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Title:

**Organic chemicals from diesel exhaust particles affects intracellular calcium, inflammation and  $\beta$ -adrenoceptors in endothelial cells**

Running title:

**DEP-OC affects calcium, inflammation and  $\beta$ ARs in endothelial cells**

Authors:

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## Highlights:

- Organic matter from DEP extracted by *n*-hexane (lipophilic) and dichloromethane (semi-lipophilic) enhanced  $[Ca^{2+}]_i$  and expression of inflammation-associated genes in HMEC-1 endothelial cells.
- Semi-lipophilic organic matter from DEP enhanced  $[Ca^{2+}]_i$  via  $\beta$ -adrenoceptors and protease activated receptor 2
- Effects of semi-lipophilic organic matter from DEP on COX-2 expression was partly dependent on enhanced  $[Ca^{2+}]_i$  and  $\beta$ -adrenoceptors.

## Abstract

Exposure to diesel exhaust particles (DEP) may contribute to endothelial dysfunction and cardiovascular disease. DEP, extractable organic material from DEP (DEP-EOM) and certain PAHs seem to trigger  $[Ca^{2+}]_i$  increase as well as inflammation *via* GPCRs like  $\beta$ ARs and PAR-2. In the present study we explored the involvement of  $\beta$ ARs and PAR-2 in effects of DEP-EOM on  $[Ca^{2+}]_i$  and expression of inflammation-associated genes in the endothelial cell-line HMEC-1. We exposed the human microvascular endothelial cell line HMEC-1 to DEP-EOM fractionated by sequential extraction with solvents of increasing polarity: *n*-hexane (*n*-Hex-EOM), dichloromethane (DCM-EOM), methanol (Methanol-EOM) and water (Water-EOM). While Methanol-EOM and Water-EOM had no marked effects, *n*-Hex-EOM and DCM-EOM enhanced  $[Ca^{2+}]_i$  (2-3 times baseline) and expression of inflammation-associated genes (IL-1 $\alpha$ , IL-1 $\beta$ , COX-2 and CXCL8; 2-15 times baseline) in HMEC-1. The expression of  $\beta$ ARs (60-80% of baseline) and  $\beta$ AR-inhibitor carazolol suppressed the increase in  $[Ca^{2+}]_i$  induced by both *n*-Hex- and DCM-EOM. Carazolol as well as the  $Ca^{2+}$ -channel inhibitor SKF-96365 reduced the DCM-EOM-induced pro-inflammatory gene-expression. Overexpression of  $\beta$ ARs increased DCM-EOM-induced  $[Ca^{2+}]_i$  responses in HEK293 cells, while  $\beta$ AR-overexpression suppressed  $[Ca^{2+}]_i$  responses from *n*-Hex-EOM. Furthermore, the PAR-2-inhibitor ENMD-1068 attenuated  $[Ca^{2+}]_i$  responses to DCM-EOM, but not *n*-Hex-EOM in HMEC-1.

The results suggest that  $\beta$ AR and PAR-2 are partially involved in effects of complex mixtures of chemicals extracted from DEP on calcium signalling and inflammation-associated genes in the HMEC-1 endothelial cell-line.

**Abbreviations:** aryl hydrocarbon receptor (AhR),  $\beta$ -adrenoceptors ( $\beta$ AR), benzo[*a*]pyrene (B[*a*]P), calcium ( $\text{Ca}^{2+}$ ), cardiovascular disease (CVD), DEP extracted by: *n*-hexane (*n*-Hex-EOM), dichloromethane (DCM-EOM), methanol (Methanol-EOM), water at 25 °C (Water-EOM), diesel exhaust particles (DEP), endothelial nitric oxide synthase (eNOS), extractable organic material of DEP (DEP-EOM), human microvascular endothelial cell-line (HMEC-1), human embryonic kidney cells (HEK293), wild type (WT), intracellular calcium concentrations ( $[\text{Ca}^{2+}]_i$ ), inositol trisphosphate (IP3), G-protein coupled receptors (GPCR), particulate matter (PM), protease-activated receptor-2 (PAR-2), polycyclic aromatic hydrocarbons (PAHs), nuclear factor- $\kappa$ B (NF- $\kappa$ B), 1-nitro-pyrene (1-NP), dimethyl sulfoxide (DMSO), cyclooxygenase 2 (COX-2), interleukin 8 (CXCL8), matrix metalloproteinase 1 (MMP-1)

**Keywords:** diesel exhaust particles, extractable organic material, endothelial dysfunction, calcium signalling, beta adrenoceptors, protease activated receptor.

## 1. Introduction

Air pollution especially particulate matter (PM), is one of the leading environmental causes of cardiovascular disease (CVD) (Brook et al., 2010; HEI, 2017). PM seems to contribute to CVD and progression of atherosclerosis *via* endothelial dysfunction (Donaldson et al., 2001; Moller et al., 2011), defined as an alteration of endothelial cells towards a pro-inflammatory and pro-constrictive phenotype (Dharmashankar and Widlansky, 2010; Ramji and Davies, 2015). However, the precise molecular mechanisms involved in this process are still largely unknown.

Diesel exhaust particles (DEP) are major constituents of urban PM and contain complex mixtures of organic chemicals adhered to the surfaces of carbon cores (Cohen A. J. et al., 2004; Grahame et al., 2014; Maricq, 2007). Many biologic effects of DEP have been attributed to soluble organic chemicals such as polycyclic aromatic hydrocarbons (PAHs), quinones and dioxins as well as modified PAHs such as nitro-PAHs (Bonvallot et al., 2001; Brinchmann et al., 2018a; Brinchmann et al., 2018b; Kawasaki et al., 2001; Keebaugh et al., 2015; Totlandsdal et al., 2012). Lipophilic compounds such as PAHs may rapidly diffuse across the alveolar-capillary barrier into the bloodstream and target the vasculature (Brinchmann et al., 2018b; Gerde, 2001; Penn et al., 2005).

Beta-adrenoceptors ( $\beta$ ARs) are G-protein-coupled receptors (GPCRs) that transmit signalling from the catecholamine hormones adrenaline and noradrenaline acting as regulators of stress responses (Santos and Spadari-Bratfisch, 2006). While  $\beta$ 1- and  $\beta$ 2ARs are expressed in the lung, heart, vasculature and peripheral tissues,  $\beta$ 3-ADRs are mainly expressed in adipose tissue (Bylund et al., 1994; Lowell and Flier, 1997). The overall role of  $\beta$ AR signalling is to regulate cardiopulmonary function and immune responses, and these receptors are thus among the main drug-targets in CVD treatment (De Backer, 2003; Kolmus et al., 2015; Wachter and Gilbert, 2012). PAHs such as pyrene, benzo[*a*]pyrene (B[*a*]P), benzo[*e*]pyrene and chrysene, as well as 1-nitropyrene (1-NP) known to be present in DEP increase intracellular calcium concentration  $[Ca^{2+}]_i$  in the human bronchial epithelial cell-line (BEAS-2B) and human microvascular endothelial cell-line (HMEC-1) (Mayati et al., 2014; Mayati et al., 2012a; Mayati et al., 2012b).  $\beta$ 2ARs were involved in effects on  $[Ca^{2+}]_i$  triggered by B[*a*]P and 1-NP (Mayati et al., 2014; Mayati et al., 2012b). Furthermore,  $[Ca^{2+}]_i$  in endothelial cells regulates blood pressure and flow *via* endothelial nitric oxide synthase (eNOS) and more directly via  $Ca^{2+}$ -regulated  $K^+$ -channels and myoendothelial microdomains (Moller et al., 2011; Sandow et al., 2009; Sandow et al., 2012). Interestingly, diesel

exhaust exposure seem to impair calcium-dependent vasomotor function in healthy men (Barath et al., 2010; Lucking et al., 2011); an effect that may relate to disturbed endothelial  $[Ca^{2+}]_i$ .

Inflammatory effects of organic chemicals known to be present on DEP, seem to depend on increased  $[Ca^{2+}]_i$  (Mayati et al., 2014; Monteiro et al., 2008; N'Diaye et al., 2006; Zhao et al., 1996). We have previously shown that  $\beta$ ARs may be involved in  $[Ca^{2+}]_i$  increase and induction of the pro-inflammatory chemokine CXCL8 in human bronchial epithelial BEAS-2B cells exposed to 1-NP, one of the dominating nitro-PAHs in DEP (Mayati et al., 2014). Other GPCRs have also been implicated in DEP-induced  $Ca^{2+}$  signaling and inflammation. Extractable organic material of DEP (DEP-EOM) increased  $[Ca^{2+}]_i$  *via* protease activated receptor 2 (PAR-2) in primary human bronchial cells (Li et al., 2011). Furthermore, DEP-induced inflammatory responses in BEAS-2B, HMEC-1 and primary endothelial cells seemed at least partly dependent on PAR-2 (Bach et al., 2015; Brinchmann et al., 2018b). PAR activation may promote conversion of endothelial cells into a pro-inflammatory phenotype. In conditions with endothelial dysfunction, PARs also mediate contraction and may thus contribute to atherosclerosis and hypertension (Alberelli and De Candia, 2014).

DEP and DEP-EOM have been shown to induce calcium signalling, and pro-inflammatory responses in endothelial cells (Brinchmann et al., 2018a; Brinchmann et al., 2018b; Lawal et al., 2015; Yin et al., 2013). Furthermore, GPCRs seem to be involved in calcium responses and activation of pro-inflammatory responses induced by organic chemicals known to be presents in DEP (Brinchmann et al., 2018a; Brinchmann et al., 2018b; Li et al., 2011; Mayati et al., 2014; Mayati et al., 2012b). Based on this, we explored the hypothesis that  $\beta$ ARs and PAR-2, are involved in effects of DEP-EOM on  $[Ca^{2+}]_i$  and expression of inflammation-associated genes in the endothelial cell-line HMEC-1.

## 2. Materials and Methods

### 2.1. Chemicals

Dimethyl sulfoxide (DMSO) and hydrocortisone purchased from Sigma-Aldrich (St. Louis, MO). L-Glutamine (200 mM) purchased from Thermo Fischer Scientific (Scotland); endothelial growth factor from Nerliens Meszansky (Oslo, Norway); penicillin and streptomycin from Lonza (Walkersville, MD, USA); MCDB 131 medium was provided by Life technologies

(NY, USA); fetal calf serum (FCS) from Biochrom AG (Berlin, Germany). Pluronic acid and fura-2 acetoxymethyl ester (Fura-2-AM) purchased from Invitrogen. Carazolol, 2'-5'-dideoxyadenosine (dd-Ado) and ICI-118,551 from Sigma Aldrich. 1-[2-(4-Methoxyphenyl)-2-[3-(4-methoxyphenyl)propoxy]ethyl-1H-imidazole hydrochloride (SKF 96365) purchased from TOCRIS (Bristol, UK).

