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Study of the self-association of molecular tweezers bearing two different arms: influence of the stereoelectronic effects of the arm substituents.

Béatrice Legouin,^a Arnaud Bondon,^a Clément Orione,^b Solenn Ferron,^a and Philippe Uriac^a

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In the general field of molecular recognition, the self-assembly of a tweezer bearing a R(+) usnic acid arm and a substituted 1,8-naphthalimide moiety as a second arm was investigated through a ¹H NMR study. Two self-association modes were evidenced: either the usnic acid arms (mode A) or the naphthalimide ones (mode B) were face to face in the center of the dimer. The existence of a third non-symmetrical mode (mode C) has not been evidenced experimentally. The behavior of these tweezers was discussed taking into account the electronic density of the naphthalimide arms and the slow mode A/mode B exchange rate (EXSY experiments). Only the mode A was observed when naphthalimide was either non-substituted (1) or substituted by Br (2). With one or two electrodonating substituents (OCH₃ (3) or (OCH₃)₂ (4) or N(CH₃)₂ (5)) both modes A and B occured in a 50/50 ratio. This ratio depended on temperature for tweezer 3.

Keywords

tweezer; self-association; supramolecular recognition, ¹H NMR

Introduction

Molecular tweezers (or clefts or clips) are preorganized synthetic receptors or hosts that have received increasing attention for the study of molecular recognition. Since the pioneering work reported by Chen and Whitlock in 1978 [1] numerous syntheses of molecular tweezers have been described in the literature [2–7]. In most of the publications the charge transfers between the tweezer and various guests (electron-poor aromatics, amino-acids, ions....) were described and characterized. More recently the interactions of tweezers with proteins or nucleic acids were even reported [8–11]. Additional papers explored their self-assembly (or self-association or dimerization) using ¹H NMR spectroscopy and X-ray diffraction, often associated with computational studies [12–19].

In a previous study, we have described the synthesis of molecular tweezers bearing two different arms and their association with the 2,4,7-trinitrofluoren-9-one (**TNF**) guest [20]. Then, we decided to investigate the self-assembly of five related tweezers (**1-5**) (Fig1). As a first common feature these compounds were bearing a R(+) usnic acid arm and as a second one various substituted 1,8-naphthalimide moities in order to evaluate the substituents influence on their self-assembly. Tweezers **6** and **7** [21, 22] (Fig1) were included as references. Our study was essentially performed by ¹H NMR in CD₂Cl₂. Then the association could have been follow through the anisotropic effect variations on the aromatic protons. Some previous results of X-ray diffraction [20–22] allowed discussion.

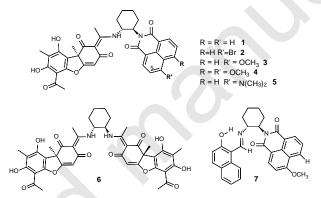


Fig1 Structure of the molecular tweezers 1-7

During the self-assembly process, two symmetric modes could occur: either the usnic acid moieties or the naphthalimide arms were face to face at the center of the dimeric structure, as illustrated by the mode **A** and the mode **B**, respectively. The characteristics of the self-assembly modes **A** and **B** will be first described with **1** and the reference compounds **6** and **7**. Then, the behavior of the four tweezers **2-5** will be reported. The weight of each self-assembly species depended on the nature of the substituents **R** and **R'**. A third non-symmetric association mode (mode **C**) was also worth considering, with both a naphthalimide and an usnic acid arms at the center of the dimer (Fig2).

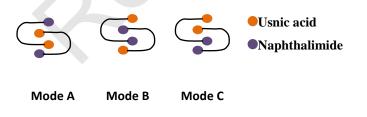


Fig2 Different self-assembly association modes

Results and discussion

The ¹H NMR signals of H-5 and CH₃ signal of the groups R and R' on the naphthalimide arm as well as the OH- β signal on the usnic arm (Fig3) were selected. Indeed, these three signals, unambiguously attributed and well isolated on the spectra, were corresponding to protons included in the maximum interference area (Fig4). Their shift variations, observed when the tweezers concentration increased allowed the self-assembly study. After preliminary experiments performed to check spectra resolution and shift variation amplitude, the 258 K temperature has been chosen (whole spectra are given in Electronic Supplementary Material - ESM).

