problem [1]. Contemporary societies are always interested in finding substitutes for these fuels which must be renewable and environmentally sustainable. Thus, the waste-based energy rich in renewable organic matter (biomass) constitutes good raw materials for many industries and the recovery of biomass by biotechnological processes would be an economical and sustainable solution of energy supply. Special attention should be paid to agricultural by-products (sugar beet, palm date, sugar cane, wheat, fruits, vegetables, etc.) since some fruits and vegetables of low quality cannot be integrated into human nutrition as they may have colors, tastes and aromas that are not appreciated [2–4]. Thus, a considerable part of this biomass is recognized as waste and not conflicting with food availability.

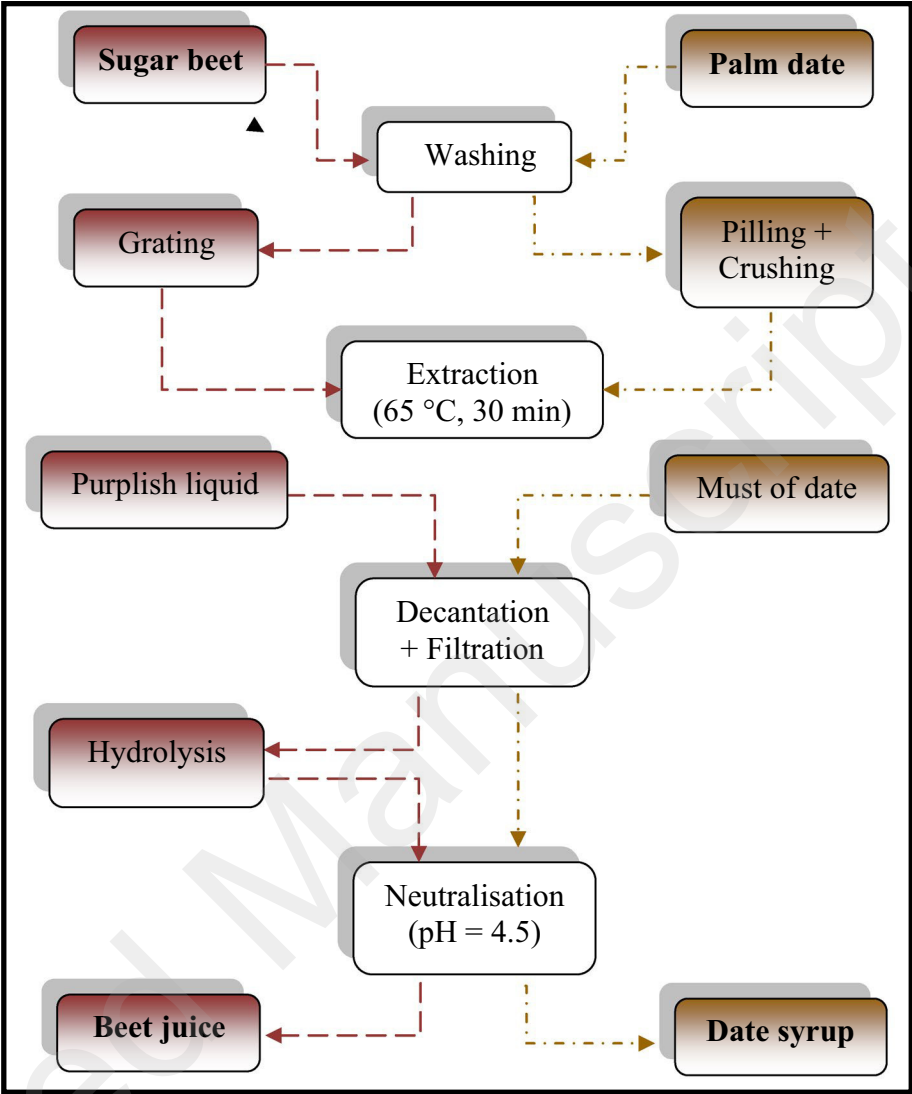
Agricultural residues are rich in organic matter but are unfortunately underutilized and often end up being pollutants of the environment. These by-products can be valorized in order to produce bioethanol which can be used as alone or can be blended with gasoline as transportation fuels [5–7]. Bioethanol can be produced through fermentation of any raw materials as long as it contains sugar [8, 9]. Vegetables and fruits are very rich in carbohydrates. A successful conversion of these carbohydrates is considered