## 2.2. Diesel exhaust particles, chemical extraction and analysis

*Diesel exhaust particles (DEP)* collected from the tail-pipe of a diesel engine (Deutz, 4 cylinder, 2.2 l, 500 rpm) running on gas oil ; were kindly provided by Flemming R. Cassee (RIVM, the Netherlands). The physicochemical characteristics of this DEP is available elsewhere (Brinchmann et al., 2018b; Totlandsdal et al., 2010; Totlandsdal et al., 2014). To avoid loss of semi-volatile organic compounds, the DEP was stored at -18 °C. Combustion technology has advanced since these particles where collected, but the car-fleet in most cities is still composed of vehicles of differing age and type. These particles where used since they contain a high level (approximately 60%) of organic carbon. DEP-EOM was extracted with a sequence of solvents of increasing polarity ranging from non-polar to polar and chemically characterized as previously described (Brinchmann et al., 2018a; Brinchmann et al., 2018b; Cochran and Kubátová, 2015). In short, DEP-EOM was extracted by sequential pressurized extraction (137 bar) with: *n*-hexane (*n*-Hex-EOM: non-polar), dichloromethane (DCM-EOM: weakly polar) and methanol (Methanol-EOM: semi-polar) at 100°C followed by a final extraction with water at 25°C (Water-EOM: highly polar). The solvents were removed by evaporating the samples to dryness under a gentle stream of nitrogen and extracted DEP-EOM re-suspended in DMSO at concentrations corresponding to extracts from 25 mg/mL of the original DEP.

Chemical composition of *DEP-EOM fractions*: The chemical composition of the four fractionated DEP-EOM fractions have been characterized elsewhere (Brinchmann et al., 2018a; Brinchmann et al., 2018b). In brief, the majority of organic chemicals extracted, where contained in the *n*-Hex- and DCM-EOM fractions. Furthermore, PAHs and aliphatic hydrocarbons, where only retrieved in *n*-Hex- and DCM-EOM. The *n*-Hex-EOM had substantially higher concentrations of PAHs and aliphatic hydrocarbons (respectively 1.5 and 100 mg/g DEP) compared to DCM-EOM (respectively 0.3 and 17.5 mg/g DEP). The dominating PAHs and PAH-derivatives contained in *n*-Hex-EOM were (in rank order): methylated phenanthrenes and/or anthracenes >> phenanthrene > pyrene > methylated fluoranthenes and/or pyrenes > chrysene > 1-nitropyrene > xanthone > fluoranthene > 9-fluorenone. In DCM-EOM the only PAHs found in considerable amounts were (in

rank order): pyrene > phenanthrene  $\approx$  fluoranthrene  $\approx$  chrysene. Methylated, oxidized or nitrated PAH-species were not detected in DCM-EOM (Brinchmann et al., 2018a). Notably, the total amount of organic carbon extracted by *n*-hexane (153 mg/g DEP), DCM (113 mg/g DEP) and methanol (62 mg/g DEP) decreased according to polarity of the solvents (Brinchmann et al., 2018b). However, organic carbon was much more evenly distributed across these three fractions, compared to PAHs and aliphatic chemicals, which were predominately extracted by *n*-hexane. The reason for this apparent discrepancy, was that the DCM and methanol extracts predominately contained higher molecular weight (MW) compounds which could not be detected by GC-MS since their boiling points were above the  $\sim 300^{\circ}\text{C}$  of the GC-injector (Brinchmann et al., 2018b).

### 2.3. Cell cultures and treatments

*Human microvascular endothelial cell-line (HMEC-1)* (LGC Standards, Germany) were maintained in MCDB131 medium containing epidermal growth factor (10 ng/mL), hydrocortisone (0.2  $\mu\text{g/mL}$ ), penicillin (50 unit/mL), and streptomycin (50  $\mu\text{g/mL}$ ) and supplemented with 10% fetal calf serum (FCS), according to the providers instructions.

*Human embryonic kidney cells (HEK293)* were maintained in Dulbecco's modified Eagle's medium, containing penicillin (50 units/mL) and streptomycin (50  $\mu\text{g/mL}$ ) and supplemented with 10% FCS. HEK293 permanently expressing  $\beta 1$ - and  $\beta 2$ ARnergic receptors ( $\beta 1\text{AR}$ ,  $\beta 2\text{AR}$ ) were obtained by  $\beta 1\text{AR}$ ,  $\beta 2\text{AR}$  cDNA transfection as previously described (Mayati et al., 2012b). Briefly, HEK293 were seeded at  $2.5 \times 10^5$  cells/well in 6-well plates, transfected with either 2.5  $\mu\text{g}$  of empty pcDNA3.1(+)*neo* vector (HEKwt) or 2.5  $\mu\text{g}$  of pcDNA3.1(+)*neo* vector containing HA-tagged human  $\beta 1\text{AR}$  or  $\beta 2\text{AR}$  ORF (HEK  $\beta 1$  and HEK  $\beta 2$ ), and subsequently selected with G418 sulfate (1 mg/mL).

Chemicals were prepared as stock solution in DMSO. The final concentration of solvent did not exceed 0.2% (v/v); control cultures received similar concentration of DMSO. In all experiments with chemical inhibitors, cells were pre-treated 30 min prior to and during exposure.

### 2.4. Calcium measurements

Variations in intracellular  $\text{Ca}^{2+}$  concentrations ( $[\text{Ca}^{2+}]_i$ ) were analysed in HMEC-1 and HEK293 exposed to DEP-EOM, by micro-spectrofluorometry using the  $\text{Ca}^{2+}$  sensitive probe Fura-2AM, as previously reported (Brinchmann et al., 2018a). Briefly, cells were incubated at  $37^{\circ}\text{C}$  in cell suspension buffer (134.8 mM NaCl, 4.7 mM KCl, 1.2 mM  $\text{K}_2\text{HPO}_4$ , 1 mM  $\text{MgCl}_2$ , 1 mM  $\text{CaCl}_2$ , 10 mM glucose, 10 mM HEPES, pH 7.4) supplemented with 1.5  $\mu\text{M}$  Fura-2AM and 0.006%



pluronic acid. After 30 min loading, cells were washed two times with the buffer before exposure. DEP-EOM corresponding to 5 µg/mL of original DEP was added to the buffer as a bolus dose, after 3 minutes of measuring baseline calcium levels.  $[Ca^{2+}]_i$  imaging in HMEC-1 exposed to all four DEP-EOM was conducted at 0 and 60 min of exposure. The experimental protocol for  $[Ca^{2+}]_i$  imaging in HEK293 and HMEC-1 exposed to *n*-Hex- or DCM-EOM with or without inhibitors, involved data acquisition every 10<sup>th</sup> second (emission at 510 nm) at 340- and 380-nm excitation wavelengths. Changes in  $[Ca^{2+}]_i$  were monitored using a DMIRB (Leica, Wetzlar, Germany) inverted microscope-based imaging system equipped with a 40×/1.35 UApo N340 high UV light transmittance oil immersion objective (Olympus, Waltham, MA, USA), a CoolSnapHQ fast-cooled monochromatic digital camera (Princeton instrument), a DG-4 Ultra High Speed Wavelength Switcher (Sutter Instruments, Novato, CA, USA) for fluorophore excitation, and METAFLUOR software (Universal Imaging, Downingtown, PA, USA) for image acquisition and analysis. Analysis involved determination of pixels assigned to each cell. The average pixel value allocated to each cell was obtained with excitation at each wavelength and corrected for background. The ratio was obtained after dividing the 340-nm by the 380-nm fluorescence image on a pixel-by-pixel base ( $R=F_{340\text{ nm}}/F_{380\text{ nm}}$ ). Results are presented as normalized calcium level compared to basal  $[Ca^{2+}]_i$  measured 3 min prior to exposure. Area under the curve (AUC) was calculated from baseline (1.0).

## 2.5. Gene expression analysis by real-time qPCR

*HMEC-1* destined for q-PCR were grown to near-confluency and serum starved for a minimum of 12 h prior to exposure. Cells were then exposed by removing the media and adding growth medium without FCS containing the various DEP-EOM (*n*-Hex-, DCM-, Methanol- or Water-EOM) or DMSO. Exposure concentrations corresponded to 5 or 50 µg/mL of original DEP. After 5 or 24 h exposure, cells were harvested and mRNA isolated. Total RNA was isolated using NucleoSpin RNA Plus (Macherey-Nagel; Düren, Germany) and reverse transcribed to cDNA on a PCR System 2400 (PerkinElmer, Waltham, MA, USA) using a High Capacity cDNA Archive Kit (Applied Biosystems, Foster City, CA, USA). Real-time qPCR was performed using pre-designed TaqMan Gene Expression Assays and TaqMan Universal PCR Master Mix and run on Applied Biosystems 7500 fast software (Applied Biosystems, Foster City, CA, USA). Gene expression of induced IL-1 $\alpha$  (Hs00174092\_m1), IL-1 $\beta$  (Hs01555410\_m1), CXCL8 (Hs00174103\_m1), COX-2 (Hs00153133\_m1), MMP-1 (Hs00899658\_m1), HO-1 (Hs01110250\_m1), PAI-2/SERPIN2 (Hs01010736\_m1),  $\beta$ 1AR (Hs02330048\_s1) and  $\beta$ 2AR (Hs00240532\_s1) were normalized against

GAPDH (Hs02758991\_g1), and expressed as fold change compared to untreated control as calculated by the  $\Delta\Delta\text{Ct}$  method ( $\Delta\text{Ct} = \text{Ct}[\text{Gene of Interest}] - \text{Ct}[\text{GAPDH}]$ ;  $\Delta\Delta\text{Ct} = \Delta\text{Ct}[\text{Treated}] - \Delta\text{Ct}[\text{Control}]$ ; Fold change =  $2^{-\Delta\Delta\text{Ct}}$ ).

## 2.6. Statistical analysis

Statistical analysis was performed by ANOVA with Holm-Sidak post-test for multiple comparisons. As ANOVA cannot be performed on normalized data, gene expressions were analyzed using the deltaCT-values from the q-PCR measurements. Effects on  $[\text{Ca}^{2+}]_i$  were quantified as area under the curve (AUC) and statistical comparison conducted with non-parametric t-tests. All calculations were performed using GraphPad Prism 7 software (GraphPad Software, Inc., San Diego, CA).

## 3. Results:

### 3.1 Effects of DEP-EOM on $[\text{Ca}^{2+}]_i$ and the involvement of $\beta\text{AR}$ signalling.

Cells were first exposed to DEP-EOM fractionated by sequential extraction with solvents of increasing polarity, *n*-Hex-, DCM-, Methanol- and Water-EOM, at concentrations corresponding to 5  $\mu\text{g}/\text{mL}$  of the original DEP.  $[\text{Ca}^{2+}]_i$  was measured prior to exposure and after 1 h. In corroboration with our previous observations (Brinchmann et al., 2018a), exposure to *n*-Hex-EOM and DCM-EOM caused a marked (2 fold)  $[\text{Ca}^{2+}]_i$  increase (Fig. 1), compared to baseline levels measured prior to exposure. The fractions extracted by the two more polar solvents, Methanol- and Water-EOM, had negligible effects on  $[\text{Ca}^{2+}]_i$  in HMEC-1 (Fig. 1). To explore if *n*-Hex- and DCM-EOM increased  $[\text{Ca}^{2+}]_i$  via  $\beta\text{AR}$  signalling, we used the potent and unselective  $\beta\text{AR}$ -antagonist carazolol (Innis et al., 1979). Cells were pre-treated with carazolol (10  $\mu\text{M}$ ) or DMSO, and exposed 1 h to *n*-Hex- or DCM-EOM at concentrations corresponding to extracts from 5  $\mu\text{g}/\text{mL}$  of original DEP. Carazolol caused a marked reduction of  $[\text{Ca}^{2+}]_i$  triggered by both *n*-Hex- and DCM-EOM in HMEC-1 (Fig. 2). While the overall suppressive effects of carazolol  $[\text{Ca}^{2+}]_i$  was only slightly larger on DCM-EOM compared to *n*-Hex-EOM (as evident from the AUC estimates), DCM-EOM induced  $[\text{Ca}^{2+}]_i$  was considerably more affected at the later time points. After 60 min exposure, carazolol treatment caused almost 70% reduction in DCM-EOM-induced  $[\text{Ca}^{2+}]_i$  increase, whereas *n*-Hex-EOM induced  $[\text{Ca}^{2+}]_i$  was only suppressed by approximately 40%. Gene-expression analysis of HMEC-1 exposed to the various DEP-EOM indicated that compounds in *n*-Hex- and DCM-EOM resulted in a downregulation of  $\beta\text{AR}$  after 24 h exposure (Fig. 3). In line with our

previous report on B[a]P (Mayati et al., 2017), this indicates that lipophilic organic chemicals extracted by the two least polar solvents interacts with and desensitizes  $\beta$ ARs. In contrast, the hydrophilic extracts had no significant effects.