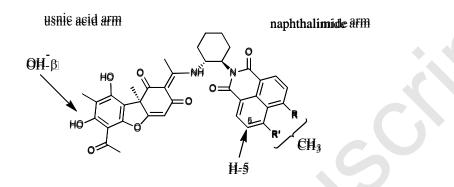


Fig3 Selected ¹H NMR signals

Description of the self-assembly modes A and B

As illustrated on X-ray diffraction (Fig4), tweezer **1** was auto-associated through the mode **A** where the usnic acid units are face to face, like the compound **6** [21, 22]. The ¹H NMR data of the aromatic proton H-5 on the naphthalimide underwent low chemical shift variation $\Delta \delta = 0.04$ ppm when increasing the concentration from 1.3 mM to 80.0 mM at 258 K, attesting the weak effect of the usnic acid arm on the external naphthalimide one. The reciprocal effect on OH- β was weaker for **1** than for **6** ($\Delta \delta = 0.05$ ppm *versus* $\Delta \delta = 0.21$ ppm).

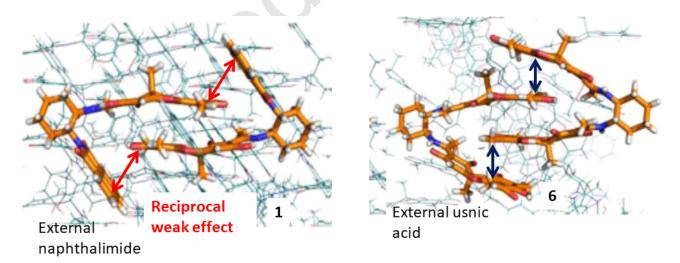


Fig4 X-ray structures of 1 and 6. Arms reciprocal effects.

Mode **B** could be evidenced by tweezer **7**. We had previously observed [23] that no charge transfer with TNF occurred with one naphthalene arm derived of the 2-hydroxynaphthaldehyde (see Supplementary Material), suggesting that if a chemical shift variation was observed for the current tweezer **7**, this meant that the naphthalimide moieties interacted with themselves within a mode **B**. Contrary to mode **A**, the methoxynaphthalimide arm protons (H-5 and H-7) were significantly shielded, exhibiting shift values of 0.22 and 0.25 ppm respectively (Fig5). It could be noted that the naphthalene arm (H-4' and N=CH) remained unaffected according to the shielding value of N=CH and H-4' of 0.03 ppm. Moreover, one can observe two sets of signals corresponding to α and β conformers (Fig6) at low temperatures. This phenomenon was induced by the non-symmetrical arms of **7** and has been widely previously discussed [20].

These results validated the selection of H-5 as self-assembly marker: its inclusion into the tweezer resulted in a large shielding (above 0.25 ppm) evidencing the mode **B** while an external position lead to a weaker shielding indicating the mode **A**, as already observed with naphthalene derivatives by Klärner [24] in D_2O . We will focus mainly on this proton for the further study of compounds 2-5.

