



**HAL**  
open science

## Update on prevalence and mechanisms of resistance to linezolid, tigecycline and daptomycin in enterococci in Europe Towards a common nomenclature

Jennifer K Bender, Vincent Cattoir, Kristin Hegstad, Ewa Sadowy, Teresa M Coque, Henrik Westh, Anette M Hammerum, Kirsten Schaffer, Karen Burns, Stephen Murchan, et al.

### ► To cite this version:

Jennifer K Bender, Vincent Cattoir, Kristin Hegstad, Ewa Sadowy, Teresa M Coque, et al.. Update on prevalence and mechanisms of resistance to linezolid, tigecycline and daptomycin in enterococci in Europe Towards a common nomenclature. Drug Resistance Updates, 2018, 40, pp.25-39. 10.1016/j.drup.2018.10.002 . hal-01940265

HAL Id: hal-01940265

<https://univ-rennes.hal.science/hal-01940265>

Submitted on 3 Dec 2018

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

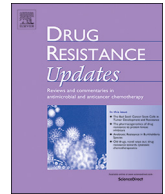


Distributed under a Creative Commons Attribution - NonCommercial - NoDerivatives 4.0 International License



Contents lists available at ScienceDirect

## Drug Resistance Updates

journal homepage: [www.elsevier.com/locate/drup](http://www.elsevier.com/locate/drup)

## Update on prevalence and mechanisms of resistance to linezolid, tigecycline and daptomycin in enterococci in Europe: Towards a common nomenclature

Jennifer K. Bender<sup>a</sup>, Vincent Cattoir<sup>b</sup>, Kristin Hegstad<sup>c</sup>, Ewa Sadowy<sup>d</sup>, Teresa M. Coque<sup>e</sup>, Henrik Westh<sup>f</sup>, Anette M. Hammerum<sup>g</sup>, Kirsten Schaffer<sup>h</sup>, Karen Burns<sup>i</sup>, Stephen Murchan<sup>j</sup>, Carla Novais<sup>k</sup>, Ana R. Freitas<sup>k</sup>, Luísa Peixe<sup>k</sup>, Maria Del Grosso<sup>l</sup>, Annalisa Pantosti<sup>l</sup>, Guido Werner<sup>a,\*</sup>

<sup>a</sup> Department of Infectious Diseases, Division of Nosocomial Pathogens and Antibiotic Resistances, Robert Koch Institute, Wernigerode, Germany

<sup>b</sup> Department of Clinical Microbiology and National Reference Center for Enterococci, University Hospital of Rennes, Rennes, France and Medical School and INSERM Unit U1230, University of Rennes, Rennes, France

<sup>c</sup> Norwegian National Advisory Unit on Detection of Antimicrobial Resistance, Department of Microbiology and Infection Control, University Hospital of North-Norway and Research group for Host-Microbe Interactions, Department of Medical Biology, Faculty of Health Sciences, University of Tromsø - The Arctic University of Norway, Tromsø, Norway

<sup>d</sup> Department of Molecular Microbiology, National Medicines Institute, Warsaw, Poland

<sup>e</sup> Ramón y Cajal Health Research Institute (IRYCIS), Ramón y Cajal University Hospital, Department of Microbiology, and Centro de Investigación Biomédica en Red de Epidemiología y Salud Pública (CIBER-ESP), Madrid, Spain

<sup>f</sup> Faculty of Health and Medical Sciences, Institute of Clinical Medicine, University of Copenhagen, Copenhagen and Department of Clinical Microbiology, Hvidovre Hospital, Hvidovre, Denmark

<sup>g</sup> Department for Bacteria, Parasites and Fungi, Infectious Disease Preparedness, Statens Serum Institut, Copenhagen S, Denmark

<sup>h</sup> Department of Clinical Microbiology, St Vincent's University Hospital, UCD School of Medicine, Dublin, Ireland

<sup>i</sup> Beaumont Hospital, Royal College of Surgeons in Ireland & Health Protection Surveillance Center, Dublin, Ireland

<sup>j</sup> Health Protection Surveillance Center, Dublin, Ireland

<sup>k</sup> UCIBIO/REQUIMTE, Departamento de Ciências Biológicas, Laboratório de Microbiologia, Faculdade de Farmácia, Universidade do Porto, Porto, Portugal

<sup>l</sup> Department of Infectious Diseases, Istituto Superiore di Sanità, Rome, Italy

## ARTICLE INFO

## Keywords:

vanA  
vanB  
VRE  
Linezolid  
Tigecycline  
Daptomycin

## ABSTRACT

Vancomycin-resistant enterococci (VRE) are important nosocomial pathogens. Invasive VRE infections are difficult to treat since common therapeutic options including ampicillin and glycopeptides often fail. *In vitro*, most VRE remain susceptible to last-resort antibiotics such as linezolid, tigecycline and daptomycin. However, neither tigecycline nor linezolid act in a bactericidal manner, and daptomycin has proven activity only at high dosages licensed for treating enterococcal endocarditis. Despite these pharmacological and therapeutic limitations, reports on resistance to these last-resort drugs in VRE, and enterococci in general, have increased in recent years. In this review, we briefly recapitulate the current knowledge on the mode of action as well as the known and novel mechanisms of resistance and describe surveillance data on resistance to linezolid, tigecycline and daptomycin in enterococci. In addition, we also suggest a common nomenclature for designating enterococci and VRE with resistances to these important last-resort antibiotics.

## Introduction

Enterococci are important commensal bacteria residing in the intestines of humans and many other animals, including invertebrates. They are known as early colonizers of neonates and constitute a substantial part of the healthy intestinal microbiota of adults. Due to their beneficial impact on gut health, enterococcal mixtures have been licensed as probiotics. Various enterococci are used for food fermentation

and preservation (Franz et al., 2011) due to their abilities to produce highly effective bacteriocins active against food-borne pathogens such as *Listeria monocytogenes*. The two species *Enterococcus faecium* and *Enterococcus faecalis* have been recognized for decades as major nosocomial pathogens in humans. Moreover, *E. faecalis* is occasionally implicated in community-associated infections such as urinary tract infections and endocarditis.

Members of the genus *Enterococcus* are known to be versatile

\* Corresponding author.

E-mail address: [WernerG@rki.de](mailto:WernerG@rki.de) (G. Werner).

<https://doi.org/10.1016/j.drup.2018.10.002>

Received 24 March 2018; Received in revised form 10 August 2018; Accepted 30 October 2018

1368-7646/© 2018 The Authors. Published by Elsevier Ltd. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

microorganisms capable of surviving under harsh conditions and quickly adapting to a changing environment. Their biological constitution allows for a high rate of DNA recombination against a background level of mutation, which then facilitates survival under varying situations at the population level. Within the *E. faecium* species, frequent DNA exchange associated with niche separation and niche adaptation have shaped a subpopulation, called clade A1, that has clearly adapted to the hospital setting (Lebreton et al., 2013). Different models exist that describe the origin of these hospital-associated *E. faecium* lineages (Lebreton et al., 2013; Numminen et al., 2016); however, it is still under debate as to whether today's hospital-associated *E. faecium* strains originated from commensal strains in humans and animals or, in fact, evolved directly from livestock animals (Lebreton et al., 2013; Raven et al., 2016; Willems et al., 2012). Niche adaptation and host speciation is much less pronounced in *E. faecalis*, although a number of clonal lineages exhibit an association with hospitals, to a certain extent (CC2 and CC9; (Kuch et al., 2012; Numminen et al., 2016; Tedim et al., 2015).

Currently, *E. faecium* and *E. faecalis* rank as the second and third most important nosocomial pathogens worldwide (Kolonen et al., 2017; Mendes et al., 2016a; Weisser et al., 2017; Zhang et al., 2017). Even though the pathogenicity of these species is controversial, enterococci can cause life-threatening infections in humans, mainly among elderly, multimorbid and/or immunocompromised patients (Arias and Murray, 2012; Murray, 1990). A wide armamentarium of natural resistance in this genus prevents a number of frequently used modern drugs such as cephalosporins and fluoroquinolones from being effective against enterococcal infections (Arias and Murray, 2012; Murray, 1990). Whereas ampicillin and glycopeptides are important first-line therapeutics, almost all hospital-associated strains of *E. faecium* have acquired ampicillin resistance (Lester et al., 2008; Top et al., 2007; Weisser et al., 2012). Moreover, since the mid-1980s, enterococci have garnered increased medical and public attention when vancomycin resistance first appeared in clinical isolates of *E. faecium* and *E. faecalis* (vancomycin-resistant enterococci; VRE) (Johnson et al., 1990; Uttley et al., 1989). Increased prevalence of ampicillin and vancomycin resistance, mostly among hospital-associated *E. faecium* isolates, has led to the therapeutic dilemma of multidrug-resistant (MDR) pathogens. Hence, these isolates, along with other superbugs such as *Staphylococcus aureus*, *Klebsiella pneumoniae*, *Acinetobacter baumannii*, *Pseudomonas aeruginosa* and *Enterobacter* species, have been dubbed the “ESKAPE” pathogens (Boucher et al., 2009). Several reports and reviews have assessed rates of VRE among clinical enterococcal isolates and in specific clinical indications (such as bloodstream infections) (Adams et al., 2016; Akpaka et al., 2017; Simner et al., 2015; Weisser et al., 2017; Werner et al., 2008a). In general, the number of infections caused by enterococcal isolates, mainly *E. faecium*, and of those that are vancomycin resistant, has increased in many industrialized countries all over the world. Consequently, VRE are also considered a significant antibiotic resistance threat, leading to in excess of 20,000 drug-resistant *Enterococcus* infections and approximately 1300 deaths due to VRE in the US each year (CDC 2013, <https://www.cdc.gov/drugresistance/pdf/ar-threats-2013-508.pdf>). Through participation in the European Antimicrobial Resistance Surveillance Network (EARS-Net), coordinated by the European Center for Disease Prevention & Control (ECDC), many European countries have experienced an increased proportion of ampicillin resistant enterococci in invasive infections, mainly *E. faecium*, which are also resistant to vancomycin (Tedim et al., 2017). However, the European-wide trend of VRE causing systemic infections has remained inconsistent ([https://ecdc.europa.eu/sites/portal/files/documents/antibiotics-EARS-Net-summary-2016\\_0.pdf](https://ecdc.europa.eu/sites/portal/files/documents/antibiotics-EARS-Net-summary-2016_0.pdf)). Ireland has been reporting the highest rate of vancomycin resistance among *E. faecium* bloodstream isolates in Europe in the last 10 years. Since 2012, more than 40% of *E. faecium* bloodstream isolates have been identified as vancomycin-resistant.