To further study the link between  $\beta$ AR and increased  $[Ca^{2+}]_i$  we explored effects of *n*-Hex- and DCM-EOM in HEK293 cells, known to express little or no  $\beta$ AR. HEK293 wild type cells (HEK WT) and HEK293 cells over-expressing  $\beta$ 1AR (HEK  $\beta$ 1) or  $\beta$ 2AR (HEK  $\beta$ 2) were then exposed to *n*-Hex- or DCM-EOM (5  $\mu$ g/mL) and  $[Ca^{2+}]_i$  measured. It seemed that *n*-Hex-EOM increased  $[Ca^{2+}]_i$  via other receptors than  $\beta$ ARs in HEK293, as  $[Ca^{2+}]_i$  was increased substantially in HEK WT (Fig. 4A). Notably, over-expression of  $\beta$ AR rather had a negative effect on *n*-Hex-EOM triggered  $[Ca^{2+}]_i$  increase. DCM-EOM on the other hand, only marginally affected  $[Ca^{2+}]_i$  in HEK WT, and  $[Ca^{2+}]_i$ -responses were considerably stronger in HEK293 overexpressing  $\beta$ 1- or  $\beta$ 2AR (Fig. 4B). Thus, DCM-EOM seemed to increase  $[Ca^{2+}]_i$  via  $\beta$ AR signalling in HEK293 as well as HMEC-1. To further clarify the role of  $\beta$ AR in mediating DCM-EOM-triggered calcium we used the more  $\beta$ 2-selective inhibitor ICI-118,551, that reduced  $[Ca^{2+}]_i$  in HMEC-1 indicating a possible role of  $\beta$ 2AR (Supplementary Fig. S1).

PAR-2 is an important mediator of endothelial dysfunction (Alberelli and De Candia, 2014). We have previously shown that PAR-2 is involved in mediating the inflammatory responses triggered by *n*-Hex- and DCM-EOM in HMEC-1 and primary human endothelial cells (Brinchmann et al., 2018b). We thus wanted to explore the role of PAR2 in calcium effects of *n*-Hex and DCM-EOM. The inhibitor ENMD-1068 (2.5 mM) did not affect *n*-Hex-EOM-induced  $[Ca^{2+}]_i$  increase (Fig 5A), but effects of DCM-EOM was reduced by ENMD-1068 treatment (Fig. 5B).

### 3.2. Effects of lipophilic DEP-EOM on inflammation-associated genes, involvement of $\beta$ ARs and $[Ca^{2+}]_i$ .

In a recent study on the currently used DEP-EOM, we found that *n*-Hex- and DCM-EOM affected inflammation-associated genes in HMEC-1 and primary endothelial cells (Brinchmann et al., 2018b). In agreement with the current results on  $[Ca^{2+}]_i$  and  $\beta$ AR expression, the hydrophilic extracts, methanol-EOM and water-EOM had little or no effect on inflammation-associated genes. To explore the role of  $\beta$ ARs with regard to expression of genes linked to inflammation, we pre-treated HMEC-1 with the  $\beta$ AR-antagonist carazolol, prior to exposure to *n*-Hex- and DCM-EOM at a concentration corresponding to 50  $\mu$ g/mL of original DEP. Carazolol did not inhibit the *n*-Hex-EOM-induced gene-expression (Fig 6A), but somewhat surprisingly

augmented expression of IL-1 $\alpha$  (from 5 fold to 8 fold). By contrast, in HMEC-1 exposed to DCM-EOM carazolol suppressed the up-regulation of COX-2 primarily, and to a certain extent CXCL8 and MMP-1 (Fig. 6B). Induction of MMP-1 was small and effects on CXCL8 were variable, thus these effects should be interpreted with caution.

In an attempt to further link inflammatory effects to  $[Ca^{2+}]_i$ , we applied SKF 96365, an inhibitor of transient receptor potential (TRP) channels. We previously found that SKF 96365 reduced effects of DCM-EOM and to a lesser degree *n*-Hex-EOM on  $[Ca^{2+}]_i$  (Brinchmann et al., 2018a). HMEC-1 pre-treated with SKF 96365 or DMSO, were exposed 5 h to *n*-Hex- or DCM-EOM (50  $\mu$ g/mL). None of the inflammation-associated genes induced by *n*-Hex-EOM were significantly affected by SKF 96365 (Fig. 7A), while DCM-EOM-induced COX-2 was markedly reduced (Fig. 7B).

#### 4. Discussion:

The intracellular second messenger calcium is kept at a low cytoplasmic concentration in resting cells (Clapham, 2007). Furthermore, inflammatory effects of xenobiotics known to be present on DEP often seem to depend on  $[Ca^{2+}]_i$  (Monteiro et al., 2008; N'Diaye et al., 2006; Ovrevik et al., 2017). Recent studies suggest that some PAHs and DEP may trigger  $[Ca^{2+}]_i$  increase as well as inflammation *via* GPCRs like  $\beta$ ARs and PAR-2 (Bach et al., 2015; Li et al., 2011; Mayati et al., 2014; Mayati et al., 2012b). Notably, effects of DEP on  $[Ca^{2+}]_i$  and MMP-1 seemed to be due to lipophilic EOMs in bronchial cells (Li et al., 2011). In line with this, we previously found that lipophilic DEP-EOM triggered pro-inflammatory responses and disrupted  $[Ca^{2+}]_i$  *via* AhR nongenomic signalling in endothelial cells (Brinchmann et al., 2018a; Brinchmann et al., 2018b). In the current study we investigated the role of the GPCRs  $\beta$ AR and PAR-2 in these effects and the role of  $[Ca^{2+}]_i$  increase in the pro-inflammatory effects of DEP-EOMs. We found that *n*-Hex- and DCM-EOM in contrast to the more polar DEP-EOM increased  $[Ca^{2+}]_i$  in HMEC-1, and that these effects partly depended on  $\beta$ ARs. Furthermore, some of the inflammation associated genes induced by DCM-EOM seemed to depend on  $\beta$ ARs, and effects on  $[Ca^{2+}]_i$  and COX-2 seemed partly interconnected.

Effects of PM depends on various physicochemical properties, implying that PM and its constituents may trigger toxic responses via multiple mechanisms (Lewtas, 2007; Longhin et al., 2016; Ovrevik et al., 2017). While the larger particles (PM<sub>10</sub>) are associated with lung damage, the

smaller particles (PM<sub>2,5</sub>) have been especially connected to CVD (Alfaro-Moreno et al., 2007; Brook et al., 2010). Interestingly, ultrafine particles with even more pro-atherogenic potential than PM<sub>2,5</sub>, contained twice as much organic chemicals (Araujo et al., 2008). Furthermore, particulates denuded of organic chemicals lost their atherogenic potential (Keebaugh et al., 2015). Thus, it seems that organic chemicals may be crucial in vascular effects caused by combustion particles. Based on this we chose a DEP sample with a high organic content.

We have previously found that DEP-EOM increased  $[Ca^{2+}]_i$  in HMEC-1 at non-cytotoxic concentrations, and the induced changes were at least partly reversible at concentrations currently used (5 µg/mL) (Brinchmann et al., 2018a; Brinchmann et al., 2018b). More specifically, *n*-Hex-EOM appeared to trigger store operated calcium entry (SOCE), while DCM-EOM activated a  $[Ca^{2+}]_i$  response resembling receptor-operated calcium entry (ROCE), that seemed totally dependent on extracellular calcium, in HMEC-1. Both effects seemed largely dependent on AhR nongenomic signalling (Brinchmann et al., 2018a). Chemical analysis of these DEP-extracts showed marked differences in chemical composition. While *n*-Hex-EOM contained a multitude of PAHs, we only detected considerable amounts of phenanthrene, fluoranthene, pyrene and chrysene in DCM-EOM. Ongoing studies have revealed that it is difficult to estimate calcium responses from chemical composition in complex mixtures, as B[a]P attenuated pyrene-induced  $[Ca^{2+}]_i$  increase (Brinchmann et al submitted). Thus, the marked differences in  $[Ca^{2+}]_i$  response patterns induced by the two DEP-EOM fractions (SOCE vs ROCE) may indeed relate to differences in chemical composition, more specifically a changed balance between specific PAHs. However, the total amount of organic chemicals was higher in *n*-Hex- (~150 mg/g of original DEP) compared to DCM-EOM (~110 mg/g), and *n*-Hex-EOM contained approximately 5-fold more PAHs and 7-fold more aliphatic compounds (Brinchmann et al., 2018a). Furthermore, we have found that *n*-Hex-EOM increased  $[Ca^{2+}]_i$  at considerably lower concentrations than DCM-EOM (Brinchmann et al., 2018a). Thus, we cannot exclude the possibility that discrepancies in  $[Ca^{2+}]_i$  response patterns may just relate to higher concentrations of organic compounds in *n*-Hex-EOM.

The most robust finding of the present study is that βARs were involved in effects of DCM-EOM on  $[Ca^{2+}]_i$  in both HMEC-1 and HEK293. Compared to HMEC-1, βARs are poorly expressed in HEK293 WT cells (Mayati et al., 2012b; von Zastrow and Kobilka, 1992). In line with this, DCM-EOM induced only marginal  $[Ca^{2+}]_i$  responses in HEK293 WT cells, but this response was substantially increased in HEK293 over-expressing β1- or β2AR. This thus indicates that βARs mediated-effects of chemicals present in DCM-EOM may not be restricted by cell type.

Acknowledging that these lipophilic DEP-EOM affect multiple mechanisms, it is tempting to speculate that DCM-EOM activates ROCE *via* the GPCRs  $\beta$ AR and PAR-2. Studies indicating that certain PAHs may act as  $\beta$ AR agonists and that DEP increase  $[Ca^{2+}]_i$  *via* PAR-2 in human bronchial cells, lend support to this suggestion (Li et al., 2011; Mayati et al., 2014; Mayati et al., 2012b; Mayati et al., 2017). However, as we have previously observed that DCM-EOM increased  $[Ca^{2+}]_i$  *via* AhR-dependent mechanisms, the possibility that GPCRs could be trans-activated downstream of AhR activation also needs to be considered.