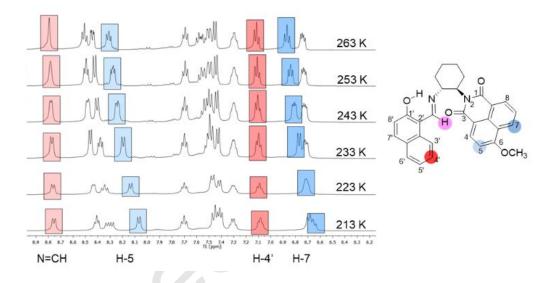


Fig5 Partial ¹H NMR spectra of 7 depending on temperature

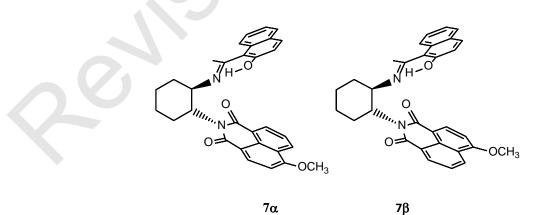


Fig6 Structures of 7α and 7β conformers

Stereoelectronic effects: tweezers 2-5

Withdrawing substituted arm tweezer 2

In the concentration range encompassing 1.1 to 69.4 mM at 258 K, tweezer 2 exhibited the same behaviour as compound 1 with a chemical shielding $\Delta \delta = 0.12$ ppm for H-5 (see Supplementary Material). One can assume than self-assembly occured according to mode **A**. A little stronger reciprocal effect on usnic arm ($\Delta \delta_{OH-\beta} = 0.11$ ppm versus 0.05 ppm) could be explained by the higher electronic density due to the Br substituent.

Electrodonating substituted arms tweezers 3, 4 and 5

At first, considering the partial ¹H NMR spectra of **3** at 258 K (Fig7), one can observe the splitting of the aromatic proton H-5. A triplet was observed at 7.02 ppm at 1.1 mM which split with different amplitude in two signals to reach 6.93 ppm and 6.35 ppm at 72.6 mM, meaning shieldings of 0.09 and 0.67 ppm respectively. The same observations can be done for the OCH₃ signal. A singlet peak was still observed at 4.5 mM (4.06 ppm) but split at 9.1 mM to give 2 singlets which differently shift when concentration increases. At 72.6 mM, signals were recorded at 3.98 ppm and 3.57 ppm leading respectively to 0.08 and 0.49 ppm shieldings. Compounds **4** and **5** exhibited the same behaviors when studying at the different concentrations. These behaviors let us to conclude than self-assembly occured according to both modes **A** and **B** for tweezers **3-5**.

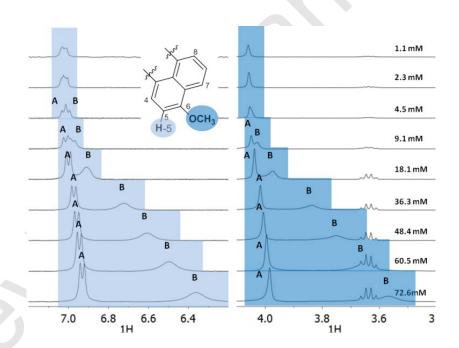


Fig7 Details of the ¹H NMR spectra of 3: signals of OCH₃ and H-5 at 258 K

Concerning the splitting of the methoxy group signal, we could note that the singlets kept the same integration with a 50/50 ratio suggesting equal affinity constants K_A and K_B (Fig6). On the two new sets of signals (Fig7) a signal broadening was observed, consistent with a relatively fast exchange in the chemical shift timescale between the self-assembled and the free monomeric form. According to it, an apparent interconversion between both forms was observed on EXSY spectra that

evidenced the high dissociation-association-rate constants for both self-assembly modes **A** and **B** (Fig8). **3** EXSY spectrum is presented Figure 9. The magnetization exchange rate constants k'_1 and k'_{-1} were 4.3 ± 0.2 s⁻¹ for compound **3** at 278 K and 72.5 mM in agreement with the low affinity constants values.

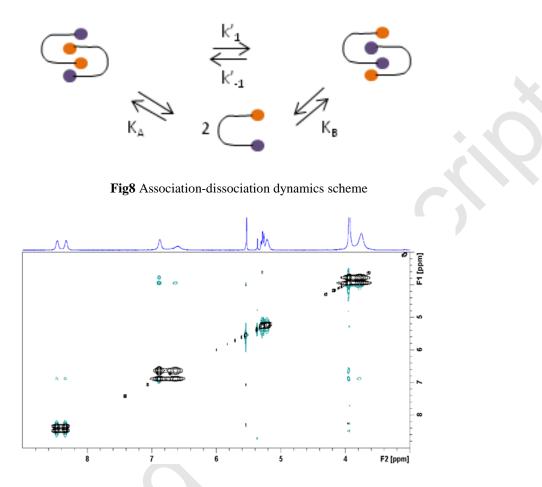


Fig9 EXSY spectrum of 3. Black cross peaks denote chemical exchange and blue ones denote NOEs.