In the hospital setting, most VRE are also resistant to first line

antibiotics (ampicillin, quinolones) with high MIC values. Furthermore, *E. faecium* and *E. faecalis* are often high-level resistant to aminoglycosides (HLAR). Hence, only a few therapeutic options, including oxazolidinones (linezolid, tedizolid), novel tetracyclines (tigecycline) and lipopeptides (daptomycin) remain to treat infections with multidrug-resistant VRE (Barber et al., 2015; Mutters et al., 2015; Whang et al., 2013; Zhao et al., 2016). The increasing prevalence of VRE in many countries and health-care settings, the expanded use of such last-resort antibiotics, and the ability of enterococci to easily acquire novel DNA fragments encoding resistance determinants currently enhance the selective pressure for developing resistance to these compounds. The genetic constitution of enterococci with a high recombination rate and the frequent occurrence of mobile genetic elements within the genome, as well as their natural habitat, predestine enterococci as a suitable indicator for novel emerging resistance phenotypes. We herein aim to summarize the general knowledge on non-susceptibility to last-resort antibiotics in *Enterococcus* spp. with a special focus on linezolid, tigecycline and daptomycin resistance. There are international surveillance systems that monitor the *in vitro* activities of these antimicrobial compounds, such as T.E.S.T. (Tigecycline Evaluation and Surveillance Trial) (Marco and Dowzicky, 2016; Vega and Dowzicky, 2017). They further serve as a basis for comprehensive databases from which resistance development can be analyzed retrospectively on a global scale (ATLAS, Antimicrobial Testing Leadership And Surveillance; supported by Pfizer). For the vast majority of the European countries, a generally low prevalence (< 1%) of isolates resistant to last-resort antibiotics has been noted (ATLAS database; <https://atlas-surveillance.com>). For this review article, we have comprehensively compared susceptibility data on enterococci with regard to linezolid, tigecycline and daptomycin and have summarized recent advances in studying the underlying resistance mechanisms in order to propose a common nomenclature for VRE with resistance to last-resort antibiotics.

#### Vancomycin resistance in *Enterococcus* spp.

Although this is not in the main focus of this report, we provide a short overview of the current state of knowledge of the different genotypes of vancomycin resistance in enterococci. At present, eight types (VanA to VanN) of vancomycin resistance are known, named according to its ligase gene (*vanA* to *vanN*). In addition to the ligase gene, two other genes encoding the sensor (*VanS*) and response regulator (*VanR*) are essential to confer the corresponding phenotype. Glycopeptide antibiotics bind to the N-terminal D-Ala-D-Ala residue present in disaccharide-pentapeptide cell wall precursors. The interplay between several expressed resistance genes allows for the downregulation of housekeeping cell wall precursor synthesis and the production of an alternative pentapeptide (the so-called pentadepsipeptide) side chain ending with D-lactate (e.g., VanA, VanB) or D-serine (e.g., VanG) (Courvalin, 2006). This alternative ending exhibits an up to 1,000-fold lower binding capacity to vancomycin (in case of D-Ala-D-Lac), which results in clinical resistance. The *vanA* and *vanB* resistance genotypes are by far the most frequently found variants in clinical *E. faecium* and *E. faecalis* isolates worldwide. The *vanA* gene cluster (carried by transposon Tn1546) is mainly plasmid-located, whereas the *vanB* gene cluster (carried by members of the integrative conjugative elements Tn1549-Tn5382) is usually present on the bacterial chromosome. For more details, the reader is referred to a number of review articles dedicated to this topic (Courvalin, 2006; Hegstad et al., 2010; Werner et al., 2013).

A specific variant of VRE is represented by vancomycin-variable enterococci (VVE); these strains were noticed only very recently. VVE are vancomycin-susceptible enterococci with a *vanA* genotype (VVE-S), which can become resistant to vancomycin (VVE-R) upon exposure to vancomycin or teicoplanin (Coburn et al., 2014; Sivertsen et al., 2016). Development of vancomycin resistance in VVE-S can arise from different mechanisms and result in the expression of *vanHAX* genes, in

either a restored vancomycin-inducible fashion (Sivertsen et al., 2016) or constitutively (Thaker et al., 2015). Outbreaks of VVE have been reported in Canada (Szakacs et al., 2014), Norway (Sivertsen et al., 2016) and Denmark (Hansen et al., 2018). Sporadic isolates of VVE have also been reported in other European countries (Jung et al., 2014; Loens et al., 2016; Sivertsen et al., 2017). Most of the reported VVE isolates have been *E. faecium*, but this phenomenon can also occur in *E. faecalis* (Sivertsen et al., 2016). The overall prevalence of VVE-S is unknown. Since the expression of vancomycin resistance genes is turned off in VVE-S, they are not easily detected with phenotypic methods. However, since vancomycin and teicoplanin exposure over days promote the development of resistance, VVE have caused treatment failure and could be considered an emerging threat (Downing et al., 2015; Sivertsen et al., 2016).

**Last-resort antimicrobials to treat VRE infections**

*Oxazolidinones*

*Linezolid: mode of action*

Linezolid (LZD) is the first representative of the oxazolidinones that received approval for clinical practice by the US Food and Drug Administration (FDA) in 2000 (and later by the European Medicines Agency, EMA). This bacteriostatic antibiotic exhibits activity against a broad range of Gram-positive bacteria, including Methicillin-resistant *Staphylococcus aureus* (MRSA), and has received licensing for the treatment of nosocomial pneumonia, complicated or uncomplicated skin and soft tissue infections (SSTIs) and for infections caused by VRE (Brickner et al., 2008; Plosker and Figgitt, 2005). Although it adds to the substances that inhibit bacterial protein synthesis, cross-resistance triggered by antibiotics with proximal target sites have not been described, which is most likely due to the distinct mechanism of LZD action (Fines and Leclercq, 2000; Kloss et al., 1999). By targeting bacterial protein translation, oxazolidinones prevent the formation of the initiation complex by binding to domain V of the 23S rRNA (Swaney et al., 1998; Wilson et al., 2008). Nevertheless, during a compassionate use program and following implementation in clinical daily routine, the detection of LZD-resistant bacteria has been reported, including those that emerged during LZD therapy (Bassetti et al., 2003; Birmingham et al., 2003; Krull et al., 2016; Seedat et al., 2006). LZD is the only antibiotic of last resort discussed in this overview that can be given also orally in addition to an intravenous application suggesting a wider selective pressure, for instance, also on bacteria with an intestinal reservoir.

*Linezolid: mechanisms of resistance*

Resistance to LZD (MIC > 4 mg/L; [www.eucast.org](http://www.eucast.org)) can be mediated by either mutational mechanisms or by gene acquisition (summarized in Table 1). The most common resistance mechanism in enterococci is based on a G-to-T conversion at position 2576 (according to *E. coli* K-12 numbering) within the genes encoding for 23S ribosomal RNAs (Klare et al., 2015; Mendes et al., 2014; Pfaller et al., 2017b). The LZD MIC increases with the copy number of mutated 23S rDNA alleles and LZD resistance in enterococcal isolates is rapidly selected under LZD therapy (Bassetti et al., 2003; Marshall et al., 2002; Seedat et al., 2006). The initial 23S rDNA mutation is a crucial step in resistance progression, as additional 23S rDNA mutation sites in the genome are most likely generated by recombination rather than by individual point mutations (Marshall et al., 2002). The classical G2576 T mutations also seem to determine resistance to tedizolid, the second clinically available oxazolidinone (Klupp et al., 2016). Acquired resistance-determining mutations appear to be stable, as 40 to 50 daily passages of a clinical LZD-resistant *E. faecium* isolates in non-selective media (ca. 300 bacterial generations) were not sufficient to revert resistance associated with heterozygous G2576 T mutations (Freitas et al., 2016; Klare et al., 2015). For one of these experiments, *E. faecium* strains (n = 3) were

**Table 1**  
Overview of acquired mutations/genes possibly contributing to linezolid resistance in enterococci.

Species	Target (gene)	Function of target	Comment	References
<i>E. faecalis</i> / <i>E. faecium</i>	23S rDNA	Ribosomal RNA	G2576 T mutation Initial point mutation, most likely followed by further allele replacement due to homologous recombination MIC levels depend on the number of alleles affected	(Klare et al., 2015; Marshall et al., 2002; Mendes et al., 2014; Pfaller et al., 2017a)
	RplC/RplD Cfr, Cfr (B), Cfr (B) variants OprA	Ribosomal proteins 23S rRNA methyltransferase ABC-F protein	Expression of <i>cfr</i> does not necessarily result in LZD insusceptibility	(Chen et al., 2013) (Bender et al., 2016; Cercenado et al., 2010; Deshpande et al., 2015; Liu et al., 2014a, 2012; Long et al., 2006) (Brenciani et al., 2016; Cai et al., 2016; Cavaco et al., 2017; Cui et al., 2016; He et al., 2016; Huang et al., 2017; Li et al., 2016; Sharkey and O'Neill, 2018; Vorobieva et al., 2017; Wang et al., 2015) (Antonelli et al., 2018) (Hua et al., 2018)
	PoxA DEGs	ABC-F protein Upregulation of genes associated with efflux pumps and biofilm formation		

isolated from a Portuguese patient taking linezolid for 61 days, who developed three variants of the same LZD-resistant clone presenting different antibiotic resistance and virulence profiles. Yet another mechanism based on nucleotide mutations has been described for genes encoding for ribosomal proteins. Here, mutations in *rplC* (L3) and *rplD* (L4) are predominantly linked to LZD resistance in enterococci that lack G2576 T mutations and that do not express the multidrug resistance gene *cfr* (Chen et al., 2013).

Cfr was first isolated from *Staphylococcus sciuri* of animal origin and soon received enormous attention, as it is capable of conferring resistance to at least five classes of antibiotics, including linezolid (but not tedizolid); it has been detected in various bacterial species and genera (Locke et al., 2014; Long et al., 2006; Schwarz et al., 2000; Shen et al., 2013). The resistance phenotype mediated by the Cfr methyltransferase is commonly referred to as PhLOPS<sub>A</sub>, an acronym for resistance to the antibacterial agents phenicol, lincosamide, oxazolidinone, pleuromutilin and streptogramin A (Long et al., 2006). Most importantly, the transfer of plasmid-borne *cfr* has been demonstrated to occur even across bacterial species and genera (Bender et al., 2016; Cafini et al., 2016).