Notably, AhR-nongenomic signalling appears to be the main triggering mechanism for both the *n*-Hex- and DCM-EOM induced  $[Ca^{2+}]_i$ -increase in HMEC-1 (Brinchmann et al., 2018a). While the carazolol-effect suggests that  $\beta$ ARs also contributed to *n*-Hex-EOM-induced  $[Ca^{2+}]_i$ -increase in HMEC-1, the results obtained in HEK293 appeared quite contradictory. In HEK293 WT, which constitutively express marginal levels of  $\beta$ AR (von Zastrow and Kobilka, 1992), *n*-Hex-EOM increased  $[Ca^{2+}]_i$  markedly, clearly showing that this effect was independent of  $\beta$ ARs and triggered *via* other signalling mechanisms. Furthermore, overexpression of  $\beta 1$ -/ $\beta 2$ AR suppressed *n*-Hex-EOM triggered  $[Ca^{2+}]_i$ -increase in HEK293, thus strongly suggesting that there also may be negative crosstalk between  $\beta 1$ -/ $\beta 2$ AR and AhR signalling pathways, in addition to the positive crosstalk previously reported (Brinchmann et al., 2018a). The nature and overall relevance of this apparent interaction between AhR- and  $\beta$ AR-signalling remains to clarify. It should however, be noted that examples from the literature indicate that increase of  $\beta$ AR-signalling may negatively affect other signalling pathways and thus  $[Ca^{2+}]_i$ . For instance, phosphatidylinositol 4,5-bisphosphate (PIP<sub>2</sub>)-dependent calcium channels will be negatively affected by  $\beta$ ARs ligands that primarily activate phospholipase C (PLC), leading to PIP<sub>2</sub> hydrolysis (Putney and Tomita, 2012; Suh and Hille, 2005). As PIP<sub>2</sub> positively regulates a wide range of ion channels, a reduction of PIP<sub>2</sub>-levels through hydrolysis would be expected to reduce  $Ca^{2+}$ -influx through these channels (Suh and Hille, 2005).

COX-2 may produce prostaglandin E<sub>2</sub>, which promotes expression of matrix metalloproteinases (MMPs), tissue destruction, cell death and destabilization of atherosclerotic plaques (Bishop-Bailey et al., 2006; Gomez et al., 2014; Gomez et al., 2013; Newby, 2016; Walton et al., 1999). In bronchial epithelial cells it has been found that lipophilic components of DEP induce MMP-1 *via* calcium signalling (Li et al., 2011). Thus, a central focus of this study was to explore whether DEP-EOM-induced increases in  $[Ca^{2+}]_i$  and gene-expression were interconnected, or rather parallel events. The  $\beta$ AR antagonist carazolol and the calcium antagonist SKF96365, had

little or no effect on *n*-Hex-EOM and most of the genes induced by DCM-EOM were unaffected. Thus,  $\beta$ ARs and  $\text{Ca}^{2+}$ -signalling did not seem to be pivotal in mediating the observed effects on inflammation-associated genes. However, DCM-EOM-induced COX-2 expression was reduced by both carazolol and SKF96365, indicating that  $\text{Ca}^{2+}$ -signalling is an upstream event at least partly involved in regulation of COX-2. This may be related to protein kinase C (PKC), which is involved in the regulation of COX-2 and is activated by  $\text{Ca}^{2+}$  and di-acyl-glycerol (DAG) (Mochly-Rosen et al., 2012). Moreover, GPCRs may activate PLC, which is detrimental to  $[\text{Ca}^{2+}]_i$ -regulation (Putney and Tomita, 2012). Thus, a possible chain of events is that certain lipophilic chemicals in DEP may activate GPCRs, directly or indirectly through AhR, subsequently triggering the PLC/DAG/ $\text{Ca}^{2+}$ /PKC-cascade leading to activation of COX-2 and other pro-inflammatory genes. In contrast, *n*-Hex-EOM-induced gene expression was not affected by neither SKF96365 nor carazolol, despite effects of these inhibitors on *n*-Hex-EOM-induced  $[\text{Ca}^{2+}]_i$  in HMEC-1. Thus,  $[\text{Ca}^{2+}]_i$  does not seem central to these responses. However, care should be taken when comparing these findings, as effects on inflammation-associated genes were examined after 5 h exposure to 10-fold higher DEP-EOM-concentrations than those used to study  $\text{Ca}^{2+}$ -signalling over the first 60 min of exposure.

In conclusion, neither  $\beta$ AR nor PAR-2 were consistently involved in effects of *n*-Hex-EOM in HMEC-1 cells, but both GPCRs seemed at least partly involved in regulation of calcium signalling and COX-2 responses in cells exposed to DCM-EOM. This discrepancy in involvement of GPCRs in cellular effects of DEP-EOM fractionated by solvents of increasing polarity is likely due to differences in chemical composition and/or the amount of active compounds extracted. Thus,  $\beta$ AR and PAR-2, does not appear to play a major role in mediating the observed effects of DEP-EOM on  $[\text{Ca}^{2+}]_i$  and inflammation-associated genes in HMEC-1, but may conceivably contribute to modulate responses.

### Competing interests

The authors report no competing interests. The authors alone are responsible for the content and writing of the paper.

### Authors' contributions

BB performed all experiments, and contributed in all experimental planning and design in collaboration with ELF, JØ, DLG and JAH. BB, NP and JØ performed data analysis and statistics. JØ conceived and coordinated the study, with support of JAH, ELF and DLG. BB drafted the first

versions of the manuscript and wrote the final version in collaboration with JØ and JAH. All authors read, commented and approved the final manuscript.

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#### Data Statement:

Data will be made available on request.

ACCEPTED MANUSCRIPT



## References:

- Alberelli, M.A., De Candia, E., 2014. Functional role of protease activated receptors in vascular biology. *Vascul Pharmacol* 62, 72-81.
- Alfaro-Moreno, E., Nawrot, T.S., Nemmar, A., Nemery, B., 2007. Particulate matter in the environment: pulmonary and cardiovascular effects. *Current opinion in pulmonary medicine* 13, 98-106.
- Araujo, J.A., Barajas, B., Kleinman, M., Wang, X., Bennett, B.J., Gong, K.W., Navab, M., Harkema, J., Sioutas, C., Lusk, A.J., Nel, A.E., 2008. Ambient particulate pollutants in the ultrafine range promote early atherosclerosis and systemic oxidative stress. *Circ Res* 102, 589-596.
- Bach, N., Bolling, A.K., Brinckmann, B.C., Totlandsdal, A.I., Skuland, T., Holme, J.A., Lag, M., Schwarze, P.E., Ovrevik, J., 2015. Cytokine responses induced by diesel exhaust particles are suppressed by PAR-2 silencing and antioxidant treatment, and driven by polar and non-polar soluble constituents. *Toxicol Lett* 238, 72-82.
- Barath, S., Mills, N.L., Lundback, M., Tornqvist, H., Lucking, A.J., Langrish, J.P., Soderberg, S., Boman, C., Westerholm, R., Londahl, J., Donaldson, K., Mudway, I.S., Sandstrom, T., Newby, D.E., Blomberg, A., 2010. Impaired vascular function after exposure to diesel exhaust generated at urban transient running conditions. *Part Fibre Toxicol* 7, 19.
- Bishop-Bailey, D., Mitchell, J.A., Warner, T.D., 2006. COX-2 in cardiovascular disease. *Arterioscler Thromb Vasc Biol* 26, 956-958.
- Bonvallot, V., Baeza-Squiban, A., Baulig, A., Brulant, S., Boland, S., Muzeau, F., Barouki, R., Marano, F., 2001. Organic compounds from diesel exhaust particles elicit a proinflammatory response in human airway epithelial cells and induce cytochrome p450 1A1 expression. *Am J Respir Cell Mol Biol* 25, 515-521.
- Brinckmann, B.C., Le Ferrec, E., Podechard, N., Lagadic-Gossman, D., Shoji, K.F., Penna, A., Kukowski, K., Kubatova, A., Holme, J.A., Ovrevik, J., 2018a. Lipophilic Chemicals from Diesel Exhaust Particles Trigger Calcium Response in Human Endothelial Cells via Aryl Hydrocarbon Receptor Non-Genomic Signalling. *International journal of molecular sciences* 19.
- Brinckmann, B.C., Skuland, T., Rambol, M.H., Szoke, K., Brinckmann, J.E., Gutleb, A.C., Moschini, E., Kubatova, A., Kukowski, K., Le Ferrec, E., Lagadic-Gossman, D., Schwarze, P.E., Lag, M., Refsnes, M., Ovrevik, J., Holme, J.A., 2018b. Lipophilic components of diesel exhaust particles induce pro-inflammatory responses in human endothelial cells through AhR dependent pathway(s). *Part Fibre Toxicol* 15, 21.
- Brook, R.D., Rajagopalan, S., 2012. Chronic air pollution exposure and endothelial dysfunction: what you can't see--can harm you. *J Am Coll Cardiol* 60, 2167-2169.
- Brook, R.D., Rajagopalan, S., Pope, C.A., 3rd, Brook, J.R., Bhatnagar, A., Diez-Roux, A.V., Holguin, F., Hong, Y., Luepker, R.V., Mittleman, M.A., Peters, A., Siscovick, D., Smith, S.C., Jr., Whitsel, L., Kaufman, J.D., American Heart Association Council on, E., Prevention, C.o.t.k.i.C.D., Council on Nutrition, P.A., Metabolism, 2010. Particulate matter air pollution and cardiovascular disease: An update to the scientific statement from the American Heart Association. *Circulation* 121, 2331-2378.
- Bylund, D.B., Eikenberg, D.C., Hieble, J.P., Langer, S.Z., Lefkowitz, R.J., Minneman, K.P., Molinoff, P.B., Ruffolo, R.R., Jr., Trendelenburg, U., 1994. International Union of Pharmacology nomenclature of adrenoceptors. *Pharmacol Rev* 46, 121-136.
- Campen, M.J., 2012. Vascular endothelium as a target of diesel particulate matter-associated toxicants. *Arch Toxicol* 86, 517-518.
- Clapham, D.E., 2007. Calcium signaling. *Cell* 131, 1047-1058.