as the most crucial step for bioethanol production. Thus, an efficient pretreatment should increase the content of fermentable sugars (glucose, fructose, galactose, etc.) and will results in a good yield of fermentative bioethanol [6, 10, 11]. Palm date is very rich in sucrose, fructose and glucose. Other simple sugars (oses) were identified in some date varieties; it is arabinose, xylose, galactose and mannose present at very low concentrations [12]. Palm date can therefore be good candidates for the production of bioenergy. In Algeria, one of the leading countries in the production and export of dates in the world, palm date cultivars are numerous and poorly exploited, with the exception of Deglat-Nour, Ghars, Deglat- Beida and Mech-Deglat, which are of major economic importance [12]. Thousands of tons of dates remain unused and can exceed 50% of year-round production, which could be valorized [13, 14]. Sugar beet is also a typical feedstock for bioethanol production which is rich in hemicelluloses (24–32%), and cellulose (22–30%) (It has very low lignin content) [15]. The juice is fermented by yeast or bacteria. The chemical composition of sugar beet reveals a high content of fermentable sugars (arabinose, mannose and xylose) [16]. The pulp, once drained, is used as animal feed or sold to the chemical, pharmaceutical or food industry.

The yeast *Saccharomyces cerevisiae* has long been an efficient agent for ethanol production at laboratory and industrial level with high efficiency [17–19]. It is considered as the world's premier industrial microorganisms being the best exploited microorganism in terms of both old and new biotechnologies [20]. The yeast tolerates a wide range of pH with an optimum under acidic

83	conditions, which makes its fermentation less susceptible	129
84	to infection than bacteria [21]. It also tolerates ethanol and	130
85	inhibitory compounds better than other ethanol produc-	131
86	ing microorganisms. <i>S. cerevisiae</i> is considered as of major	132
87	economic and social significance in human culture since it	133
88	has long been used to produce alcoholic beverages (beer	134
89	and wine) and ferment bread.	
90	The objective of the present project was to propose a	135
91	method of valorization of agricultural products in order	
92	to produce second-generation bioethanol. We opted for	
93	Algerian palm dates and sugar beets of low quality. <i>Sac-</i>	
94	<i>charomyces cerevisiae</i> was the microorganism used to	
95	perform the alcoholic fermentation. The main goal was	
96	to develop an effective pretreatment of the selected raw	
97	material and alternative and efficient process for bioetha-	
98	anol production from a large variety of agro-industrial	
99	wastes.	
100	2 Materials and methods	
101	2.1 Reagents	
102	All the chemicals used in the present work (acids, nutrient	
103	salts, etc.) were purchased from Merck.	
104	2.2 Selection and pretreatment of raw material	
105	(palm date and sugar beet)	
106	The sugar beet, <i>Beta vulgaris</i> from Algeria was used in the	
107	present work. It is a vegetable with a tuberous root, in	
108	which high amounts of sugar accumulate. The palm date	
109	used was Deglet-Nour which is the edible fruit of the date	
110	palm <i>Phoenix dactylifera</i> . It is highly coveted inside and	
111	outside the country because of its flavor and nutritional	
112	quality; it is a fleshy fruit containing an elongated core	
113	marketed mostly in the form of dry dates. The two prod-	