Regarding enterococci, *cfr* was first reported from *E. faecalis* of animal origin (Liu et al., 2012). Follow-up studies in enterococci identified several distinct conjugative plasmids that harbored the multidrug resistance gene (Liu et al., 2013, 2012). Transfer of the *cfr*-harboring plasmids was demonstrated and the wildtype *cfr* (staphylococci homolog) and derivatives thereof, named *cfr(B)* and *cfr(B)* variants, located on plasmids or within the bacterial chromosome emerged in *Enterococcus* spp. from animal and human samples alike (Bender et al., 2016; Deshpande et al., 2015; Diaz et al., 2012; Liu et al., 2014a, 2013; Patel et al., 2013). It is worth noting that *E. faecium* isolates containing the *cfr* gene are increasingly observed (Cercenado et al., 2010). Nevertheless, the contribution of *cfr* and *cfr(B)* variants to elevated LZD MICs in *Enterococcus* spp. is still under debate. It has been shown that, probably due to yet unknown strain-specific reasons, Cfr and Cfr(B) variants fail to demonstrate LZD resistance and the entire PhLOPS<sub>A</sub> phenotype in enterococci (Bender et al., 2016; Liu et al., 2014b).

Further studies on enterococci with a silent *cfr* gene, but exhibiting LZD resistance, have demonstrated a role for a second LZD (and tedizolid) resistance determinant, the oxazolidinone and phenicol transferable resistance A (OptrA) (Brenciani et al., 2016; Wang et al., 2015). First detected in *E. faecalis* of human origin, further studies have been published reporting *optrA* in isolates of *E. faecalis* and *E. faecium* and most recently also in *S. aureus*, *S. sciuri* and *Streptococcus suis* (Cai et al., 2016; Cui et al., 2016; Guo et al., 2018; He et al., 2016; Huang et al., 2017; Li et al., 2016). Surveillance studies from China have shown that *optrA* is detected more frequently in enterococci from food-producing animals than in humans (Wang et al., 2015). *OptrA* encodes for an ATP-binding cassette (ABC)-F protein and mediates resistance through target protection (Sharkey and O'Neill, 2018). Releasing ribosome-targeted antibiotics such as phenicols, compounds commonly used in the animal production, highlights the relevance of selected antibiotic usage in animal husbandry for the emergence of bacteria carrying this resistance determinant (Wang et al., 2015). Moreover, reports from animals raised in Korea, Tunisia and Colombia or residual wastewaters in Tunisia where *E. faecalis* isolates with *optrA* have been described pointed towards an important animal reservoir as well as the possibility of ongoing environmental dissemination of this resistance gene [(Cavaco et al., 2017; Tamang et al., 2017) A.R.F. and L.P., unpublished data]. Recently, the analysis of 26 *E. faecalis* isolates from humans with LZD MIC > 4 mg/L obtained from the SENTRY program (see above) revealed a high prevalence (53.5%) of *optrA*-positive *E. faecalis* from humans; however, not all of these isolates exhibited LZD resistance (Mendes et al., 2016b). Congruently, the German National Reference Center (NRC) for staphylococci and enterococci at the Robert Koch Institute (RKI) have observed an increase in *optrA*-positive and LZD-resistant clinical (human) *E. faecalis* from 2007 to 2016, with 90.9% of

all LZD-resistant *E. faecalis* (n = 35) isolates being *optrA*-positive in 2016 (Bender et al., 2018). Moreover, all LZD-resistant *E. faecalis* isolates reported to the Norwegian National Advisory Unit on Detection of Antimicrobial Resistance since December 2015 harbored *optrA* (Hegstad et al., unpublished). In a study on Polish LZD-resistant enterococci (Gawryszewska et al., 2017), two *E. faecalis* strains were positive for *optrA* with the resistance determinant located on a transferable plasmid identical to pE394, initially reported in China (Wang et al., 2015). With respect to the One Health dimension, *optrA* has been reported from an ST22 *E. faecium* strain isolated from imported turkey meat and an ST22 *E. faecalis* strain isolated from Danish veal meat (Cavaco et al., 2017). Likewise, an ST16 *E. faecalis* isolate with *optrA* has been reported in a Danish patient (Vorobieva et al., 2017).

Exceeding what is known for *cfr*, a high diversity of *optrA* nucleotide sequences and a plethora of variable genetic environments embedding the resistance gene in either the chromosome or on diverse plasmids and/or within transposon structures (Tn554, Tn558, flanked by IS1216 or similar IS elements) have been reported (Bender et al., 2018; He et al., 2016; Huang et al., 2017; Mendes et al., 2016b). Hence, rapid spread of the resistance locus is conceivable with the possibility of global distribution being underestimated to date.

Recently, another member of the ABC-F protein subfamily, PoxTA, has been discovered in MRSA of clinical origin, clostridia and enterococci, exerting its protective effect against phenicols, oxazolidinones and tetracyclines (Antonelli et al., 2018). The prevalence of *poxTA* amongst the enterococcal population is under current investigation.

Although insufficiently studied, cell wall thickness and biofilm formation were discussed as putative alternative resistance pathways to acquire LZD resistance (Tian et al., 2014); however, the authors had no knowledge of *optrA* or *poxTA* at the time when the article was published and thus did not screen for this particular resistance loci. Also, it remains to be determined as to how the combination of determinants and/or genetic background with respect to species impacts LZD MICs. A first report by Mendes and co-workers and as also observed by the NRC for Staphylococci and Enterococci in Germany and the Norwegian National Advisory Unit on Detection of Antimicrobial Resistance demonstrated that the frequency of particular LZD resistance mechanisms seems to differ between *E. faecalis* and *E. faecium*. A tendency of 23S rDNA mutation being mostly common in *E. faecium* and *optrA* dominating in *E. faecalis* was generally observed (Mendes et al., 2016b).

A plethora of LZD resistance mechanisms already exists in enterococci, and novel ones have emerged relatively rapidly. Hence, it is conceivable that additional factors might be discovered with the potential to equally contribute to infections of and outbreaks caused by linezolid-resistant enterococci (LRE). In fact, LRE without any of the currently known resistance mechanisms have been reported, both associated with infection and colonization (A.R.F. and L.P., pers. communication; J.K.B. and G.W.; pers. communication; K.H. pers. communication). Recently, Hua et al. demonstrated differential expression of genes related to efflux pumps and or biofilm formation in strains showing low-level resistance to LZD and lacking known resistance mechanisms using a whole transcriptomic approach (Hua et al., 2018). Alarmingly, gene acquisition or mutational changes do not seem to impact bacterial fitness, as Hegstad and colleagues noticed the persistence of an ST117 LZD-resistant *E. faecium* clone in a Scandinavian hospital for over one year (Hegstad et al., 2014). Likewise, acquisition of an *optrA*-plasmid by *E. faecalis* did not affect the growth rates of the transconjugant compared to the plasmid-free recipient (Gawryszewska et al., 2017).

Thus, prudent use of LZD is indispensable to prevent the emergence and dissemination of multidrug-resistant strains. Again, it must be stressed that the occurrence of enterococci susceptible to linezolid but carrying silent *cfr* and/or *optrA* might hamper their identification in a clinical setting (Brenciani et al., 2016). The potential to disseminate a highly promiscuous resistance locus to other bacteria imposes an

unpredictable risk to different sectors.

#### Linezolid: resistance surveillance data

Infections due to LRE and their nosocomial spread have been described in several case reports and outbreak investigations for both vancomycin-resistant and vancomycin-susceptible *E. faecium* and *E. faecalis* (Gomez-Gil et al., 2009; Gonzales et al., 2001; Hegstad et al., 2014; Herrero et al., 2002; Ntokou et al., 2012; O'Driscoll et al., 2015; Rahim et al., 2003). LZD exposure, prolonged antibiotic treatment, hospitalization in a hematological ward, immunodeficiency, prior surgery and patient-to-patient transmission have been identified as risk factors to acquire LRE infections (Hayakawa et al., 2012; Kainer et al., 2007; Scheetz et al., 2008; Souli et al., 2009). However, LZD resistance does not necessarily only emerge upon LZD therapy (Mutters et al., 2015).

Several national and international programs exist that started to monitor LZD resistance development in the early 2000s. The SENTRY Antimicrobial Surveillance Program, the Linezolid Experience and Accurate Determination of Resistance (LEADER) initiative, ZAAPS (Zyvox® Annual Appraisal of Potency and Spectrum) and T.E.S.T. concordantly report a sustained high potency of LZD against the entire spectrum of tested bacteria with very low rates of resistance development over the last two decades (Flamm et al., 2016; Mendes et al., 2016b, c; Mutnick et al., 2003; Pfaller et al., 2017a, b; Streit et al., 2015). As for other Gram-positive bacteria, the general prevalence of LRE still remains below 1%. It is also worth noting that no MIC creep was observed amongst the isolates tested (Mendes et al., 2016c; Streit et al., 2015). In a very recent review, enterococcal data from the US LEADER and the global ZAAPS studies were summarized, again confirming the overall low prevalence of < 1% LZD resistance among clinical *Enterococcus* spp. (Bi et al., 2017). The presence of common resistance mechanisms was also analyzed, demonstrating the presence of *cfr* in 4.7% of linezolid-resistant *E. faecalis* and in 4.8% of linezolid-resistant *E. faecium* isolates. Altogether, 32 cases of *optrA*-positive enterococci were found.

Specialized reference laboratories throughout Europe have reported a greater recognition of LRE. However, differences in resistance rates might also depend on the species analyzed; for instance, a recent report from China described that the occurrence of LZD non-susceptibility was greater in *E. faecalis* compared to *E. faecium* (Tian et al., 2014).

In Germany, the national resistance surveillance scheme ARS (<https://ars.rki.de>) has not observed any trend for linezolid resistance among clinical *E. faecalis* or *E. faecium* isolates in recent years (< 1%). However, the NRC recognized an increased frequency of LRE from < 1% in 2008 to > 9% in 2014 among all strains received annually for subsequent strain characterization on a voluntary basis (Klare et al., 2015).

Since the first finding of LZD-resistant *E. faecalis* in 2012 (Thilesen et al., 2014), only 20 LRE have been reported in Norway up to January 2, 2017 (<https://unn.no/fag-og-forskning/k-res/betydelig-oktforekomst-av-linezolidresistente-enterokokker-i-2016-i-norge>). Half of these isolates were found in 2016, representing a significant increase in prevalence compared to previous years.

In Denmark, LRE have not been sent to the antimicrobial reference laboratory on a routine basis. However, during 2016, 14 LZD-resistant *E. faecalis* (1.3%) and 27 LZD-resistant *E. faecium* (4.2%) were detected from Rigshospitalet, in the capital region in Denmark (DANMAP 2016). In the first nine months of 2017, the incidence of LZD-resistant *E. faecalis* was reduced at Rigshospitalet to 0.5%, and that of LZD-resistant *E. faecium* to 1.7%. This correlated with a reduction program for linezolid usage at Rigshospitalet (J. D. Knudsen, pers. communication). In 2017, two other Copenhagen hospitals had three isolates of LZD-resistant *E. faecium* (0.1%) (H.W., pers. communication).