- Cochran, R.E., Kubátová, A., 2015. Pressurised fluid extraction of polycyclic aromatic hydrocarbons and their polar oxidation products from atmospheric particles. *International Journal of Environmental Analytical Chemistry* 95, 434-452.
- Cohen A. J., Anderson H. R., Ostro B., Pandey K. D., Krzyzanowski M., Künzli N., Gutschmidt K, Pope III C. A., Romieu I., Samet J. M., R., S.K., 2004. Urban Air Pollution. In: Ezzati, M., Lopez, A.D., Rodgers, A., Murray, C.J.L. (Eds.) *Comparative Quantification of Health Risks - Global and Regional Burden of Diseases Attributable to Selected Major Risk Factors*. World Health Organization, World Health Organization.
- De Backer, G., 2003. European guidelines on cardiovascular disease prevention in clinical practice Third Joint Task Force of European and other Societies on Cardiovascular Disease Prevention in Clinical Practice (constituted by representatives of eight societies and by invited experts). *European Heart Journal* 24, 1601-1610.
- Dharmashankar, K., Widlansky, M.E., 2010. Vascular endothelial function and hypertension: insights and directions. *Curr Hypertens Rep* 12, 448-455.
- Donaldson, K., Stone, V., Seaton, A., MacNee, W., 2001. Ambient Particle Inhalation and the Cardiovascular System: Potential Mechanisms. *Environ Health Perspect*.
- Edwards, M.R., Haas, J., Panettieri, R.A., Jr., Johnson, M., Johnston, S.L., 2007. Corticosteroids and beta2 agonists differentially regulate rhinovirus-induced interleukin-6 via distinct Cis-acting elements. *J Biol Chem* 282, 15366-15375.
- Gerde, 2001. The rapid alveolar absorption of diesel soot-adsorbed Benzo[a]Pyrene: bioavailability, metabolism and dosimetry of an inhaled particle-borne carcinogen. *Carcinogenesis*.
- Gomez, I., Benyahia, C., Louedec, L., Leseche, G., Jacob, M.P., Longrois, D., Norel, X., 2014. Decreased PGE(2) content reduces MMP-1 activity and consequently increases collagen density in human varicose vein. *PLoS One* 9, e88021.
- Gomez, I., Foudi, N., Longrois, D., Norel, X., 2013. The role of prostaglandin E2 in human vascular inflammation. *Prostaglandins Leukot Essent Fatty Acids* 89, 55-63.
- Grahame, T.J., Klemm, R., Schlesinger, R.B., 2014. Public health and components of particulate matter: the changing assessment of black carbon. *Journal of the Air & Waste Management Association* (1995) 64, 620-660.
- HEI, 2017. *STATE OF GLOBAL AiR /2017*. Boston, MA:Health Effects Institute and The Institute for Health Metrics and Evaluation.
- Holden, N.S., Rider, C.F., Bell, M.J., Velayudhan, J., King, E.M., Kaur, M., Salmon, M., Giembycz, M.A., Newton, R., 2010. Enhancement of inflammatory mediator release by beta(2)-adrenoceptor agonists in airway epithelial cells is reversed by glucocorticoid action. *Br J Pharmacol* 160, 410-420.
- Innis, R.B., Correa, F.M., Synder, S.H., 1979. Carazolol, an extremely potent beta-adrenergic blocker: binding to beta-receptors in brain membranes. *Life Sci* 24, 2255-2264.
- Kawasaki, S., Takizawa, H., Takami, K., Desaki, M., Okazaki, H., Kasama, T., Kobayashi, K., Yamamoto, K., Nakahara, K., Tanaka, M., Sagai, M., Ohtoshi, T., 2001. Benzene-extracted components are important for the major activity of diesel exhaust particles: effect on interleukin-8 gene expression in human bronchial epithelial cells. *Am J Respir Cell Mol Biol* 24, 419-426.
- Keebaugh, A.J., Sioutas, C., Pakbin, P., Schauer, J.J., Mendez, L.B., Kleinman, M.T., 2015. Is atherosclerotic disease associated with organic components of ambient fine particles? *Sci Total Environ* 533, 69-75.

- Kolmus, K., Tavernier, J., Gerlo, S., 2015. beta2-Adrenergic receptors in immunity and inflammation: stressing NF-kappaB. *Brain, behavior, and immunity* 45, 297-310.
- Krishnan, R.M., Adar, S.D., Szpiro, A.A., Jorgensen, N.W., Van Hee, V.C., Barr, R.G., O'Neill, M.S., Herrington, D.M., Polak, J.F., Kaufman, J.D., 2012. Vascular responses to long- and short-term exposure to fine particulate matter: MESA Air (Multi-Ethnic Study of Atherosclerosis and Air Pollution). *J Am Coll Cardiol* 60, 2158-2166.
- Lawal, A.O., Zhang, M., Dittmar, M., Lulla, A., Araujo, J.A., 2015. Heme oxygenase-1 protects endothelial cells from the toxicity of air pollutant chemicals. *Toxicol Appl Pharmacol* 284, 281-291.
- Le Ferrec, E., Øvrevik, J., 2018. G-protein coupled receptors (GPCR) and environmental exposure. Consequences for cell metabolism using the  $\beta$ -adrenoceptors as example. *Current Opinion in Toxicology* 8, 14-19.
- Lewtas, J., 2007. Air pollution combustion emissions: characterization of causative agents and mechanisms associated with cancer, reproductive, and cardiovascular effects. *Mutat Res* 636, 95-133.
- Li, J., Kanju, P., Patterson, M., Chew, W.L., Cho, S.H., Gilmour, I., Oliver, T., Yasuda, R., Ghio, A., Simon, S.A., Liedtke, W., 2011. TRPV4-mediated calcium influx into human bronchial epithelia upon exposure to diesel exhaust particles. *Environ Health Perspect* 119, 784-793.
- Longhin, E., Gualtieri, M., Capasso, L., Bengalli, R., Mollerup, S., Holme, J.A., Ovrevik, J., Casadei, S., Di Benedetto, C., Parenti, P., Camatini, M., 2016. Physico-chemical properties and biological effects of diesel and biomass particles. *Environ Pollut* 215, 366-375.
- Lowell, B.B., Flier, J.S., 1997. Brown adipose tissue, beta 3-adrenergic receptors, and obesity. *Annual review of medicine* 48, 307-316.
- Lucking, A.J., Lundback, M., Barath, S.L., Mills, N.L., Sidhu, M.K., Langrish, J.P., Boon, N.A., Pourazar, J., Badimon, J.J., Gerlofs-Nijland, M.E., Cassee, F.R., Boman, C., Donaldson, K., Sandstrom, T., Newby, D.E., Blomberg, A., 2011. Particle traps prevent adverse vascular and prothrombotic effects of diesel engine exhaust inhalation in men. *Circulation* 123, 1721-1728.
- Maricq, M., 2007. Chemical characterization of particulate emissions from diesel engines: A review. *Journal of Aerosol Science* 38, 1079-1118.
- Mayati, A., Le Ferrec, E., Holme, J.A., Fardel, O., Lagadic-Gossmann, D., Ovrevik, J., 2014. Calcium signaling and beta2-adrenergic receptors regulate 1-nitropyrene induced CXCL8 responses in BEAS-2B cells. *Toxicol In Vitro* 28, 1153-1157.
- Mayati, A., Le Ferrec, E., Lagadic-Gossmann, D., Fardel, O., 2012a. Aryl hydrocarbon receptor-independent up-regulation of intracellular calcium concentration by environmental polycyclic aromatic hydrocarbons in human endothelial HMEC-1 cells. *Environ Toxicol* 27, 556-562.
- Mayati, A., Levoine, N., Paris, H., N'Diaye, M., Courtois, A., Uriac, P., Lagadic-Gossmann, D., Fardel, O., Le Ferrec, E., 2012b. Induction of intracellular calcium concentration by environmental benzo(a)pyrene involves a beta2-adrenergic receptor/adenylyl cyclase/Epac-1/inositol 1,4,5-trisphosphate pathway in endothelial cells. *J Biol Chem* 287, 4041-4052.
- Mayati, A., Podechard, N., Rineau, M., Sparfel, L., Lagadic-Gossmann, D., Fardel, O., Ferrec, E.L., 2017. Benzo(a)pyrene triggers desensitization of beta2-adrenergic pathway. *Sci Rep* 7, 3262.
- Mochly-Rosen, D., Das, K., Grimes, K.V., 2012. Protein kinase C, an elusive therapeutic target? *Nature reviews Drug discovery* 11, 937-957.
- Moller, P., Mikkelsen, L., Vesterdal, L.K., Folkmann, J.K., Forchhammer, L., Roursgaard, M., Danielsen, P.H., Loft, S., 2011. Hazard identification of particulate matter on vasomotor dysfunction and progression of atherosclerosis. *Crit Rev Toxicol* 41, 339-368.

- Monteiro, P., Gilot, D., Le Ferrec, E., Rauch, C., Lagadic-Gossmann, D., Fardel, O., 2008. Dioxin-mediated up-regulation of aryl hydrocarbon receptor target genes is dependent on the calcium/calmodulin/CaMKII $\alpha$  pathway. *Mol Pharmacol* 73, 769-777.
- N'Diaye, M., Le Ferrec, E., Lagadic-Gossmann, D., Corre, S., Gilot, D., Lecureur, V., Monteiro, P., Rauch, C., Galibert, M.D., Fardel, O., 2006. Aryl hydrocarbon receptor- and calcium-dependent induction of the chemokine CCL1 by the environmental contaminant benzo[a]pyrene. *J Biol Chem* 281, 19906-19915.
- Newby, A.C., 2016. Metalloproteinase production from macrophages - a perfect storm leading to atherosclerotic plaque rupture and myocardial infarction. *Experimental physiology* 101, 1327-1337.
- Ovrevik, J., Refsnes, M., Holme, J.A., Schwarze, P.E., Lag, M., 2013. Mechanisms of chemokine responses by polycyclic aromatic hydrocarbons in bronchial epithelial cells: sensitization through toll-like receptor-3 priming. *Toxicol Lett* 219, 125-132.
- Ovrevik, J., Refsnes, M., Lag, M., Brinchmann, B.C., Schwarze, P.E., Holme, J.A., 2017. Triggering Mechanisms and Inflammatory Effects of Combustion Exhaust Particles with Implication for Carcinogenesis. *Basic & clinical pharmacology & toxicology* 121 Suppl 3, 55-62.
- Penn, A., Murphy, G., Barker, S., Henk, W., Penn, L., 2005. Combustion-Derived Ultrafine Particles Transport Organic Toxicants to Target Respiratory Cells. *Environmental Health Perspectives* 113, 956-963.
- Putney, J.W., Tomita, T., 2012. Phospholipase C signaling and calcium influx. *Advances in biological regulation* 52, 152-164.
- Ramji, D.P., Davies, T.S., 2015. Cytokines in atherosclerosis: Key players in all stages of disease and promising therapeutic targets. *Cytokine Growth Factor Rev* 26, 673-685.
- Sandow, S.L., Haddock, R.E., Hill, C.E., Chadha, P.S., Kerr, P.M., Welsh, D.G., Plane, F., 2009. What's where and why at a vascular myoendothelial microdomain signalling complex. *Clin Exp Pharmacol Physiol* 36, 67-76.
- Sandow, S.L., Senadheera, S., Grayson, T.H., Welsh, D.G., Murphy, T.V., 2012. Calcium and endothelium-mediated vasodilator signaling. *Adv Exp Med Biol* 740, 811-831.
- Santos, I.N., Spadari-Bratfisch, R.C., 2006. Stress and cardiac beta adrenoceptors. *Stress (Amsterdam, Netherlands)* 9, 69-84.
- Suh, B.C., Hille, B., 2005. Regulation of ion channels by phosphatidylinositol 4,5-bisphosphate. *Current opinion in neurobiology* 15, 370-378.
- Totlandsdal, A.I., Cassee, F.R., Schwarze, P., Refsnes, M., Lag, M., 2010. Diesel exhaust particles induce CYP1A1 and pro-inflammatory responses via differential pathways in human bronchial epithelial cells. *Part Fibre Toxicol* 7, 41.
- Totlandsdal, A.I., Herseth, J.I., Bolling, A.K., Kubatova, A., Braun, A., Cochran, R.E., Refsnes, M., Ovrevik, J., Lag, M., 2012. Differential effects of the particle core and organic extract of diesel exhaust particles. *Toxicol Lett* 208, 262-268.
- Totlandsdal, A.I., Ovrevik, J., Cochran, R.E., Herseth, J.I., Bolling, A.K., Lag, M., Schwarze, P., Lilleaas, E., Holme, J.A., Kubatova, A., 2014. The occurrence of polycyclic aromatic hydrocarbons and their derivatives and the proinflammatory potential of fractionated extracts of diesel exhaust and wood smoke particles. *J Environ Sci Health A Tox Hazard Subst Environ Eng* 49, 383-396.
- von Zastrow, M., Kobilka, B.K., 1992. Ligand-regulated internalization and recycling of human beta 2-adrenergic receptors between the plasma membrane and endosomes containing transferrin receptors. *J Biol Chem* 267, 3530-3538.