114	ucts used in this work are of lower quality to be taken as	
115	residues of agriculture.	
116	For the preparation of date syrup and beet juice, some	
117	pretreatments were carried out. They differ according to	
118	the nature of the raw material; the protocol was described	
119	in Fig. 1. In the case of sugar beet, two kinds of juice were	
120	prepared: beet juice with liming (BJL) and beet juice with-	
121	out liming. The operation of liming consists of a clarifica-	
122	tion of beet juice by calco-carbonic purification (addition	
123	of CaCO_3) in order to precipitate impurities. To obtain BJL,	
124	40 ml of CaCO_3 (10 g/L) was added to the filtrate. Calcium	
125	carbonate fixes and precipitates impurities. The precipitate	
126	was then removed by a second filtration which led to a	
127	clear beet juice. Hydrolysis and neutralization were then	
128	carried out in the same way as for pretreatment without	
	liming. The extracted date syrups and beet juices were	129
	stored in hermetic flasks at 4 °C until use.	130
	The extraction was operated in a 500-ml flask equipped	131
	with a reflux condenser at a temperature of 65 °C during	132
	30 min. This allowed a better extraction of sugars and a	133
	good consumption of the vegetable pulp.	134
	2.3 Characteristics of <i>Saccharomyces cerevisiae</i>	135
	<i>Saccharomyces cerevisiae</i> selected in the present work	136
	was that marketed in Algeria. It was inoculated in nutri-	137
	ent broth (supplemented with glucose and salts) at 30 °C	138
	under shaking conditions to ensure the growth and activ-	139
	ity conditions of the yeast. Cell density referred to the num-	140
	ber of living cells per unit volume was measured at 600 nm	141
	(see Analytical techniques).	142
	2.4 Alcoholic fermentation experiments	143
	The fermentation assays were carried out in a 1 L-batch	144
	bioreactor under magnetic stirring. The temperature was	145
	maintained at 30 ± 2 °C. A volume of 500 ml of slightly	146
	heated juice or syrup (pH=4.5) was placed in the fer-	147
	menter supplemented with nutrients necessary to pro-	148
	mote rapid cell growth. Two grammes of yeast were then	149
	added to the reactor which must be quickly closed with a	150
	hermetic plug to ensure anaerobic conditions. Fermenta-	151
	tion began once the release of CO_2 was observed. The fer-	152
	mentation time was fixed at 72 h after which the bioetha-	153
	anol was recovered by distillation of the fermented mixture.	154
	The distillation temperature was of 78 °C. It was carried out	155
	in conventional equipment containing a heating bottle,	156
	refrigeration columns and a distillate recovery bulb.	157
	Liquid samples for bioethanol determination were col-	158
	lected at the end of fermentation and filtered by a 0.45 μm	159
	Millipore hydrophilic filter. The amount of CO_2 released	160
	was measured by the liquid displacement method [13]	161
	(Fig. 2).	162
	2.5 Analytical techniques	163
	2.5.1 Characterization of beet juice and date syrup	164
	Juice and syrup produced from raw materials were charac-	165
	terized by some physicochemical and biochemical param-	166
	eters such as pH, mass soluble solids (Brix), dry matter,	167
	total sugars and cell density.	168
	• The Brix, measured by a refractometer, is a measure	169
	of total soluble solids (including sugars). It is based	170
	on the ability of a juice's sugar to deflect light. In the	171
	present work, the Brix of beet juice and date syrup	172