In Spain, an *E. faecalis* clinical isolate was the first identified LRE in 2003 (Sanchez-Gomez et al., 2006). Since then, LRE, either *E. faecalis* or *E. faecium*, have been detected in different hospitals, mostly associated

with the use of this antibiotic in neutropenic patients (Alonso et al., 2014; Sanchez-Diaz et al., 2014), eventually causing nosocomial outbreaks (Gomez-Gil et al., 2009). Data from T.E.S.T. for tigecycline and comparator compounds in Spain, which included isolates from 27 centers obtained from 2004 to 2014 (Marco and Dowzicky, 2016), reflected linezolid resistance of 0.1% for *E. faecalis* (MIC<sub>90</sub> = 2; range 0.5–8 mg/L) and 0.6% for *E. faecium* (MIC<sub>90</sub> = 2, range 0.5–16 mg/L). Another collaborative study performed in Spain from 2015, including data from 23,907 patients in 200 intensive care units as part of a strategic program to reduce the risk of selection and spread of antibiotic resistance in this country (Plan Nacional de resistencias a los antibióticos, Programa PRAM, <http://www.resistenciaantibioticos.es>), documented the occurrence of resistance to linezolid in 1.5% of *E. faecalis* and 3.3% of *E. faecium* strains (Palomar et al., unpublished data). Finally, the first phenotypically LZD-resistant *E. faecalis* strain in wild animals was documented in Spain; however, LZD resistance was not confirmed by a second, independent method (Navarro-Gonzalez et al., 2013).

When analyzing T.E.S.T. data, no LRE were detected in Italy between 2012 and 2014 (Stefani and Dowzicky, 2016). The international ZAAPS program assessing LZD resistance rates in 32 countries reported in 2015 a single Italian *E. faecium* isolate with a confirmed resistance phenotype and a known G2756 T mutation (see below) (Pfaller et al., 2017a). The national Italian Surveillance system AR-ISS has reported, amongst others, linezolid resistance in isolates from bloodstream infections. Linezolid resistance was detected in 0.6% (n = 9/1,582) of *E. faecalis* strains in 2015 and 0.3% (n = 5/1,579) in 2016 and in 0.4% (n = 3/745) of *E. faecium* strains in 2015 and 1.1% (n = 10/930) in 2016.

Poland obtained the first LZD-resistant isolate of *E. faecium* as early as in 2003 (Krawczyk et al., 2004). Since 2012, the National Reference Center for Susceptibility Testing (NRCST) has witnessed both increasing numbers of strains that were received by the NRCST as well as centers that reported the occurrence of LRE. An analysis performed on 50 isolates in the period from 2008 to 2015 revealed the predominance of hospital-associated *E. faecium* (82%); LZD-resistant *E. faecalis* and a single isolate of *E. avium* were identified as well (Gawryszewska et al., 2017). In that study, the G2576 T mutation was the most prevalent mechanism of resistance, but isolates with *optrA* were also observed.

Out of 3,974 enterococcal clinical isolates sent to the French NRC from 2006 to 2016, only 9 (0.2%) were resistant to linezolid (MIC > 4 mg/L; V.C. unpublished data). The first isolate (found in 2010) possessed ribosomal mutations, whereas the more recent isolates (one in 2013, one in 2014, two in 2015 and four in 2016) harbored the plasmid-mediated *optrA* gene. Five strains isolated before 2016 were all *vanA*-positive *E. faecium*, whereas strains isolated in 2016 were all susceptible to vancomycin (one *E. faecium* and three *E. faecalis*).

In Ireland, LZD resistance in *E. faecium* blood culture isolates was reported for the first time in 2004, when four of 78 *E. faecium* isolates tested against LZD were reported as being resistant (5.2%). In subsequent years (2005–2016), LZD resistance continued to be reported from blood culture isolates. Overall, among 4073 isolates tested against LZD from 2002 to 2016, 75 were found to be resistant to LZD (1.8%). Of 3100 *E. faecalis* isolates tested against LZD between 2002 and 2016, 40 were found to be resistant (1.0%). LZD resistance in VRE-*E. faecium* isolates from sites other than the bloodstream, along with outbreaks of LZD-resistant VRE-*E. faecium* in tertiary hospitals, have also been reported in Ireland since 2012. Surveillance data in one institution showed an increase in LZD resistance in VRE-*E. faecium* from four isolates in 2015 to 16 isolates in 2016 from both clinical and screening specimens. The international ZAAPS program reported an Irish *E. faecalis* isolate from 2015 with LZD resistance and an *optrA* gene (Pfaller et al., 2017a).

In Portugal, available data from three major hospitals in the north and south of the country from 2009 to 2016 showed the occurrence of 35 *Enterococcus* spp. with LZD resistance. In each hospital, the incidence

**Table 2**  
Overview of acquired mutations/genes possibly contributing to tigecycline resistance in enterococci.

Species	Target (gene)	Function of target protein	Comment	Reference
<i>E. faecalis</i> / <i>E. faecium</i>	RpsJ	Ribosomal protein S10 (positions 54-61, <i>E. coli</i> numbering)	Some mutations are also present in TGC-susceptible isolates	(Beabout et al., 2015a, b; Cattoir et al., 2015; Fiedler et al., 2016; Niebel et al., 2015)
	Tet(M)	Ribosomal protection protein		(Beabout et al., 2015b; Fiedler et al., 2016)
	Tet(L)	MFS-type efflux pump		(Fiedler et al., 2016)

of linezolid resistance was below 1% (L.P., pers. communication). Data from T.E.S.T., including 400 isolates from four Portuguese centers collected from 2004 to 2016, confirmed the very low incidence of *Enterococcus* spp. resistant to LZD. Some of those isolates were also co-resistant to vancomycin or tigecycline. LRE (n = 7) and enterococci with intermediate resistance (n = 10; according to the Clinical & Laboratory Standards Institute criteria) were identified among 14 sampled hospital wastewaters (Santos Arronches, 2014).

Besides the occurrence of LZD-resistant enterococci in hospitals, their occurrence in other sources and habitats is of special concern. Recent reports from the European Food Safety Authority (EFSA), published together with the ECDC, indicated the presence of LZD resistance in enterococcal indicator isolates (i.e., non-selective screening) collected in 2012 and 2013 from food, especially from bovine and broiler samples, but also from pigs and cattle. The samples originated from various European countries including the Netherlands, France, Croatia, Belgium and Spain (<http://ecdc.europa.eu/en/publications/Publications/antimicrobial-resistance-zoonotic-bacteria-humans-animals-food-EU-summary-report-2013.pdf>; last access 30/10/2018).

### Tigecycline

#### Tigecycline: mode of action

Tigecycline (TGC) is the first member of the class of glycylicyclines (Pankey, 2005). Structurally, it is a derivative of minocycline with a bulky *ter*-butyl-glycylamido side chain at the position 9 of the D ring (Thaker et al., 2010). It exerts bacteriostatic activity against a broad range of bacterial pathogens, including Gram-positive and Gram-negative bacteria, anaerobes and atypical organisms (Pankey, 2005; Stein and Babinchak, 2013). It is currently approved for the treatment of complicated SSTIs and complicated intra-abdominal infections (2005), as well as community-acquired bacterial pneumonia (2008) (Stein and Babinchak, 2013).

Like other tetracyclines, TGC inhibits bacterial protein synthesis at the elongation stage by preventing the binding of the ternary complex amino acid-tRNA-EF-Tu-GTP to the small 30S ribosomal subunit (Brodersen et al., 2000; Chopra and Roberts, 2001; Pioletti et al., 2001) due to an overlap in the primary binding site (named Tet-1) with the position of the anticodon stem loop of the A-site tRNA (Wilson, 2014). Both tetracycline (TET) and TGC reversibly bind through an Mg<sup>2+</sup>-mediated interaction to the Tet-1 site, located in a pocket formed between helices h34 and h31 (16S rRNA) but in different orientations (Bauer et al., 2004; Grossman, 2016). As compared to classical tetracyclines, TGC has an increased binding affinity that results from a stacking interaction between the 9-*ter*-butyl-glycylamido moiety and nucleotide C1054 in h34 (Jenner et al., 2013; Olson et al., 2006).

Interestingly, glycylicyclines are not or only poorly affected by major classical TET resistance determinants, i.e. TET-specific MFS efflux pumps [e.g. Tet(A) and Tet(B) in Gram-negative bacteria; Tet(K) and Tet(L) in Gram-positives] and ribosomal protection proteins [e.g. Tet(M) and Tet(O)] (Grossman, 2016; Thaker et al., 2010). This stronger activity is related to the inhibition of bacterial protein synthesis with a potency three- and 20-fold greater than those of minocycline and TET as well as to a binding to 30S and 70S ribosomes with five- and > 100-fold greater affinity than minocycline and TET, respectively (Bergeron et al., 1996; Olson et al., 2006; Rasmussen et al., 1994).

#### Tigecycline: mechanisms of resistance

Clinical resistance to TGC is almost exclusively described in multi-drug-resistant Gram-negative bacteria, for which this antibiotic has been used as a last-resort option, usually in combination therapy (Sun et al., 2013). It commonly results from the overexpression of resistance-nodulation-cell division (RND) efflux pumps, such as AdeABC in *A. baumannii*, AcrAB-TolC in *K. pneumoniae* and *Enterobacter* spp. or SdeXY-HasF in *Serratia marcescens* (Sun et al., 2013). Of note, both *P. aeruginosa* and *Proteus mirabilis* show an intrinsic reduced susceptibility to TGC associated with the expression of MexXY-OprM and AcrAB-TolC RND-type systems, respectively (Dean et al., 2003; Visalli et al., 2003).

The role of active efflux has also been extensively studied in Gram-positive *S. aureus* mutants obtained *in vitro*, whereas no naturally-occurring clinical isolates with decreased TGC susceptibility have been reported to date (McAleese et al., 2005). In these *in vitro* mutants, TGC resistance was due to overexpression of the *mepA* gene (coding for an efflux pump belonging to the MATE family) caused by the inactivation of its transcriptional repressor MepR (Kaatz et al., 2005; Kumaraswami et al., 2009; McAleese et al., 2005).