Wachter, S.B., Gilbert, E.M., 2012. Beta-adrenergic receptors, from their discovery and characterization through their manipulation to beneficial clinical application. *Cardiology* 122, 104-112.

Walton, L.J., Franklin, I.J., Bayston, T., Brown, L.C., Greenhalgh, R.M., Taylor, G.W., Powell, J.T., 1999. Inhibition of prostaglandin E2 synthesis in abdominal aortic aneurysms: implications for smooth muscle cell viability, inflammatory processes, and the expansion of abdominal aortic aneurysms. *Circulation* 100, 48-54.

Yin, F., Ramanathan, G., Zhang, M., Araujo, J.A., 2013. Prooxidative effects of ambient pollutant chemicals are inhibited by HDL. *Journal of biochemical and molecular toxicology* 27, 172-183.

Zhao, M., Lytton, J., Burchiel, S.W., 1996. Inhibition of sarco-endoplasmic reticulum calcium ATPases (SERCA) by polycyclic aromatic hydrocarbons: lack of evidence for direct effects on cloned rat enzymes. *International journal of immunopharmacology* 18, 589-598.

Øvrevik, J., Arlt, V.M., Øya, E., Nagy, E., Mollerup, S., Phillips, D.H., Låg, M., Holme, J.A., 2010. Differential effects of nitro-PAHs and amino-PAHs on cytokine and chemokine responses in human bronchial epithelial BEAS-2B cells. *Toxicology and Applied Pharmacology* 242, 270-280.

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## Figure legends:

**FIGURE 1. Effects of DEP-EOM on  $[Ca^{2+}]_i$  in HMEC-1 cell-line.** A: Cells plated on glass lamellas were loaded with the  $Ca^{2+}$ -sensitive probe Fura2-AM and then exposed to DEP-EOM (corresponding to 5  $\mu$ g DEP/mL).  $[Ca^{2+}]_i$  level measured by normalized ratio of the Fura2-AM probe before exposure and after 60 min is presented. B: HMEC-1 cells exposed to the four DEP-extracts: *n*-Hex- (i), DCM- (ii), Methanol- (iii) and Water-EOM (iv) are visualized as presented. Results are expressed as mean  $\pm$  SEM; *n*-Hex-EOM and DCM-EOM: n=3; Methanol- and Water-EOM: n=2). \*Statistically significant different from baseline.

**FIGURE 2. Effects of the  $\beta$ AR antagonist carazolol on *n*-Hex- and DCM-EOM triggered  $[Ca^{2+}]_i$  in HMEC-1.** Cells were pre-treated with the unselective  $\beta$ AR antagonist carazolol (10  $\mu$ M) or vehicle (DMSO) 30 min prior to exposure. Three min after measurements were started, cells were exposed to *n*-Hex- or DCM-EOM (5  $\mu$ g/mL).  $[Ca^{2+}]_i$  level measured by normalized ratio of the Fura2-AM probe during exposure is presented as graph and the area under the curve (AUC) 0-60 min, as mean and mean  $\pm$  SEM (n=3). \*Statistically significant different from no inhibitor.

**FIGURE 3. DEP-EOM affects  $\beta$ AR expression in HMEC-1.** Cells were exposed to DEP-EOM (5 and 50  $\mu$ g/mL) ranging from lipophilic to hydrophilic, *n*-Hexane, DCM-, Methanol-, Water-EOM or vehicle (DMSO) alone. The expression of ADR $\beta$ 1 and ADR $\beta$ 2 was measured by q-PCR after 2 and 24 h. The m-RNA levels are relative to DMSO, represented by the dotted line at 1. Results are expressed as mean  $\pm$  SEM (n=4). \*Statistically significant different from unexposed controls.

**FIGURE 4. Effects of DEP-EOM on  $[Ca^{2+}]_i$  in HEK293 WT,  $\beta$ 1 and  $\beta$ 2.** A: Three min after measurements were started, HEK WT or HEK cells over expressing  $\beta$ 1AR or  $\beta$ 2AR were exposed to *n*-Hex- or DCM-EOM (5  $\mu$ g/mL).  $[Ca^{2+}]_i$  level measured by normalized ratio of the Fura2-AM probe during exposure is presented as graph and AUC 0-60 min, as mean and mean  $\pm$  SEM (n=3). # and \*statistically significant different from HEK WT.

**FIGURE 5. Inhibition of PAR-2 and  $[Ca^{2+}]_i$  increased by *n*-Hex- and DCM-EOM in HMEC-1.** Cells were incubated in buffer with or without the PAR-2 inhibitor ENMD-1068 (2.5 mM) 30 min prior to exposure. Three min after measurements were started, the cells were exposed to *n*-Hex- or DCM-EOM at concentrations corresponding to 5  $\mu$ g/mL of the original DEP or vehicle control

(DMSO).  $[Ca^{2+}]_i$  level measured by normalized ratio of the Fura2-AM probe during exposure is presented as graph and the area under the curve (AUC) 0-45 min, as mean and mean  $\pm$  SEM (*n*-Hex-EOM: *n*=1; DCM-EOM: *n*=3), respectively. \*Significantly different from no inhibitor.

**FIGURE 6. Effects of carazolol on *n*-Hex- and DCM-EOM induced genes in HMEC-1.** Cells were pre-treated with carazolol (10  $\mu$ M) 30 min and exposed to 50  $\mu$ g/mL of the lipophilic fractions, *n*-Hex- (A) or DCM-EOM (B), or vehicle (DMSO). Gene expression measured after 5 h by q-PCR. The m-RNA levels are expressed relative to DMSO, represented by the dotted line at 1. The results are expressed as mean  $\pm$  SEM (*n*=3). \*Statistically significant difference from unexposed controls. #Statistically significant difference from cells exposed to DEP-EOM without inhibitor.

**FIGURE 7. Effects of the calcium channel inhibitor SKF 96365 on *n*-Hex- and DCM-EOM induced genes in HMEC-1.** Cells were pre-treated with SKF96365 (10  $\mu$ M) 30 min and exposed to 50  $\mu$ g/mL of the lipophilic fractions, *n*-Hex- (A) or DCM-EOM (B), or vehicle (DMSO). Gene expression measured after 5 h by q-PCR. The m-RNA levels are expressed relative to DMSO, represented by the dotted line at 1. The results are expressed as mean  $\pm$  SEM (*n*=3). \*Statistically significant difference from unexposed controls. #Statistically significant difference from cells exposed to DEP-EOM without inhibitor.

Figures:

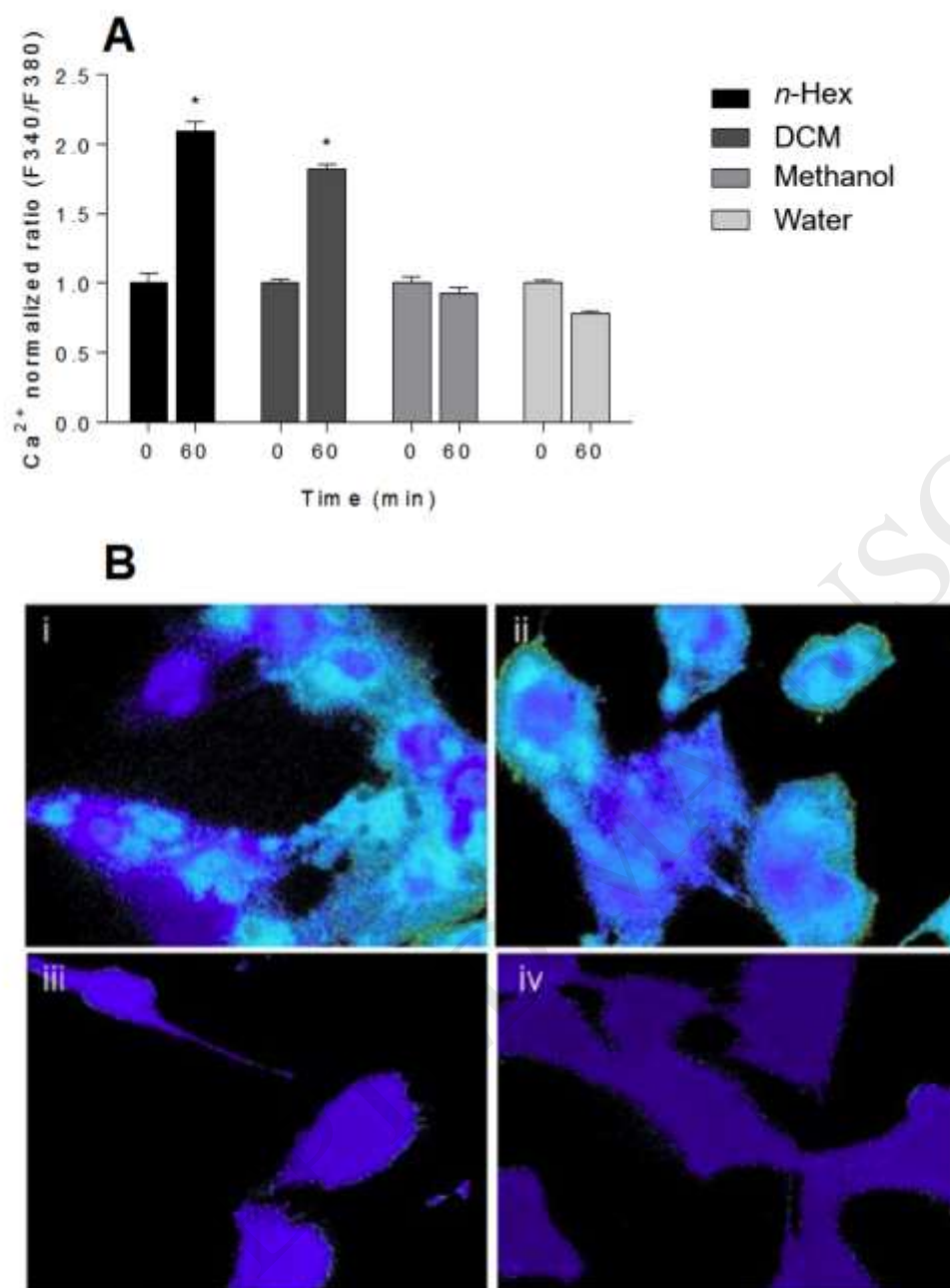


FIGURE 1.