On Figure 10, the shielding values of both ortho protons (H-5) and X-CH₃ protons, measured in the concentration range 1-70 mM for tweezers **3-5**, are presented for both self-assembly modes **A** and **B**. Concerning the self-assembly mode **A**, the reciprocal effect of the usnic arm on H-5 and CH₃ (R,R') remained low: $\Delta\delta < 0.1$ ppm for **3** and **5** in good agreement with effects observed with **1** and **2**. The disubstituted tweezer **4** exhibited slightly higher values ($\Delta\delta_{H-5} = 0.25$ ppm, $\Delta\delta_{OMe} = 0.15$ ppm) but still consistent with the mode **A**.

In the self-assembly mode **B**, chemical shift variations of the naphthalimide protons dramatically differed for **3** and **4**. They reached $\Delta \delta_{\text{H-5}} \approx 0.7$ and $\Delta \delta_{\text{OMe}} = 0.5$ ppm at about 70 mM (only one signal was observed for both methoxy groups). The chemical shift variation was reduced for **5** (<0.10 ppm for both signals).

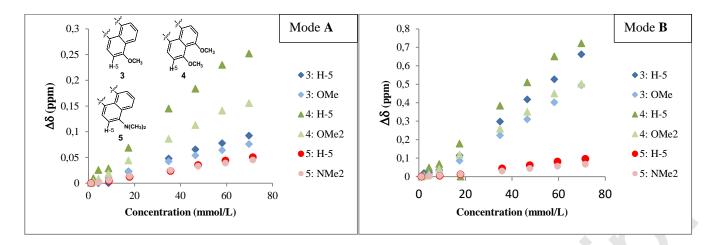


Fig10 Shielding of some protons of **3**, **4** and **5** at 258 K according to mode A (left) and mode B (right) (numbering is given on the structures)

In the literature [25, 26] the importance of electrostatic interaction between static molecular charge distributions to explain non-covalent interaction and molecular recognition events has been highlighted. Thus, in table 1 calculated electronic potential values (see Supplementary Material) are reported. When core charge is negative (1, 2), only mode A was observed whereas both modes A and B exist when it becomes positive (3-5). In mode B, in addition of the electronic potential values, the observed chemical shift variation of 4 (table 1) was modulated by the steric hindrance between the two methoxy substituents. The weak shielding observed for 5 was related to an ineffective delocalization of the lone pair electrons of nitrogen owing to its pseudo equatorial position [27] and to the steric hindrance. In the case of 3 and 4, the high shielding of the CH₃ groups ($\Delta\delta$ close to 0.70 ppm) gave evidence of their position and of their closeness with the naphthalimide aromatic rings [28]. The resulting CH/ π interactions were also in favor of a charge transfer between two naphthalimide arms [29].

	1	2	3	4	5
Core	-0.22	-0.12	0.08	0.31	0.03
charge					
		6387	85	283	-28-

 Table 1 Electrodonating substituents impact on naphthalimide arm: calculated core charge and electrostatic potential.

 (Negative and positive potentials are respectively red and blue).

Existence of mode C?

Mode C was characterised by an acid usnic and a naphthalimide moiety in the center of the tweezer (Fig2). Such an interaction should lead to four non-equivalent magnetical moieties. ¹H NMR spectra in accordance with hypothesis were never observed meaning that the lifetime of such species was not large enough to permit detection.

Temperature influence on self-association

In order to better understand the self-assembly process, the temperature influence on the mode A/mode B ratio was explored using a concentrated solution (about 70 mM) for 3 and 4 which presented the highest shielding values. If the ratio remained constant at 50/50 for 4, it reached 35/65 for 3 at 238 K. As expected, a temperature decreasing induced a $\Delta\delta$ increasing for both tweezers.

In the Figure 11 were reported the shielding variations $\Delta\delta$ ($\Delta\delta = \delta_{\rm T} - \delta_{298}$) of the OCH₃ signals (at 400 MHz) for the two modes (See spectra in Supplementary Material). We could note that in the mode **B** at 238 K, the shielding effect was nearly twice as high for **4** than for **3** ($\Delta\delta = 1.10$ ppm and 0.58 ppm, respectively).