In enterococci, the mechanism of TGC decreased susceptibility was first deciphered in *E. faecium* (Cattoir et al., 2015); resistance is defined by a MIC > 0.5 mg/L ([www.eucast.org](http://www.eucast.org)). TGC resistance determinants are summarized in Table 2. Mutants with reduced susceptibility to TGC were obtained *in vitro* by serial passages using two *E. faecium* reference strains, Aus0004 and HM1070 (Cattoir et al., 2015). The derivative mutants, AusTig and HMTig, exhibited MICs 8-fold higher than those of the parental strains (0.25 mg/L vs. 0.03 mg/L, respectively), with a concomitant 4-to-8-fold increase in MICs of TGC, minocycline and doxycycline. Importantly, decreased susceptibility was stable over two weeks without selective pressure, and no decrease in the MIC of TGC was evident in the presence of efflux pump inhibitors such as reserpine or pantoprazole. In addition, two *vanA*-positive *E. faecium* clinical isolates (named EF16 and EF22), collected in 2010 and 2013 in France, were found to have reduced susceptibility to TGC (Cattoir et al., 2015). EF16 was positive for *tet(M)* with a MIC of 0.5 mg/L while EF22 harbored both *tet(M)* and *tet(L)* genes with a MIC of 0.25 mg/L. Following a comparative genomic analysis between AusTig and Aus0004, four different non-synonymous nucleotide mutations were found, including one in the *rpsJ* gene, which encodes for the S10 ribosomal protein of the 30S ribosomal subunit. This mutation led to the D60Y substitution (*E. coli* numbering), which was also detected in HMTig mutants. A close mutation in the S10 ribosomal protein (K57E) was also detected in EF16, whereas no *rpsJ* mutation was found in EF22.

The role of RpsJ in TGC-reduced susceptibility in *E. faecium* was subsequently confirmed (Niebel et al., 2015). In this study, three different pairs of isogenic clinical isolates were characterized, each pair comprising a susceptible strain before TGC therapy and a non-susceptible strain following TGC exposure. The three pre-treatment strains (1S, 2S, and 3S) presented MICs of 0.12 mg/L, whereas among the post-exposure isolates, two (1R and 2R) were resistant (MICs of 8 mg/L) and one isolate (3I) showed reduced susceptibility (MIC of 0.5 mg/L). Comparative genomic analysis showed that all modifications occurred in a small portion of the S10 ribosomal protein comprised between positions 52 and 60, including A54E and H56R substitutions in 3I, a Δ52-56IRATH deletion in 2R and a Δ52I deletion plus Y58H in 1R.

The role of the ribosomal S10 protein as a general target of TGC

insusceptibility was confirmed across several bacterial species. Before *E. faecium*, it was first described in a KPC-producing *K. pneumoniae* clinical isolate, with the description of a substitution V57 L in RpsJ (Villa et al., 2014). Using an experimental evolution approach, several strains of *A. baumannii*, *E. faecium*, *E. coli* and *S. aureus* were adapted to growth in elevated TGC concentrations (Beabout et al., 2015a). At the end of the adaptation period, 35 out of 47 replicate populations presented clones with a mutation in the *rpsJ* gene. The residues where mutations in *rpsJ* were identified in Gram-positives were at nucleotide positions 53–56, 58 and 60 while a mutation observed in both Gram-positive and Gram-negative species was found only at position 57. All identified mutations were found on the vertex of an extended loop of the S10 ribosomal protein (positions 53–60), which is in close proximity to the 16S rRNA TGC binding pocket (Beabout et al., 2015a). The mechanism of reduced susceptibility could result from changes in the conformation or conformational dynamics of 16S rRNA, leading to reduced TGC binding affinity and/or a reduction in translational inhibition by TGC favoring tRNA entry and binding to the ribosome. Of note, the role of *rpsJ* mutations was also described in the early steps of TGC resistance in mutants of *Streptococcus pneumoniae* obtained *in vitro* (Lupien et al., 2015).

A recent study explored the acquisition of TGC resistance in *E. faecalis* using an experimental evolution strategy in a bioreactor setup (Beabout et al., 2015b). In two biological replicates, 24 mutations were found, but only two candidate loci seemed tightly linked to resistance: the 5'UTR of the *tet(M)* gene carried by Tn916 ( $\Delta$ 87-bp and  $\Delta$ 125-bp 51-bp and 36-bp upstream of *tet(M)*, respectively) and the *rpsJ* gene (R53Q- $\Delta$ 54-57ATHK). In this study, the authors also showed that deletions of regulatory elements in the 5'UTR of *tet(M)* led to constitutive overexpression of *tet(M)* and a hyper-conjugative phenotype of Tn916. This study highlights the possibility of rapid spread of a resistance allele throughout the population.

An increase in TGC-resistant enterococci was reported in Germany from 2007 to 2015 ( $n = 73$  *E. faecalis* and *E. faecium*) (Fiedler et al., 2016). In this study, the authors analyzed three isogenic *E. faecium* strains, including the resistant parental strain UW8175 (MIC = 8 mg/L), a susceptible revertant UW8175- (MIC = 0.03 mg/L) and a high-level resistant mutant UW8175+ (MIC = 64 mg/L). By comparative genomic analysis, they found some differences with the parental strain: six SNPs in both UW8175+ and UW8175- mutants, a loss of a 38-kb plasmid harboring *tet(L)* and *tet(M)* in UW8175- and a F62 V substitution in Tet(L) in UW8175+. The authors also demonstrated increased expression of *tet(M)* and *tet(L)* (that were co-transcribed) and an increased plasmid copy number of the very same plasmid in UW8175+ as compared to the parental strain. The putative role of plasmid-associated *tet* genes was confirmed with other clinical isolates. Finally, it was also demonstrated that Tet(M) and Tet(L) were jointly responsible for TGC resistance in another bacterial host (namely *L. monocytogenes* EGD-e).

An uncommon mechanism of TGC resistance is provided by enzymatic inactivation mediated by the flavin-dependent monooxygenase Tet(X) (Forsberg et al., 2015). First described in *Bacteroides fragilis*, then in environmental and clinical isolates of Enterobacteriaceae and Pseudomonadaceae (Grossman, 2016; Moore et al., 2005), it has not been yet described in Gram-positive bacteria. However, *tet(X)* has been detected in the human gut microbiota and demonstrated to be harbored by mobile genetic elements (Aminov, 2013). The overexpression of *tet(X)* in *E. coli* is responsible for a 32-fold increase in the MIC of TGC, and it has also been predicted that its activity against TGC can be mutationally increased (Grossman, 2016; Linkevicius et al., 2016).

#### Tigecycline: resistance surveillance data

Recent surveillance and application studies have shown high levels of TGC susceptibility among Gram-positive clinical isolates worldwide, with no increase in resistance over time (Hoban et al., 2015; Sader et al., 2014b). Out of 20,782 enterococcal isolates (14,615 *E. faecalis* and 6167 *E. faecium*) collected globally between 2004 and 2013, 99.7%

were categorized as susceptible according to the CLSI susceptibility breakpoint ( $\leq 0.25$  mg/L) with a MIC<sub>50</sub> of 0.25 mg/L (MIC range from  $\leq 0.008$  to 1 mg/L) (Hoban et al., 2015). Similar rates of < 1% were reported based on the most recent T.E.S.T. data from Latin America (Vega and Dowzicky, 2017). TGC susceptibility is not included in EARS-Net reporting, so available data on resistance to this agent derive from other sources, studies and surveillance schemes.

In enterococci, only a few TGC non-susceptible clinical isolates have been initially reported with no characterization regarding the mechanism of resistance. This includes one clinical isolate of *E. faecalis* (MIC = 2 mg/L) recovered in Germany in 2007 (Werner et al., 2008b), 10 clinical and non-clinical isolates (MICs = 0.5–1 mg/L) collected in Portugal between 1999 and 2007, of which seven were *E. faecalis*, one *E. gallinarum*, one *E. hirae* and one *Enterococcus* spp. (Freitas et al., 2011), and one clinical isolate of *E. faecalis* (MIC = 2 mg/L) reported in the UK in 2011 (Cordina et al., 2012).

In Spain, TGC resistance is still sporadic for *E. faecalis* (0.2%; range = 0.008–2; MIC<sub>50</sub> = 0.12 mg/L and MIC<sub>90</sub>  $\leq 0.25$  mg/L) and not reported for *E. faecium* (range = 0.015–0.5; MIC<sub>50</sub> = 0.06 mg/L and MIC<sub>90</sub>  $\leq 0.12$  mg/L), according to data from the T.E.S.T. survey that analyzed 1205 *E. faecalis* and 535 *E. faecium* isolates collected from 27 medical centers in Spain between 2004 and 2014 (Marco and Dowzicky, 2016).

In Portugal, data from four major hospitals in the north, center and south of the country showed the occurrence of two TGC non-susceptible *E. faecalis* of ST34 and ST319 from 2002, two from 2015 and two from 2016 [(Freitas et al., 2011); L.P., pers. communication]. In each hospital, the incidence of isolates non-susceptible to TGC was below 1% (L. P., pers. communication). Data available from T.E.S.T. for TGC and comparator compounds in Portugal, including 400 isolates from four centers assessed from 2004 to 2016, showed a single TGC-resistant *E. faecalis* isolate (MIC = 1 mg/L). Five *E. faecalis*, one *E. gallinarum*, one *E. hirae* and eight *Enterococcus* spp. were non-susceptible to TGC (MIC = 0.5–1.0 mg/L), originating from human colonization, chicken carcasses, piggyeries and hospital wastewaters, respectively (Freitas et al., 2011); [https://fenix.tecnico.ulisboa.pt/downloadFile/1970719973965985/Dissertacao\\_Ana%20Arronches.pdf](https://fenix.tecnico.ulisboa.pt/downloadFile/1970719973965985/Dissertacao_Ana%20Arronches.pdf).

In Germany, only limited data exist for TGC susceptibility among clinical enterococcal isolates. As part of the T.E.S.T. surveillance study (2004–2009) performed in France, Germany, Italy, Spain and the UK, overall very low resistance rates were reported for *E. faecium* (2/893; 0.2%) (Aznar et al., 2012). Earlier data for Germany (2004–2007) also revealed very low TGC resistance rates of < 1% (Seifert and Dowzicky, 2009). A German TGC resistance trial called G-TEST (parts I and II) flanking the introduction of this antibiotic into clinical practice in Germany revealed zero (part I) (Kresken et al., 2009) or very low TGC resistance rates (part II and subsequent years) (Kresken et al., 2011, 2009). However, the German NRC recognized an increasing number of confirmed TGC-resistant enterococci, mainly *E. faecium*, in recent years (unpublished data).

Out of 3974 enterococcal clinical isolates received by the French NRC from 2006 to 2016, none were categorized as resistant (MIC > 0.5 mg/L), but two isolates (0.05%) exhibited decreased susceptibility to TGC (MICs = 0.25–0.5 mg/L) (Cattoir et al., 2015) (V. C., unpublished data).