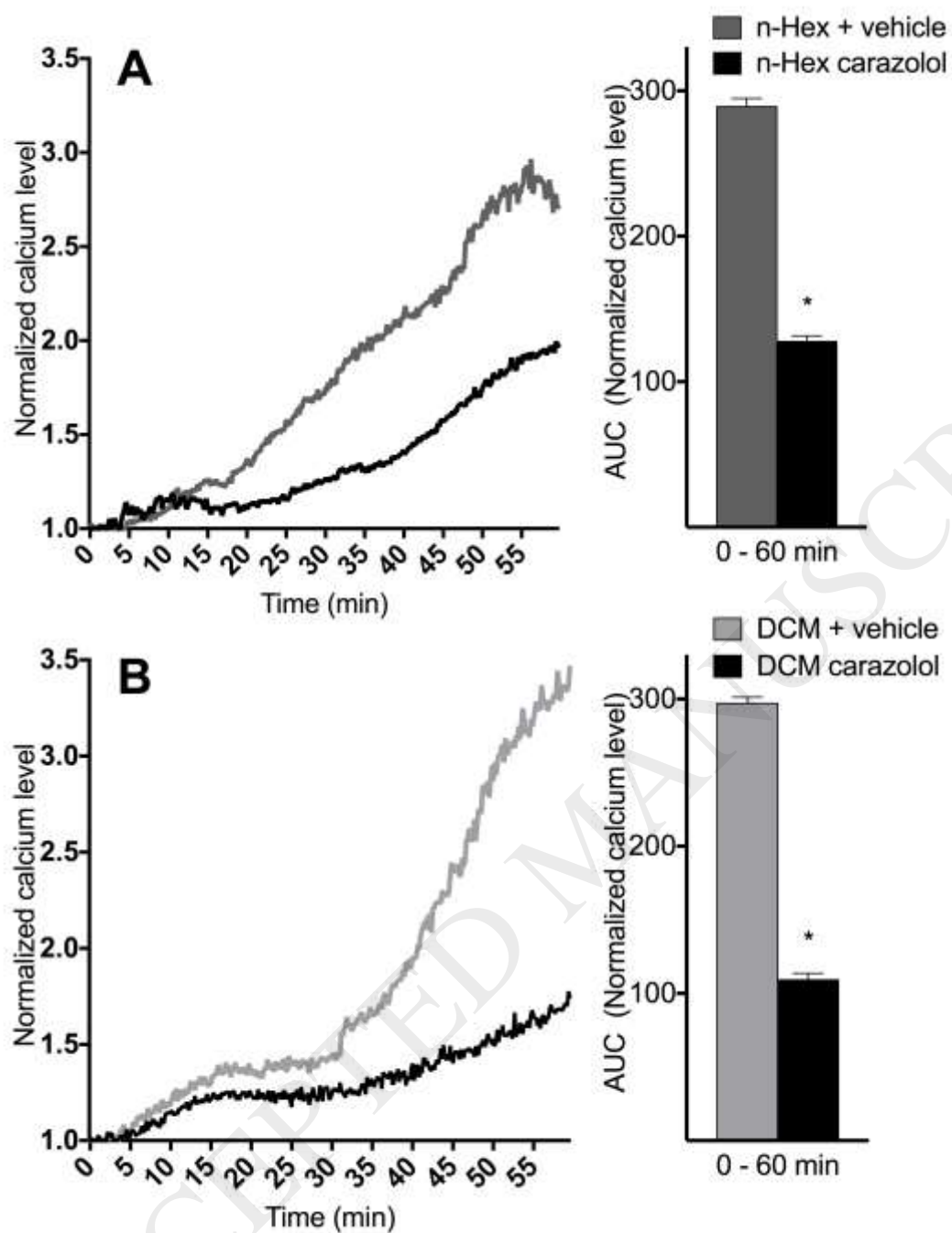


FIGURE 2

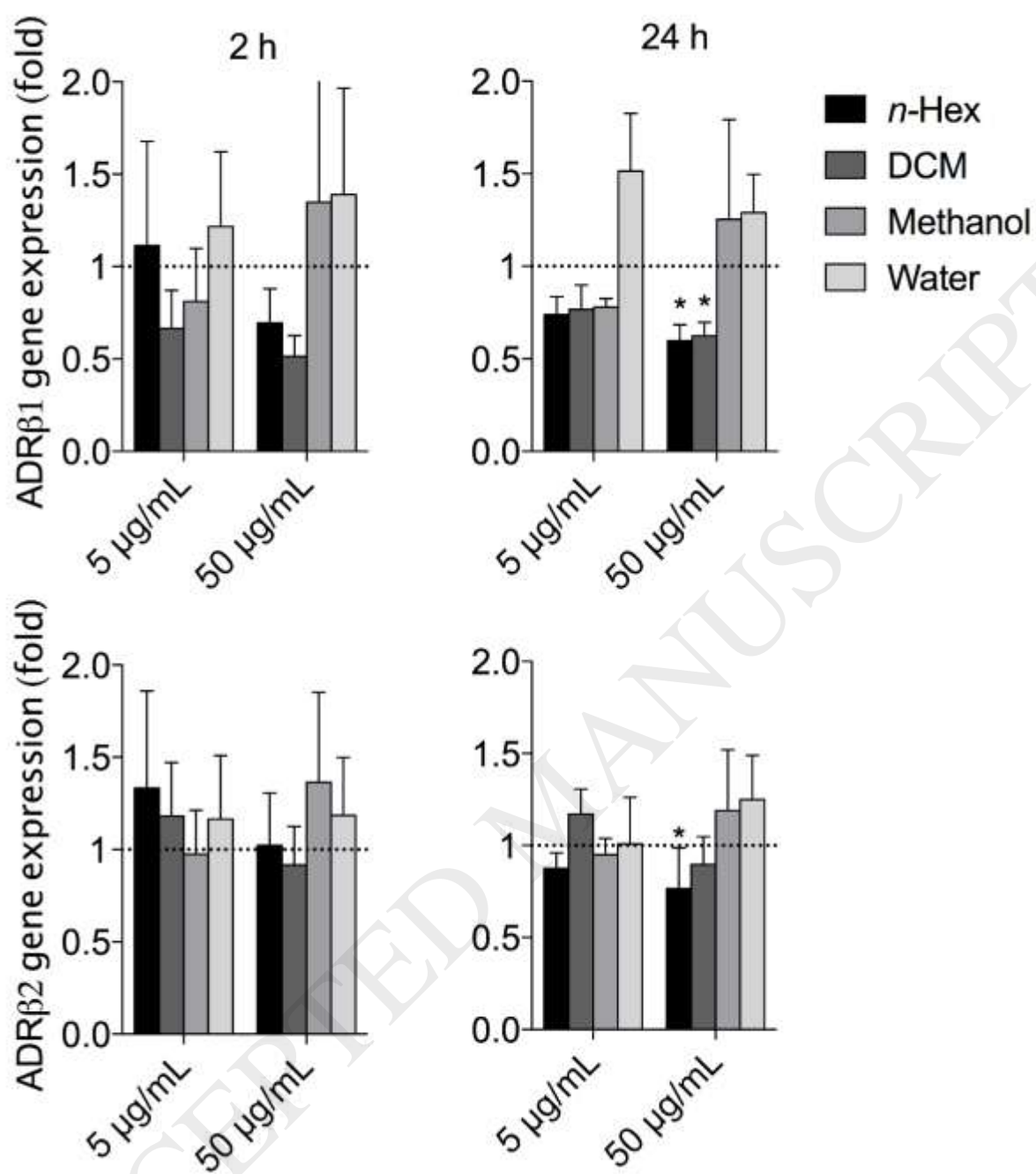


FIGURE 3

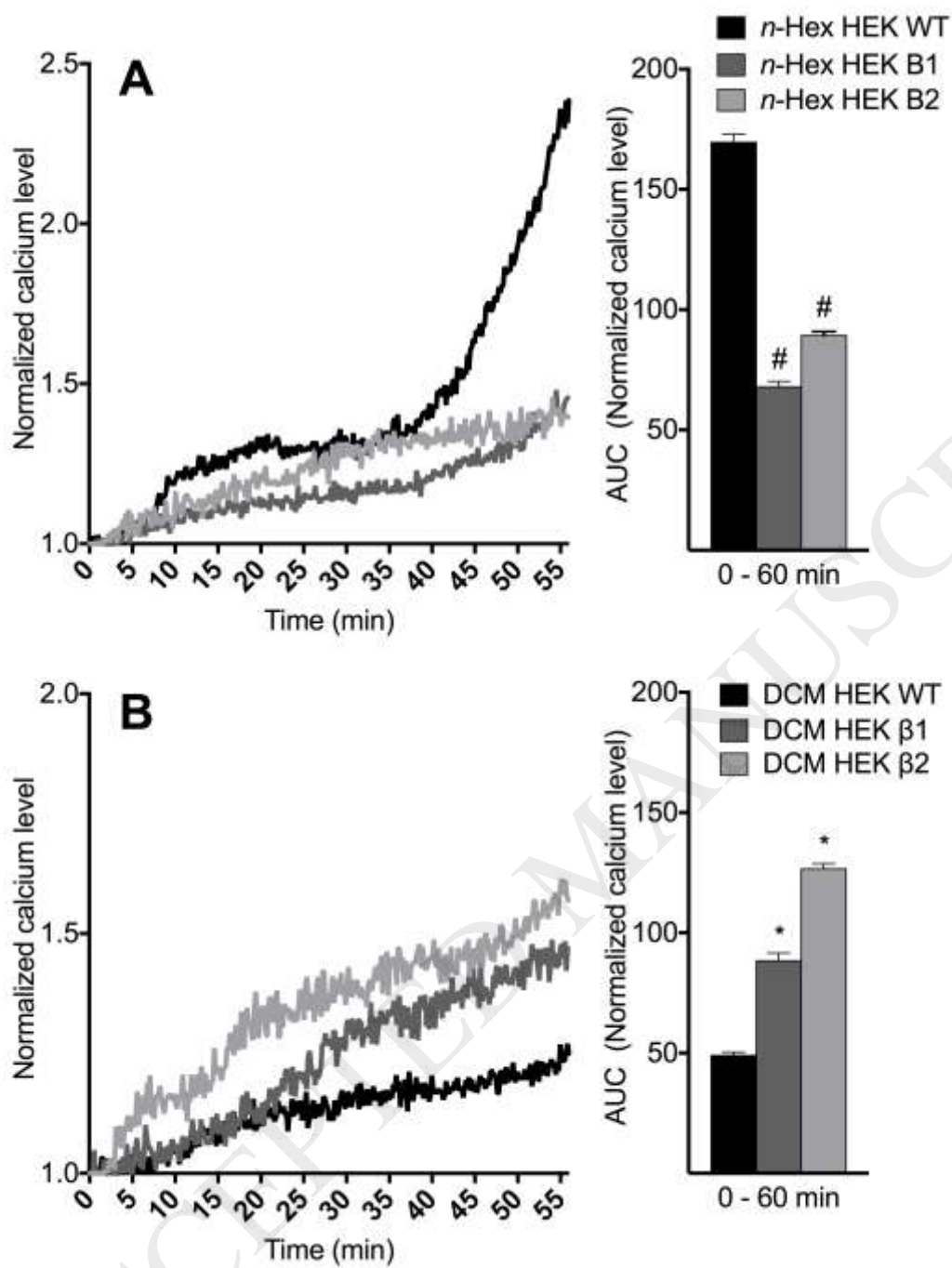


FIGURE 4.