Fig. 1 Pretreatments of sugar beet and palm date for the extraction of beet juice and date syrup



was made by measuring the refractive index at 20 °C using an ATAGO RX-5000 type refractometer.

- The pH which is essential for the control of the substrate before and during fermentation is an indicator of the metabolic activity of the yeast during the transformation of sugars. Its measure was carried out by a pH-meter Inolab pH 7110.
- The dry matter (DM) was determined by drying in an oven at a temperature of 105 °C until a constant weight a sample of 10 ml of juice or syrup. It is expressed as:

$$DM(\%) = \frac{M_1 - M_0}{M_2 - M_0} \times 100$$

M_0 is the mass of the empty capsule (g), M_1 is the mass of the capsule and the residue after desiccation (g), M_2 is the mass of the capsule and the sample (g).

- Total sugars were determined by the phenol–sulfuric acid method based on the absorption of light at 448 nm [22]. A Secomam Prime-type spectrophotometer was used for this purpose.

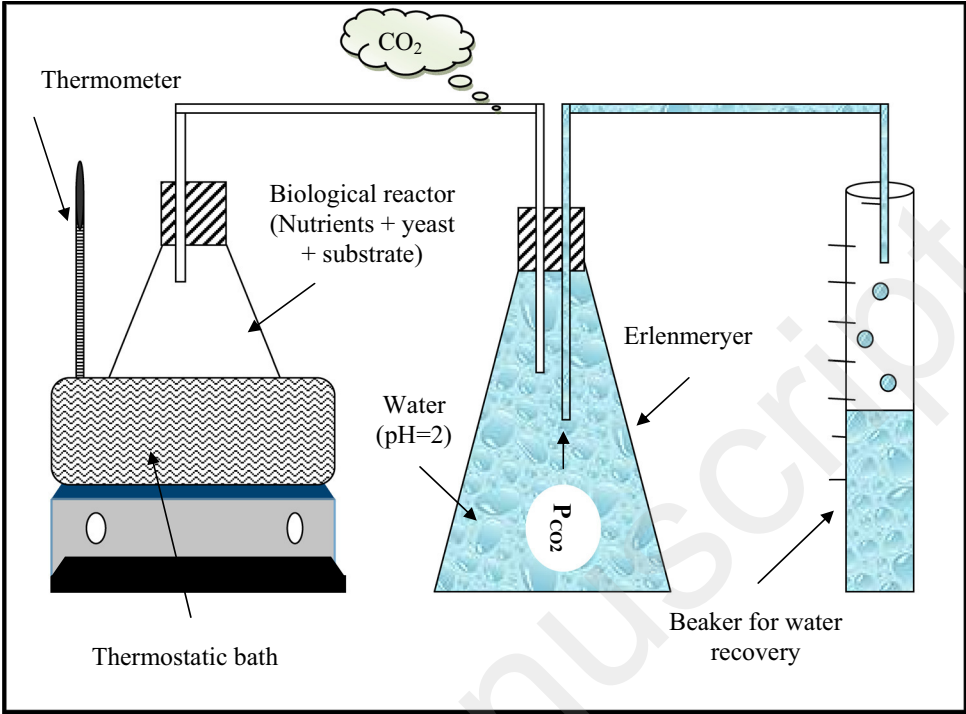
2.5.2 Cell density

Cell density was measured at 600 nm (A600) using a Secomam Prime-type spectrophotometer. A600 values were converted into cell density (10³ kcell/mL) by using a standard curve [23].

2.5.3 Characterization of bioethanol

The fermentative bioethanol was characterized by its physical appearance, smell and flammability. Its density, weight and concentration were also measured. Common techniques were used for this purpose.

Fig. 2 Experimental setup of alcoholic fermentation



2.5.4 Pressure of CO₂

The biological conversion was followed by measuring the CO₂ release resulting from the fermentation following the degradation of sugars; CO₂ production was measured by displacement of water (Fig. 2). The reactor was connected via a tube to an Erlenmeyer flask filled with water at pH 2. The latter was connected by a second tube to an empty container. The CO₂ produced in the reactor enters the flask exerting a pressure that will expel a quantity of water proportional to this pressure. This quantity of water recovered in the second container has a volume equivalent to the CO₂ pressure.

The results were analyzed using a one-way analysis of variance (ANOVA). Comparison between the different treatments was statistically analyzed and the validity of investigation was expressed as probability value of $p < 0.05$.

3 Results and discussion

3.1 Physicochemical proprieties of beet juice and date syrup

Pretreatment is an important step in the conversion of biomass to bioethanol. In this study, an effective acid pretreatment was chosen to treat date and beet in order to convert the biomass into fermentable products. The different proprieties of beet juice and date syrup before conducting fermentation are given in Table 1. The five substrates appeared syrupy and viscous with a density varying between 0.99 and 1.05. Their color differs according to the nature of the raw material. Liming operation led to a beet juice with a lighter red color probably due to the elimination of impurities and some pigments. The pH of concentrated and diluted beet juice was around

Table 1 Physicochemical proprieties of beet juice and date syrup before fermentation

Parameter	Beet juice			Date syrup	
	4.8 °Brix	3.1 °Brix ^a	2.2 °Brix	22.1 °Brix	12 °Brix
pH (20 °C)	4.18	6.66	4.29	6.30	4.10
Density (20 °C)	1.01	1.05	0.99	0.99	0.99
Total sugars (mg/L)	47.14	30.07	17.21	220.1	110.8
Color	Dark red	Very light red	Light red	Dark brown	Light brown

Three independent experiments were performed for each assay. The values are expressed as mean ± SD where $0.01 \leq SD \leq 0.5$

^aBeet juice with liming

4.2 which is an optimal value for the development of *S. cerevisiae*. The beet juice with liming had a pH value close to neutrality due to CaCO_3 supplied during the liming operation. Concentrated beet juice was richer in total sugars than the two others with a value of 47.14 mg/L. The liming operation was probably responsible for the lost of sugar amount from the juice (30.07 mg/L). Concentrated date syrup containing 220.1 mg/L of total sugar is considered as the most sugar-rich raw material used in the present study with a concentration of 220.1 mg/L.