In Ireland, 560 *E. faecium* isolates from bloodstream infections (source: EARS-Net Ireland) were tested against TGC between 2009 and 2016; of these, four were resistant (0.7%). In one institution, TGC resistance was detected in 11 VRE isolates between 2012 and 2017, in one VR-*E. faecalis* and ten VR-*E. faecium* strains. The isolates were cultured from eight clinical specimens and three screening samples. One TGC-resistant VR-*E. faecium* isolate was resistant to LZD as well. Of note, TGC susceptibility was analyzed using automated testing (VITEK II) only and resistance was not confirmed by a second independent method.

TGC resistance was low among enterococcal isolates from



**Table 3**  
Overview of acquired mutations possibly contributing to daptomycin resistance in enterococci.

Species	Target (gene)	Function of target	Comment	References
<i>E. faecalis</i>	LiaFSR ( <i>liaFSR</i> )	Regulators involved in cell envelope stress response	Initial tolerance MIC 3–4 mg/L	(Arias et al., 2011; Miller et al., 2013)
	Cis ( <i>cis</i> )	Cardiolipin synthetase involved in phospholipid metabolism	Results in non-susceptibility in combination with <i>liaFSR</i> mutations	(Miller et al., 2013; Palmer et al., 2011)
	GdpD ( <i>gdpD</i> )	Glycerophosphoryl diester phosphodiesterase involved in phospholipid metabolism	Mutations in <i>gdpD</i> alone do not modify daptomycin susceptibility	(Arias et al., 2011)
	YvlB ( <i>yvlB</i> )	Putative LiaFSR pathway target protein	* Mutation in <i>yvlB</i> alone results in non-susceptibility	(Miller et al., 2013; Palmer et al., 2011)
	GdpP ( <i>yylP</i> )	Cyclic dinucleotide phosphodiesterase associated with the membrane stress response		(Miller et al., 2013; Palmer et al., 2011)
	GshF ( <i>gshF</i> )	Glutathione synthase involved in the oxidative stress response		(Miller et al., 2013)
	MdpA ( <i>mdpA</i> )	Putative multidrug resistance pump		(Miller et al., 2013)
	EF2470	Protein with unknown function		(Palmer et al., 2011)
		Contains an HD domain, which may be involved in nucleic acid metabolism and signal transduction		
		Membrane protein with a serine incorporator domain		
<i>E. faecium</i>	EF1797 ( <i>drrmA</i> )	May aid in serine and phospholipid biosynthesis and promote transport of polar serine into the hydrophobic cell membrane		(Miller et al., 2013; Palmer et al., 2011)
	EF1753	Protein with auto-transporter adhesin domain that is likely involved in cell adhesion		(Palmer et al., 2011)
	EF0716	Membrane protein with FamA-like transporter family domain		(Palmer et al., 2011)
	LiaFSR ( <i>liaFSR</i> )	Regulators involved in the cell envelope stress response	Most common initial mutations in daptomycin non-susceptible isolates	(Diaz et al., 2014; Panesso et al., 2015) (Sinel et al., 2016a)
	YycFG ( <i>yycFG</i> )	Regulators of cell wall synthesis and homeostasis	Detected in both daptomycin resistant and tolerant isolates	(Diaz et al., 2014; Tran et al., 2013a)
	Cis ( <i>cis</i> )	Cardiolipin synthetase involved in phospholipid metabolism	Mutations in additional target needed to mediate non-susceptibility	(Diaz et al., 2014; Sinel et al., 2016a; Tran et al., 2013a)
	GdpD ( <i>gdpD</i> )	Glycerophosphoryl diester phosphodiesterase involved in phospholipid metabolism		(Diaz et al., 2014)
	Cfa ( <i>cfa</i> )	Cyclopropane synthase involved in phospholipid metabolism		(Diaz et al., 2014; Tran et al., 2013a; Werth et al., 2014)
		HD domain containing protein similar to that in <i>E. faecalis</i>		(Diaz et al., 2014; Lellek et al., 2015; Tran et al., 2013a)
		23S rRNA methyltransferase		(Diaz et al., 2014; Tran et al., 2013a)
<i>E. faecalis</i>	TelA	Tellurite resistance protein		(Diaz et al., 2014)
		Alpha-mannosidase		(Humphries et al., 2012)
	<i>aad</i>	Alcohol dehydrogenase		(Humphries et al., 2012)
	EzrA	Transmembrane negative regulator of the septation ring formation protein FtsZ		(Humphries et al., 2012)
		Protein with unknown function		(Humphries et al., 2012)
	PTS IID	Mannose-specific IID component of PTS		(Humphries et al., 2012)
	<i>relA</i>	Involved in the stringent response		(Honsa et al., 2017)
			* Tolerance to daptomycin	

\* Potentially contributing to non-susceptibility.

bloodstream infections reported by the national Italian surveillance system (AR-ISS). Among the *E. faecalis* isolates tested in 2015 and 2016, 0.8% (n = 9/1,105) and 0.16% (n = 2/1,244) were TGC-resistant, respectively. Corresponding data for *E. faecium* revealed TGC resistance in 0.4% (n = 2/490; 2015) and 0.3% (n = 2/611; 2016).

In Poland, Norway and Denmark TGC-resistant enterococcal isolates have not been reported to date.

### Daptomycin

#### Daptomycin: mode of action

Daptomycin (DAP) is a cyclic anionic lipopeptide synthesized from the lipopeptide antibiotic A21978C complex produced by *Streptomyces roseosporus* (Debono et al., 1988; Eliopoulos et al., 1985). In 2003, the USA FDA approved the use of DAP for the treatment of complicated SSTIs at a dosage of 4 mg/kg/day, and later for the treatment of *S. aureus* bacteremia with or without right-sided infective endocarditis at a dosage of 6 mg/kg/day (<https://www.fda.gov/Drugs/>). The EMA approved DAP for the same indications in January 2006. Treatment dosage ranges from 4 to 10 mg/kg/day. Different studies have documented the safety and tolerability of high-dose daptomycin (8–12 mg/kg/day) (Casapao et al., 2013).

DAP shares structural and functional similarities with cationic antimicrobial peptides produced by the mammalian innate immune system (Jung et al., 2004). The bactericidal effect of DAP involves alterations to bacterial cell envelope homeostasis through interactions with membrane phospholipids; however, this process is not yet fully understood (Tran et al., 2015). Insertion of DAP into the bacterial cytoplasmic membrane is dependent on the divalent cation form of calcium ( $\text{Ca}^{2+}$ ) (Jung et al., 2004; Taylor et al., 2016). In addition, calcium is required for the formation of DAP micellar structures proposed to facilitate delivery of the antibiotic to the bacterial cytoplasmic membrane (Ho et al., 2008; Scott et al., 2007; Taylor et al., 2016). Important steps in the DAP interaction with Gram-positive cell membranes after insertion include oligomerization of DAP-calcium complexes in the outer leaflet (Muraih et al., 2012, 2011) and subsequent translocation of DAP oligomers into the inner leaflet of the cell membrane to form functional ring-like pores of two opposing oligomers. Oligomerization is dependent on interactions with the phospholipid phosphatidylglycerol (Muraih et al., 2012, 2011), and translocation of the oligomers is negatively influenced by the presence of the negatively charged phospholipid cardiolipin (Zhang et al., 2014b). DAP oligomerization is required for its antibacterial activity (Zhang et al., 2013). DAP pores are cation- and size-selective with greater permeability to  $\text{Na}^+$ ,  $\text{K}^+$  and other metal cations (Zhang et al., 2014a). The presence of pores in the membrane presumably results in ion leakage, membrane depolarization and cell death without lysis (Silverman et al., 2003; Zhang et al., 2014a).

In addition, DAP induces abnormal septation events (Cotroneo et al., 2008; Hachmann et al., 2009) and cell wall stress responses (Muthaiyan et al., 2008), suggesting that this compound causes significant cell wall damage. A second explanation for cell death due to the insertion of DAP into the cell membrane could be that DAP oligomers induce curved membrane patches (Jung et al., 2008; Pogliano et al., 2012). These curved patches are then incorrectly identified by an essential cell division protein as a potential site of division, inducing local activation of peptidoglycan biosynthesis which causes striking cell wall and membrane deformities and finally a rupture in the cell membrane and cell death (Pogliano et al., 2012).

Chen et al. proposed the extraction of lipids from the cell membrane as yet another alternative mode of action for DAP (Chen et al., 2014). They discovered that the interaction between DAP and giant unilamellar vesicles resulted in a lipid extraction effect, i.e. removing lipids from vesicles. This effect was only observed in the presence of phosphatidylglycerol and calcium. However, the authors did not present any evidence that lipid extraction is required for the antibacterial action of

DAP.

#### Daptomycin: mechanisms of resistance

Neither the EUCAST nor the CLSI have established (clinical) breakpoints for DAP resistance in enterococci, but they have set the epidemiological cut-off (ECOFF) value for *E. faecium* and *E. faecalis* to  $\leq 4 \mu\text{g/ml}$  for DAP ([http://www.eucast.org/fileadmin/src/media/PDFs/EUCAST\\_files/General\\_documents/EUCAST\\_daptomycin\\_guidance\\_note\\_20160924.pdf](http://www.eucast.org/fileadmin/src/media/PDFs/EUCAST_files/General_documents/EUCAST_daptomycin_guidance_note_20160924.pdf)) (Humphries et al., 2013). Unfortunately, this value is considered as a clinical breakpoint by some experts (Shukla et al., 2016). For the purposes of this review, enterococci with DAP MIC  $\geq 8 \mu\text{g/mL}$  will be referred to as “DAP-resistant”.

DAP resistance in enterococci has been associated with distinct structural alterations to the cell envelope, including increased cell wall thickening and decreased ability of DAP to alter cell membrane integrity. Moreover, the development of DAP resistance is associated with a compelling reduction in cell membrane fluidity and phosphatidylglycerol content and an accompanying increase in glycerolphosphodiglycodyciacylglycerol (Arias et al., 2011; Mishra et al., 2012; Steed et al., 2011). Acquired DAP resistance is associated with mutations in intrinsic genes (summarized in Table 3). Much of the knowledge regarding which mutations are involved in DAP resistance comes from studies comparing the genomes, membrane potential and cell envelope composition of susceptible -and resistant more-or less isogenic mutants developed either in the laboratory or in patients during DAP therapy. In enterococci, DAP resistance can be linked to mutations in two major groups of genes, including genes encoding regulatory pathways involved in the coordination of the cell envelope stress response, such as LiaFSR and YycFGHLJ, and genes encoding enzymes involved in the metabolism of phospholipids important for cell membrane homeostasis, such as glycerophosphoryl diester phosphodiesterase (*gdpD*) and cardiolipin synthetase (*cls*) (Arias et al., 2011; Miller et al., 2016; Tran et al., 2015).