As previously described for **2** and **7**, two favored conformers α and β existed due to the non-symmetry of either usnic acid (**2**) or naphthalene (**7**). The equilibrium rate decreased when temperature decreased leading to two distinguishable populations α and β at 238 K. The $\alpha\alpha$, $\beta\beta$ and $\alpha\beta$ associations were identical for **4** but they clearly differed for **3** for which *anti* associations ($\alpha\beta$) were favored over the *syn* ones ($\alpha\alpha$ or $\beta\beta$) in which the two methoxy interfere (Fig12). This behavior could explain the ratio between modes **A** and **B** which reached 35:65 at 238 K while it is 50:50 at higher temperatures.

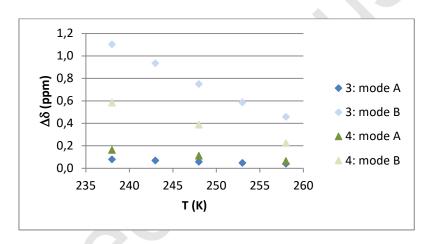


Fig11 $\Delta\delta$ ($\delta_T - \delta_{298}$) in ppm for OCH₃ signal vs temperature in K for modes A and B for compounds 3 and 4

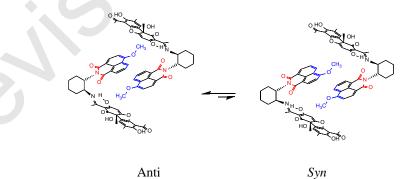


Fig12 Syn and anti associations of 3

Conclusion

This second contribution devoted to the design and the synthesis of tweezers possessing two different arms confirms the interest of such non-symmetrical molecules for studying molecular recognition phenomena. The self-association can occurred according to two different modes. The substituents stereoelectronical effects on the charge transfer and then on the self-association modes was highlighted like in the previous paper concerning the tweezer interaction with the **TNF** [20]. Thus, little modifications can induce a strong change in the recognition. Such observations are extremely frequent in medicinal chemistry [30, 31]. The design and the study of non-symmetrical tweezers with moderate affinity constant *K* could be helpful for the drug design but also for mechanistic propositions in chemistry [32].

Experimental Section

Materials and methods

All reagents of high quality were purchased from commercial suppliers and used without further purification. ¹H and ¹³C NMR spectroscopic data were recorded on Bruker apparatus at either 300, 400 or 500 MHz and either 75, 100 or 125 MHz, respectively. ¹H NMR titration experiments were performed at 500 MHz. The undeuterated residue of the solvent was used as internal standard. The δ values are given in ppm and the coupling constants in Hz. The multiplicity of the signals is reported as s (singlet), d (doublet), t(triplet), q (quadruplet) and m (multiplet). As previously reported, ¹³C signals could be either coalescent (signals are indicated by the symbol *) or split in two signals. HRMS analyses are obtained using an Orbitrap instrument for ESI. Melting points are uncorrected. Thin layer chromatography was performed using precoated Merck silica gel plate 60 F254 (0.2 mm thickness) and the Rf values are given for the eluent used during purification. Column chromatography was done using silica gel 0.063-0.2 mm. The chromatography solvents were eliminated from the tweezers by using vacuum pomp (10⁻² mm Hg) and heating (CH₂Cl₂, CH₃CO₂Et, C₂H₅OH) but traces are present in some spectra.

The ¹H NMR self-assembly studies were performed at 400 MHz using CD_2Cl_2 as solvent and all spectra are given in SM. The ¹H NMR spectra were recorded on a Bruker AVANCE III 400 MHz Ultrashield Plus spectrometer operating at 400.13 MHz for ¹H observation. A 5 mm Bruker PABBO BB-1H/D Z-GRD probehead was used for the ¹H analysis. NMR experiments were acquired using zg30 Bruker sequences supplied in TopSpin 2.1, processed and analyzed in MestReNova 10.0.0 (Mestrelab Research S.L). The temperature of each experiment was controlled by a VTU Unit with a thermocouple. T. For the 1D ¹H NMR spectra, a 1 second relaxation delay and a 5.452 seconds acquisition time were used.