3.2 Fermentation monitoring

Saccharomyces cerevisiae represents the organism of choice for the production of ethanol at laboratory and industrial scale [23]. In this work, the time course of fermentation process was studied using the five substrates as a carbon source during 72 h. The medium was, as described above, adjusted to initial pH of 4.5 and fermented at temperature of 30 ± 2 °C. The Yeasts also require appropriate supplies of nutrients, in order to efficiently carry out fermentation. Typical profiles of CO_2 production were obtained for all the five substrates (Fig. 3). It was possible to define three standardized phases in the curves: (1) a lag phase (of about a few tens

of minutes) during which the yeasts adapt to the process conditions, (2) an exponential phase leading to assess the maximal biomass, and (3) a stationary phase. The highest biomass was reached at 3.0, 3.5 and 3.6 h in the presence of beet juice with 2.2, 4.8 and 3.2 °Brix, respectively (Fig. 3a). The yeast reached its highest biomass in the presence of date syrup (12 °Brix) after 10 h of fermentation (Fig. 3b). The concentrated syrup appeared to be less favorable than the diluted one for the development of yeast; this effect is probably attributed to the osmotic stress resulting from high sugar concentration of concentrated date syrup [23] or caused by some by-products (acids, aldehydes, esters, etc.) and/or non-metabolized components which can inhibit the yeast growth [24]. From these results, it is expected that the diluted date syrup would give a good quality bioethanol since alcohol production and yeast growth are inextricably linked [25].

3.3 Physicochemical and biochemical characterization of fermentative products

The fermentable sugars are metabolized by the yeasts that carry out bioconversion into bioethanol via fermentation. As well known, *S. cerevisiae* conduct fermentative metabolism to ethanol and carbon dioxide (as the primary fermentation metabolites) and also produce numerous secondary metabolites [25]. After alcohol fermentation, it was observed a decrease in the pH for all the substrates used in the resent work. This decrease was due to the formation of the carbon dioxide following the conversion of sugars into ethanol and making the medium more acid. This result was reported in several studies related to fermentation process with yeasts [3, 26]. In a similar trend, total soluble solids (Brix) and total sugars decreased for the five substrates. Total soluble solids decreased by more than 60% in the case of beet juice (4.8 °Brix) and date syrup (12.0 °Brix). The yeast consumed more than 50% sugars of beet after 72 h of fermentation. Total sugars contained in concentrated syrup date were not consumed more than 18% while those contained in the diluted syrup date were converted to more than 63%. The cell density of yeast led to estimate the quality of the fermentation process. The maximum cell density of *S. cerevisiae* in the present fermentation conditions was $8.4 \cdot 10^3$ kcell/mL with 12 °Brix of date syrup while the minimum cell density ($3.6 \cdot 10^3$ kcell/mL) was obtained for concentrated date syrup. This result was confirmed by the formation of dense foam during the fermentation process. Finally, the results summarized in Table 2 show that the diluted syrup date represents the best substrate for the development of *S. cerevisiae* and the conversion of sugars.

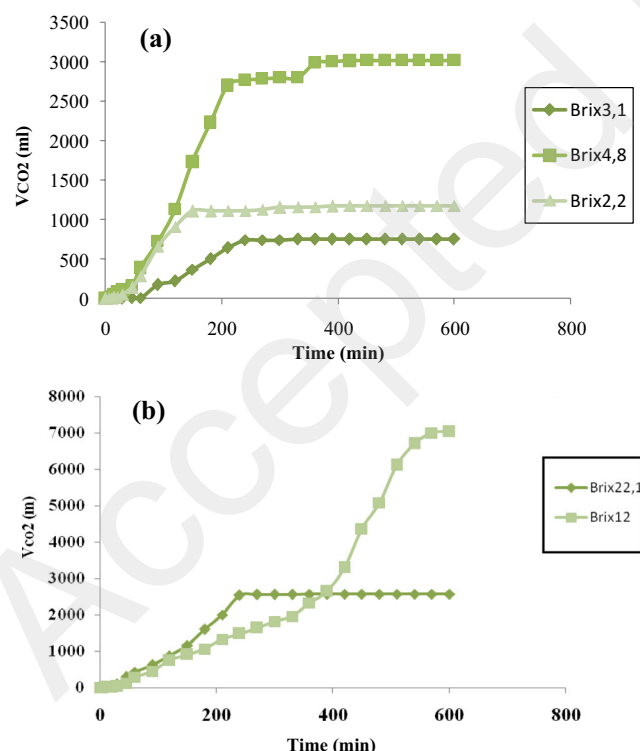


Fig. 3 Time course of fermentation for a beet juice and b date syrup for different Brix values