In *E. faecalis*, initial DAP tolerance is due to the redistribution of cell membrane cardiolipin-rich microdomains, which diverts DAP from its preferred binding site at the septum. Tolerance with a MIC close to the susceptibility breakpoint is associated with mutations within LiaFSR. High levels of MICs can be reached when additional mutations occur in *gdpD* and *cls* that alter the phospholipid content of the cell membrane (Arias et al., 2011; Mishra et al., 2012; Munita et al., 2013; Reyes et al., 2015; Tran et al., 2015, 2013b). A mutation in *gdpD* does not modify DAP susceptibility on its own (Arias et al., 2011). In agreement with this, adaptation of *E. faecalis* to gradually increasing concentrations of DAP revealed that the critical first step to resistance involves changes in the LiaFSR pathway. Almost half of the endpoint strains also had a mutation in *cls* (Miller et al., 2013).

Mutations in several other genes have been described in DAP-resistant *E. faecalis*, but further experiments are needed to reveal the exact function of these genes in *E. faecalis* and confirm that the mutations are causatively linked to DAP resistance. Potential resistance mutations have been reported in genes encoding: 1) GdpP (encoded by *yybT*), a cyclic dinucleotide phosphodiesterase associated with the membrane stress response (Miller et al., 2013; Wang et al., 2017); 2) YvIB, a putative LiaFSR pathway target protein; 3) GshF, a glutathione synthase; 4) MdpA putative multidrug resistance pump (Miller et al., 2013); 5) a protein with unknown function containing an HD domain found in an enzyme superfamily with phosphohydrolase activity, which may be involved in nucleic acid metabolism and signal transduction; 6) a membrane protein with a serine incorporator domain that may aid in serine and phospholipid biosynthesis and promote transport of the polar serine into the hydrophobic cell membrane; 7) a protein with an auto-transporter adhesin domain that likely is involved in cell-adhesion; and 8) a membrane protein with an EamA-like transporter family domain (Palmer et al., 2011).

DAP resistance development seems to be more common in *E. faecium* than in *E. faecalis* (Miller et al., 2014). Initial DAP resistance in *E.*

*faecium* is probably due to electrostatic repulsion of the positively charged DAP-Ca<sup>2+</sup> complex from the cell membrane, as seen in *S. aureus* and *Bacillus subtilis*. Resistance is not associated with the redistribution of anionic microdomains as in *E. faecalis*, although mutations in *liaFSR* are the most common changes seen in daptomycin-resistant *E. faecium* (Diaz et al., 2014; Tran et al., 2015). Deletion of *liaR* from clinical strains harboring mutations in both the LiaFSR and YycFG pathways reverses DAP resistance and restores the binding of DAP to the *E. faecium* cell surface (Panesso et al., 2015). The second most common modifications in *E. faecium* are mutations in YycFG and the accessory protein YycHIJ, detected in both DAP-resistant and tolerant isolates. Mutations in *cls* are further commonly related with DAP resistance in *E. faecium*, and mutations in *gdpD* can occasionally be found (Diaz et al., 2014; Tran et al., 2013a). *Cls* and YycFG changes only are probably not sufficient to mediate resistance (Diaz et al., 2014; Tran et al., 2015, 2013a). In *E. faecium*, mutations in LiaFSR appear before *cls* mutations, indicating a sequential progression towards resistance. Interestingly, when selective pressure is removed, *E. faecium* reverts to become hyper-susceptible through the insertion of IS elements in *liaFSR* (Sinel et al., 2016a). There are also several reports of mutations in the *cfa* gene encoding a cyclopropane synthase involved in phospholipid metabolism in *E. faecium* (Diaz et al., 2014; Tran et al., 2013a; Werth et al., 2014). The involvement of *cfa* mutations in resistance is supported by a significant increase in cyclopropane fatty acids in DAP-resistant *E. faecium*. These fatty acids aid in maintaining the cell membrane in the presence of different stressors (Mishra et al., 2012).

Comparing the genomes of DAP-susceptible and DAP-resistant *E. faecium* strains has suggested several other mutant genes potentially associated with DAP resistance, but further functional evidence is needed to support these genetic findings. These genes encode: 1) an HD domain-containing protein also linked to DAP resistance in *E. faecalis* (Diaz et al., 2014; Lellek et al., 2015; Tran et al., 2013a); 2) RrmA 23S rRNA methyltransferase (Diaz et al., 2014; Tran et al., 2013a); 3) the tellurite resistance protein TelA also linked to DAP resistance in *E. faecalis* (Diaz et al., 2014); 4) alpha-mannosidase; 5) alcohol dehydrogenase; 6) EzrA, a transmembrane negative regulator of the septation ring formation protein FtsZ; 7) a protein with unknown function; and 8) the mannose-specific IID component of PTS (mannose-specific PTS mutations in the IIC and IID domains have been linked to resistance to class IIa cationic antimicrobial peptide bacteriocins; (Humphries et al., 2012). Furthermore, whole genome sequencing of isogenic strains revealed the loss of a 43 kb chromosomal fragment with 42 genes, presumably leading to DAP resistance as no mutations were observed in genes previously described to confer DAP resistance (Gumpert et al., manuscript submitted). Altered cell surface charges or the modification of PTS as a receptor are likely to result in DAP resistance in *E. faecium* (Humphries et al., 2012). A mutation in *relA* developed during vancomycin-resistant *E. faecium* bacteremia in a leukemia patient caused a constitutively activated stringent response with elevated baseline levels of the alarmone guanosine tetraphosphate and tolerance to high doses of both DAP and LZD in a biofilm. This explained why this infection persisted despite appropriate antibacterial treatment (Honsa et al., 2017). Analyses of a DAP-resistant *E. faecium* that emerged during DAP therapy of endocarditis in a leukemia patient showed among other mutations alterations in the DNA repair proteins MutL and RecJ. These changes caused a significantly enhanced mutation frequency that possibly contributed to development of high-level DAP resistance (Matono et al., 2016).

So far, no mutations were identified in *mprF* (multiple peptide resistance factor) or the *dltABCD* operon that could be directly linked to DAP resistance in enterococci (Arias et al., 2011; Montero et al., 2008).

The examples above demonstrate that changes in many possible target genes can result in DAP tolerance and resistance and that these mutations are easily acquired, especially under selective pressure in *E. faecium*.

#### Daptomycin: resistance surveillance data

Most enterococcal isolates (> 99.8%) are susceptible to DAP worldwide (Sader et al., 2014a), and this trend did not change in European and US hospitals in the period from 2009 to 2013 (Sader et al., 2015). The occurrence of DAP-resistant enterococci is most often associated with prior DAP exposure (DiPippo et al., 2016; Lellek et al., 2015; Lewis et al., 2016) and may correspond to the use of too low dosages of DAP not providing high enough plasma concentrations to kill enterococci (Hall et al., 2012; Sinel et al., 2016b; Werth et al., 2014). A recent study provided evidence of the frequent *in vitro* increase of MIC values to DAP among *E. faecalis* strains highly resistant to gentamicin (Pericas et al., 2017). The results suggest the convenience of testing the MIC values of combination compounds in the presence of subinhibitory concentrations of DAP to predict the efficacy of combination therapy for multidrug resistant enterococci (e.g. DAP plus ampicillin). When DAP is used in combination with other antibiotics, the possibility of selecting MDR phenotypes might be increased (Cercenado and Pachon, 2012). DAP susceptibility data for enterococci cannot be included in EARS-Net data (due to a missing EUCAST breakpoint), so resistance data are derived from other sources, studies and surveillance schemes.

Although *in vitro* resistance arises spontaneously at low frequencies, the emergence of resistance is increasingly selected during DAP therapy. It could be related to the mutant selection window (MSW), which lies between the MIC and the mutant prevention concentration (MPC) and corresponds to the concentration range within which free C<sub>max</sub> (taking into account the protein binding of DAP, approx. 90%) values for DAP usually fell into the MSW when using low dosages (4–6 mg/kg), thus suggesting a risk of *in vivo* selection of resistant mutants, especially at sites of infection where the concentration of DAP is low (Sinel et al., 2016b). Therefore, high daily doses of DAP (≥ 10 mg/kg) may be considered for the treatment of infections due to *E. faecium*.

The transmission of DAP- and vancomycin-resistant enterococci within the hospital environment has been documented (Lellek et al., 2015). Moreover, population genetics assessing DAP-resistant *E. faecium* from a medical center in New York showed that a novel clone, ST736, was associated with resistance in this region (Wang et al., 2014).

The European Cubicin Outcomes Registry and Experience (EU-CORE) had been introduced as a retrospective, European, post-marketing, non-interventional registry of DAP use in patients who received at least one DAP dose. The database provides information from 6075 patients representing 18 countries and 314 study sites. The primary objective was to evaluate the clinical outcomes of patients treated with this drug (Cogo et al., 2015; Gonzalez-Ruiz et al., 2015).

In Spain, DAP is mostly used as a rescue therapy, at 6 mg/kg/day, in both hospitalized and non-hospitalized patients according to data from the EU-CORE database (Spain, with 47 hospitals, is the best represented country in this database) (Cercenado and Pachon, 2012; Sandoval et al., 2015). The first DAP-resistant enterococcal isolate in Spain was an ST117 *E. faecium* strain (MIC = 12 mg/L) recovered from a 46-year-old woman following kidney transplantation in 2005. The isolate showed a novel substitution in the response regulator LiaR and also harbored a change in the active site of the cardiolipin synthase (Sorlozano et al., 2015). The proportions of DAP-resistant *E. faecalis* and *E. faecium* are 3.23% and 10.53%, respectively, according to a national collaborative study performed in Spain (M. Palomar, unpublished data). The mechanisms of resistance were not explored in this study.

In France, a recent increase in the MIC of DAP in *E. faecium* has been observed, particularly in the proportion of strains with MICs > 4 mg/L (6–12 mg/L): 0% for the period 2006–2013, 1.1% (3/267) in 2014, 7.0% (24/344) in 2015 and 8.2% (31/379) in 2016.

In Ireland, among 60 *E. faecium* isolates from bloodstream infections (source: EARS-Net Ireland) tested against DAP between 2009 and 2014, four were resistant (7%). A tertiary care referral center has detected 12

**Table 4**

Proposal for a common nomenclature of VRE demonstrating resistance to linezolid, daptomycin and/or tigecycline. We propose abbreviating Vancomycin (V), Linezolid (L), Daptomycin (D) and Tigecycline (T) as single letters; however, combinations of several resistances should be written as given in the Table with abbreviated antibiotics in an alphabetical order.