Electrostatic potential has been calculated with Jaguar, as implemented in Maestro v.11.5 (Schrödinger). The surface potential was determined after geometry optimization of each molecule using B3LYP functional and $6-31G^{**}$ basis set (SCF convergence criterion of 5.10^{-5} Ha).

EXSY NMR spectra were recorded at 278 K on a Bruker Avance 500 spectrometer equipped with a BBO probe using 5 mm sample tubes and deuterated dichloromethane as solvent. The spectra were recorded in the phase-sensitive mode using standard Bruker sequences and 8 transients for each *t*1 value. Spectral windows were of 5000 Hz in both dimensions with 4k x 256 data points, mixing times τm of 3 μs and 0.5 s were used, while keeping a constant repetition time of 1.9 s. Before the Fourier transformation, the data were processed with a sine-bell function in both *F*1 and *F*2 domains, and zero filling was applied to obtain a 2k x1k transformed matrix. Peak volumes were integrated using Bruker NMR software TOPSPIN. The calculations of the exchange rates were performed using the EXSYCalc software [33].

Synthesis of **9**

1,8-Dimethoxynaphthenedione can occur according to S.Lahore [34]

4,6-Dimethoxy-1,8-naphthalimide 9 [35] 285 mg (1.18 mmol) of 1,8-Dimethoxynaphthenedione and 1.07 g of oxone (7.06 mmol; 6 eq) were suspended in 20 mL of methanol and refluxed for 96 h. After cooling the mixture was poured into water and the obtained solid was washed using methanol and dried under vacuum. 310 mg of a very insoluble solid was obtained and used without any purification (estimated purity 90%). <u>**HRMS(ESI)**</u>: $(M+H)^+ C_{14}H_{11}O_5$ Calc. 259.06065 Found. 259.0606. <u>**HRMR**</u> (CDCl₃, 300 MHz, 298 K): 8.64 (d, 2H), 7.15 (d,2H), 4.15 (s, 6H).

Synthesis of 11 [21]

270 mg of **9** (1.05 mmol) and 360 mg of 1,2-diaminocyclohexane **8** (3 eq) in 15 mL of ethanol were refluxed for 48 h. The solvent was evaporated under vacuum and **11** was purified by chromatography on silica gel using CH₂Cl₂:MeOH 90:10 as eluent (blue fluorescence). Only 100 mg (Yield 29%) of a yellow-brown solid were obtained probably because of the very bad solubility of **9**. **Rf** 0.56 **F**°**C**: 244-246; **HRMS(ESI)**: $(M+H)^+C_{20}H_{23}N_2O_4$ calc.: 355.16578 found.: 355.1659. ¹H NMR (CDCl₃, 300 MHz, 298K): 8.53 (d 2H), 7.00 (d, J=9Hz, 2H), 4.75 (td, J=3.7 and 11Hz, 1H), 3.78 ((td, J=3.7 and 11Hz, 1H), 4.07 (s, 6H), 2.51 (qd, J=3.7 and 12Hz, 1H), 2.08 (dm, J=11Hz, 1H), 1.70-1.85 (m, 3H), 1.45 (m, 2H), 1.25 (m, 1H) NH₂ not attributed. ¹³C NMR (CD₂Cl₂, 75.4 MHz, 298 K): 165.6, 164.5, 162.9, 134.5, 133.7, 132.3, 115.5, 114.8, 106.3, 60.8, 56.4, 50.4, 37.0, 28.6, 26.2, 25.5.

Synthesis of tweezers 4 and 7 (Scheme 1)

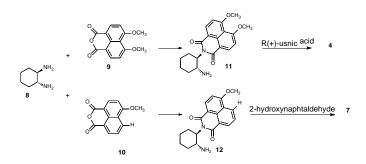
The synthesis strategy, previously reported for tweezers 1, 2, 3 and 5[20–22], was employed to prepare 4 and 7 (scheme 1) from the RR (+) diaminocyclohexane 8. The naphthalimides were introduced in the first step and then the (+)-usnic acid and the 2-hydroxynaphthaldehyde led us to obtain the tweezers 4 and 7 respectively.