Table 2 Physicochemical and biochemical characteristics of fermentative products

Parameter	Beet juice			Date syrup	
	4.8 °Brix	3.1 °Brix ^a	2.2 °Brix	22.1 °Brix	12 °Brix
pH (20 °C)	3.54	4.06	3.70	4.70	3.90
Density (20 °C)	0.99	1.04	0.97	0.99	0.96
Total soluble solids (°Brix)	1.8	2.4	1.5	19.5	4.5
Total sugars (mg/L)	21.93	19.57	5.14	180.5	40.5
Cell density (10 ³ kcell/mL)	6.7	5.6	6.0	3.6	8.4
Dry matter (g/L)	6.7	2.5	4.9	21.3	43.5

Three independent experiments were performed for each assay. The values are expressed as mean ± SD where 0.01 ≤ SD ≤ 0.5

^aBeet juice with liming

Table 3 Characterization of bioethanol produced by distillation

Substrate	d (/)	Volume (ml)	Yield ^b (g/kg DM)	Con- centra- tion (g/L)
Beet juice (4.8 °Brix)	0.86	17	28.95	36.5
Beet juice ^a (3.1 °Brix)	0.85	14.5	24.40	30.8
Beet juice (2.2 °Brix)	0.88	8.0	14.22	17.6
Date syrup (22.1 °Brix)	0.86	36.4	63.24	78.2
Date syrup (12 °Brix)	0.88	42.0	74.67	92.4

^aBeet juice: Beet juice with liming

^bThe yield is defined as ratio of bioethanol to the mass of substrate

concentration. Joshi et al. [24] on the other hand, assessed that the low efficiency of ethanol production may be due to an inhibition of the yeast growth, and consequently alcohol fermentation caused by some by-products or non-metabolized components. A flammability test was conducted with this bioethanol; a very intense and sustainable flame was obtained proving its good quality as a fuel. Although alcoholic fermentation with *S. cerevisiae* has long been improved, the present results showed that lower quality of Algerian dates, which cannot be integrated into human nutrition and are useless as animal feed due to low protein content [27] can be valorized for the production of ethanol with good quality that could be used as fuel. On the other hand, sugar beet requires high pretreatments to hydrolyze some complex molecules (starch, cellulose, etc.) into fermentable sugars.

3.4 Bioethanol yield and concentration

The physical and chemical pretreatments performed on beet and date materials were required to reduce recalcitrance biomass for hydrolyzing complex sugars (mainly sucrose) into simple intermediate products and increase the reactive surface area [2–21]. A comparison of the yield and concentration of ethanol of all fermentation processes is presented in Table 3. The values showed that alcohol obtained from the three substrates extracted from sugar beet had low concentration and yield (< 29 g/kg DM) suggesting that this raw material requires high pretreatments to hydrolyze sucrose into fermentable sugars. In parallel, for an initial soluble solids concentration of 12.0 °Brix, 92.4 g/L of bioethanol was produced by *S. cerevisiae* from date syrup. This concentration corresponded to an ethanol yield of 74.67 g/kg DM. In the presence of concentrated date syrup (22.2 °Brix), a lower efficiency of ethanol production was observed (78.2 g/L). These results corroborate with those of Chniti et al. [23] which attributed this effect to the osmotic stress resulting from high sugar

4 Conclusion

Regarding the efficiency of the fermentation, in respect to *S. cerevisiae* growth and ethanol yield, the results seemed to be promising. The ethanol yield was 74.67 g/kg dry matter (DM) of original date material with a concentration of 92.4 g/L. Syrup extracted from Algerian dates of lower quality has the potential to be an industrially useful substrate for producing second-generation bioethanol. However, sugar beet requires high pretreatments to improve biomass digestibility by hydrolyzing complex sugars into fermentable sugars. Finally, to increase even more the productivity of this alcoholic fermentation process, factors inhibiting the bioethanol production should be identified and overcome. Thus, the extrapolation of the present process to the semi-pilot and pilot scale will be possible. Also, the search for other microorganisms capable of accelerating the process of alcoholic fermentation, increasing the conversion rate and producing a good ethanol is strongly encouraged.

Author contributions All authors contributed to the study conception and design. Material preparation, data collection and analysis were performed by Ibtissem FENNOUCHE and Nabila KHELLAF. The first draft of the manuscript was written by Nabila KHELLAF and all authors commented on previous versions of the manuscript. All authors read and approved the final manuscript.

Compliance with ethical standards

Conflict of interest The authors declare that there is no conflict of interest.

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