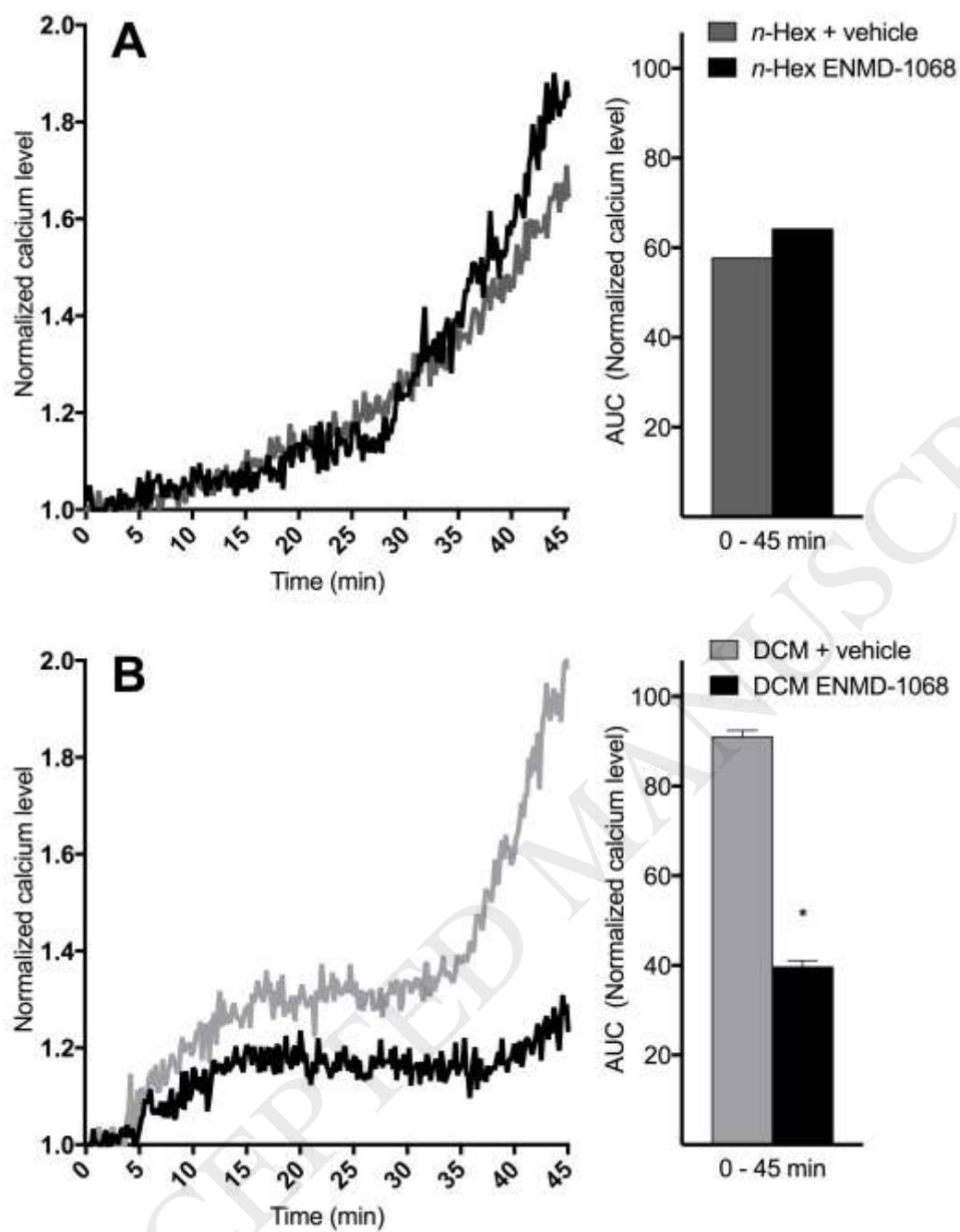


FIGURE 5

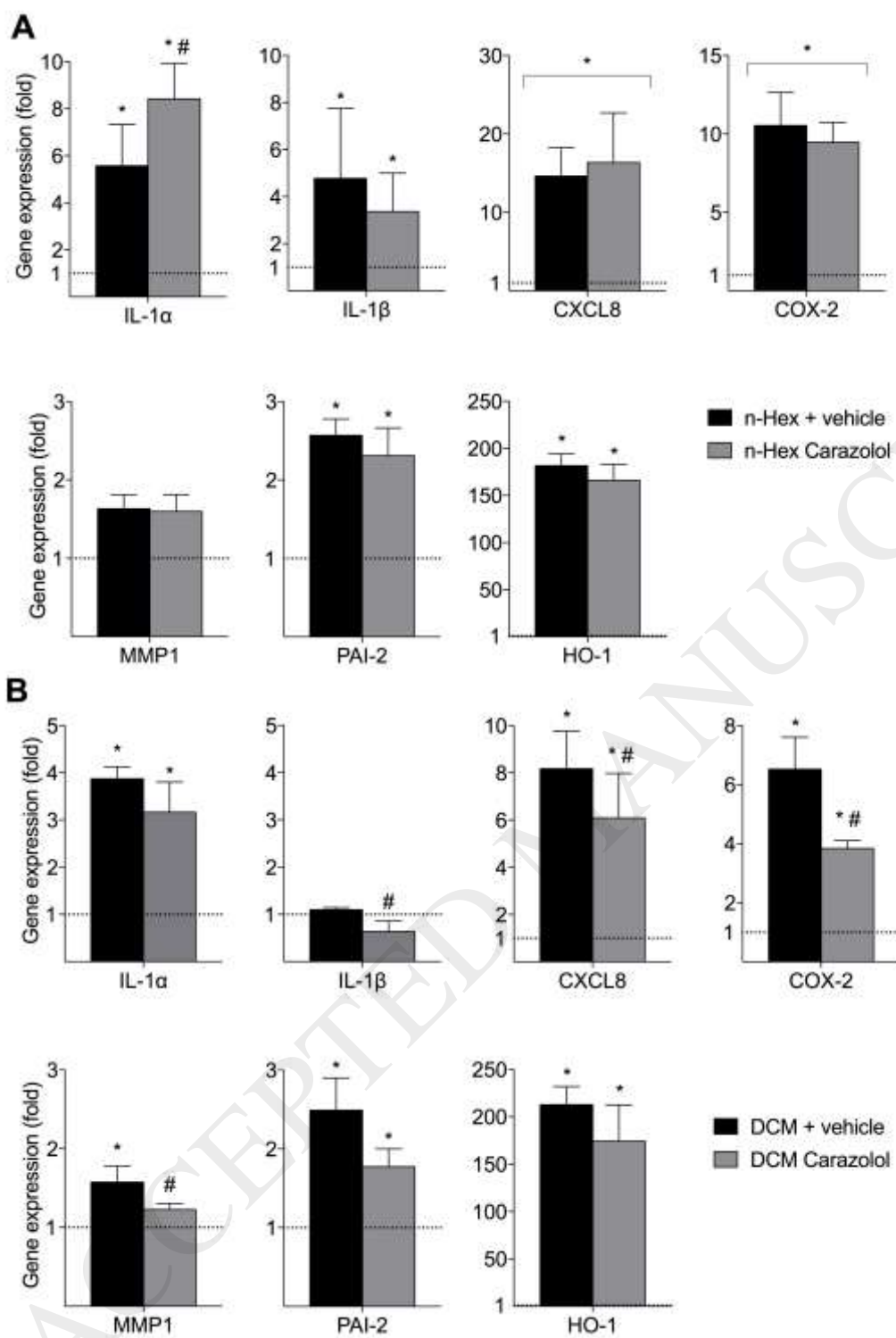


FIGURE 6

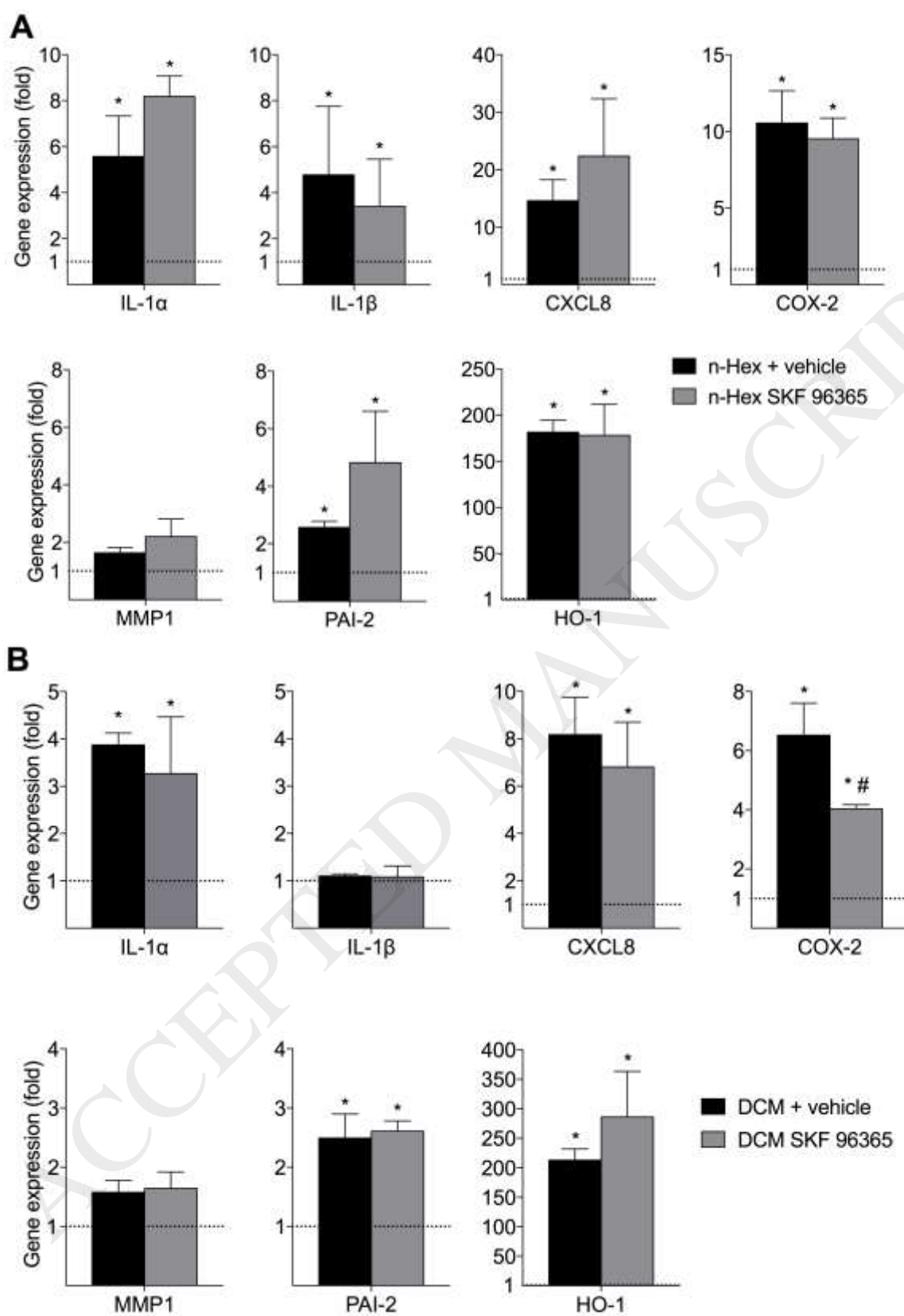


FIGURE 7