To 175 mg of **10** (0.54 mmol) in 10 mL of C₂H₅OH were added 93 mg of 2-hydroxynaphthaldhyde (0.54 mmol). After 48h of refluxing, the solvent is removed under vacuum and **7** was purified by chromatography on silica gel using CH₂Cl₂/CH₃OH-99/1 as eluent. 76 mg of **7** were obtained as a brown oil giving a solid after 2 weeks. Yield: 16% (not optimized). <u>**Rf**</u>: 0.4. <u>**HRMS(ESI)**</u>: (M+H)⁺ C₃₀H₂₇N₂O₄ Calc: 479.19708 Found: 479.1974. <u>¹H NMR</u> (CDCl₃, 300 MHz, 298 K): 14.31 (broad s, 1H), 8.74 (s, 1H), 8.55 (dd, J=1.2 Hz and J=7.3 Hz, 1H), 8.50 (d, J=8.3 Hz, 1H), 8.43 (d, J=8.3 Hz, 1H), 7.62 (m, 2H), 7.50 (d, J=9.3 Hz, 1H), 7.45 (dd, J=1.2 Hz and J=8.0 Hz, 1H), 7.26 (m, 1H), 7.10 (m, 1H), 6.94 (d, J=8.2 Hz, 1H), 6.78 (d, J=9.2 Hz, 1H), 5.32 (m, 1H), 4.72 (td, J=3.9 Hz and J=10.8 Hz, 1H), 4.05 (s, 3H), 2.60 (m, 1H), 2.19 (d, 1H), 1.48-2.03 (m, 6H) + water; <u>¹³C NMR</u> (CDCl₃, 75.4 MHz, 298 K): 174.3, 165.4, 164.9, 164.4, 164.1, 160.9, 157.3, 136.4, 133.6, 129.6, 129.0, 128.8, 127.7, 126.2, 126.0, 124.3, 123.4, 122.5, 118.1, 106.9, 105.3, 63.0, 57.2, 56.2, 35.2, 28.5, 25.8, 24.9.

Synthesis of 4 [21]

100 mg of **11** (0.28 mmol) and 95 mg of usnic acid (1 eq) in 7 mL of ethanol were refluxed for 4h. The solvent was evaporated under vacuum and **4** was purified by chromatography on silica gel using CH_2Cl_2 :MeOH 99:1 as eluent (blue fluorescence). 170 mg of a yellow solid were obtained (yield: 89 %). **<u>Rf</u>** 0.65 <u>**HRMS(ESI)**</u>: $(M+Na)^+ C_{38}H_{36}N_2O_{10}Na$ calc.: 703.22622 found: 703.226. <u>¹H NMR</u> (CDCl₃, 300 MHz, 298 K): 13.30 (s, 2H), 11.82 (s*, 1H), 8.57 (s*, 1H), 8.48 (s*, 1H), 6.99 (d, J=8.7 Hz, 2H), 5.60 (s, 1H), 5.24 (td, J=3.6 and 11.0 Hz, 1H), 5.11 (qd, J=3.6 and 10.0 Hz, 1H), 4.04 (s, 6H), 2.59 (s, 3H), 2.58 (m, 1H), 2,53 (s,

3H), 2.17 (m, 1H), 2.03 (s, 3H), 1.92 (m, 3H), 1.50-1.80 (m,3H), 1.44 (s, 3H). ¹³C NMR (CDCl₃, 75.4 MHz, 298 K): 200.8, 198.1, 174.5, 173.6, 164.9*, 163.5, 163.4, 158.4, 156.0, 135.3, 134.1, 132.5, 114.3, 107.9, 106.6, 105.2, 102.7, 102.1, 101.3, 56.8, 56.6, 56.0, 53.8, 34.0, 31.8, 31.4, 28.5, 25.6, 24.7, 18.1, 7.6.



Scheme 1 Synthesis strategy to prepare compounds 4 and 7

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