Resistance to	Vancomycin VRE	Linezolid LRE	Daptomycin DRE	Tigecycline TRE
<b>Vancomycin</b>		LVRE	DVRE	TVRE
<b>Linezolid</b>			DLRE	LTRE
<b>Daptomycin</b>				DTRE

DAP-resistant *E. faecium* isolates since 2016; six *E. faecium* isolates were sensitive to vancomycin and resistant to DAP, the other six isolates were VR-*E. faecium* resistant to DAP. All isolates were susceptible to LZD and TGC. One patient who had received a lengthy course of DAP while in the ICU was found to carry VR-*E. faecium* resistant to DAP both in the intra-abdominal drain fluid and blood culture. All DAP-resistant isolates were grown from clinical specimens (six blood cultures, one wound swab and five drain fluids).

Data on DAP susceptibility in clinical enterococci in Italy is limited, as only one laboratory tests and reports data from enterococcal bacteremia to the national Italian surveillance system (AR-ISS). In 2015 and 2016, a total of 181 isolates were tested, of which 125 were *E. faecalis*, with two that were DAP-resistant. Of 56 *E. faecium* isolates, none were DAP-resistant.

In Denmark, DAP-resistant enterococci are not sent to the antimicrobial reference laboratory on a routine basis. However, during 2017, resistance to DAP was detected in two *E. faecalis* strains and one *E. faecium* isolate at Rigshospitalet, Denmark (H.W. and J. D. Knudsen, pers. communication).

In Germany, DAP-resistant enterococci are sent to the reference centre on a voluntary basis. During recent years, resistance to DAP was confirmed for a few *E. faecalis* and *E. faecium* isolates per year only, performed and analysed by two independent methods (broth micro-dilution and Etest<sup>®</sup>; J.B. and G.W. unpublished data).

No DAP-resistant enterococcal isolates have been noted in Poland, Norway and Portugal so far.

Individual resistance development under therapy and subsequent transmission of successful DAP-resistant clones warrant careful surveillance of these isolates in order to prevent their wider spread and to keep DAP as a valuable therapeutic alternative.

## Conclusions and proposal for a common nomenclature

The review summarizes the current knowledge on mechanisms of resistance to the last-resort antibiotics linezolid, tigecycline and daptomycin in *Enterococcus* spp., and where available provides relevant surveillance data, partly unpublished, for many European countries.

We advocate using a suitable method for confirmation of these resistances, such as the broth dilution assay, and additionally recommend submitting relevant strains to reference laboratories for external confirmation, to support their reference and alert functions and to elucidate the corresponding resistance mechanisms. Using a common and agreed-upon standard is essential and available for Europe using the EUCAST methodology, clinical breakpoints and guidelines ([www.eucast.org](http://www.eucast.org)).

What has been causally indicative for VRE prediction on the basis of the presence of *vanA* or *vanB* genes is far from easy for the resistance phenotypes discussed in this review, which in turn require more sophisticated confirmatory assays based on whole genome sequence (WGS) data, as well as sophisticated transcriptomic and genetic analyses. Loss of susceptibility to linezolid, tigecycline and daptomycin in enterococci may be associated with a variety of resistance mechanisms (target mutations, gene acquisitions, “gene dosage effects” and many others); a number of them also appear in a combined fashion. Resolving

complex resistance mechanisms requires high throughput data entering and hypothesis-free approaches, thus allowing an “open view” (Chacko et al., 2018). Considering this potential complexity, extracting a clinical breakpoint (an MIC) for linezolid, tigecycline and daptomycin for *Enterococcus* isolates or predicting potential therapeutic success from WGS data seems challenging and, at present, requires additional and comprehensive isolate collections and more dedicated studies to support this concept. Genome-wide association studies will potentially pave the way towards the better prediction of resistance phenotypes out of multiple genome comparisons (Power et al., 2017).

Resistance to last-resort antibiotics in enterococci which, in general, serve as important indicator bacteria, requires contemporaneous and strict surveillance to detect the emergence and dissemination of these important resistance mechanisms. With this review, we also propose a common nomenclature for VRE additionally exhibiting these resistance phenotypes (Table 4).

## References

- Adams, D.J., Eberly, M.D., Goudie, A., Nylund, C.M., 2016. Rising vancomycin-resistant *Enterococcus* infections in hospitalized children in the United States. *Hosp. Pediatr.* 6, 404–411.
- Akpaka, P.E., Kisson, S., Jayaratne, P., Wilson, C., Golding, G.R., Nicholson, A.M., Lewis, D.B., Hermelijn, S.M., Wilson-Pearson, A., Smith, A., 2017. Genetic characteristics and molecular epidemiology of vancomycin-resistant *Enterococci* isolates from Caribbean countries. *PLoS One* 12, e0185920.
- Alonso, M., Marin, M., Iglesias, C., Cercenado, E., Bouza, E., Garcia de Viedma, D., 2014. Rapid identification of linezolid resistance in *Enterococcus* spp. based on high-resolution melting analysis. *J. Microbiol. Methods* 98, 41–43.
- Aminov, R.I., 2013. Evolution in action: dissemination of tet(X) into pathogenic microbiota. *Front. Microbiol.* 4, 192.
- Antonelli, A., D'Andrea, M.M., Brenciani, A., Galeotti, C.L., Morroni, G., Pollini, S., Valardo, P.E., Rossolini, G.M., 2018. Characterization of *poxtA*, a novel phenicol-oxazolidinone-tetracycline resistance gene from an MRSA of clinical origin. *J. Antimicrob. Chemother.*
- Arias, C.A., Murray, B.E., 2012. The rise of the *Enterococcus*: beyond vancomycin resistance. *Nat. Rev. Microbiol.* 10, 266–278.
- Arias, C.A., Panesso, D., McGrath, D.M., Qin, X., Mojica, M.F., Miller, C., Diaz, L., Tran, T.T., Rincon, S., Barbu, E.M., Reyes, J., Roh, J.H., Lobos, E., Sodergren, E., Pasqualini, R., Arap, W., Quinn, J.P., Shamoo, Y., Murray, B.E., Weinstock, G.M., 2011. Genetic basis for in vivo daptomycin resistance in enterococci. *N. Engl. J. Med.* 365, 892–900.
- Aznar, J., Lepe, J.A., Dowzicky, M.J., 2012. Antimicrobial susceptibility among *E. Faecalis* and *E. Faecium* from France, Germany, Italy, Spain and the UK (T.E.S.T. Surveillance study, 2004-2009). *J. Chemother.* 24, 74–80.
- Barber, K.E., King, S.T., Stover, K.R., Pogue, J.M., 2015. Therapeutic options for vancomycin-resistant enterococcal bacteremia. *Expert Rev. Anti. Ther.* 13, 363–377.
- Bassetti, M., Farrel, P.A., Callan, D.A., Topal, J.E., Demby, L.M., 2003. Emergence of linezolid-resistant *Enterococcus faecium* during treatment of enterococcal infections. *Int. J. Antimicrob. Agents* 21, 593–594.
- Bauer, G., Berens, C., Projan, S.J., Hillen, W., 2004. Comparison of tetracycline and tigecycline binding to ribosomes mapped by dimethylsulphate and drug-directed Fe<sup>2+</sup> cleavage of 16S rRNA. *J. Antimicrob. Chemother.* 53, 592–599.
- Beabout, K., Hammerstrom, T.G., Perez, A.M., Magalhaes, B.F., Prater, A.G., Clements, T.P., Arias, C.A., Saxer, G., Shamoo, Y., 2015a. The ribosomal S10 protein is a general target for decreased tigecycline susceptibility. *Antimicrob. Agents Chemother.* 59, 5561–5566.
- Beabout, K., Hammerstrom, T.G., Wang, T.T., Bhatti, M., Christie, P.J., Saxer, G., Shamoo, Y., 2015b. Rampant parasexuality evolves in a hospital pathogen during antibiotic selection. *Mol. Biol. Evol.* 32, 2585–2597.
- Bender, J.K., Fleige, C., Klare, I., Fiedler, S., Mischnik, A., Mutters, N.T., Dingle, K.E., Werner, G., 2016. Detection of a cfr(B) variant in German *Enterococcus faecium* clinical isolates and the impact on linezolid resistance in *Enterococcus* spp. *PLoS One* 11, e0167042.
- Bender, J.K., Fleige, C., Lange, D., Klare, I., Werner, G., 2018. Rapid emergence of highly variable and transferable oxazolidinone and phenicol resistance gene *optrA* in German *Enterococcus* spp. clinical isolates. *Int. J. Antimicrob. Agents.*
- Bergeron, J., Ammirati, M., Danley, D., James, L., Norcia, M., Retsema, J., Strick, C.A., Su, W.G., Sutcliffe, J., Wondrack, L., 1996. Glycylcyclines bind to the high-affinity tetracycline ribosomal binding site and evade Tet(M)- and Tet(O)-mediated ribosomal protection. *Antimicrob. Agents Chemother.* 40, 2226–2228.
- Bi, R., Qin, T., Fan, W., Ma, P., Gu, B., 2017. The emerging problem of linezolid-resistant enterococci. *J. Glob. Antimicrob. Resist.* 13, 11–19.
- Birmingham, M.C., Rayner, C.R., Meagher, A.K., Flavin, S.M., Batts, D.H., Schentag, J.J., 2003. Linezolid for the treatment of multidrug-resistant, gram-positive infections: experience from a compassionate-use program. *Clin. Infect. Dis.* 36, 159–168.
- Boucher, H.W., Talbot, G.H., Bradley, J.S., Edwards, J.E., Gilbert, D., Rice, L.B., Scheld, M., Spellberg, B., Bartlett, J., 2009. Bad bugs, no drugs: no ESKAPE! An update from the Infectious Diseases Society of America. *Clin. Infect. Dis.* 48, 1–12.
- Brenciani, A., Morroni, G., Vincenzi, C., Manso, E., Mingoa, M., Giovanetti, E., Valardo, P.E., 2016. Detection in Italy of two clinical *Enterococcus faecium* isolates carrying

- both the oxazolidinone and phenicol resistance gene *optrA* and a silent multi-resistance gene *cf*. J. Antimicrob. Chemother. 71, 1118–1119.
- Brickner, S.J., Barbachyn, M.R., Hutchinson, D.K., Manninen, P.R., 2008. Linezolid (ZYVOX), the first member of a completely new class of antibacterial agents for treatment of serious gram-positive infections. J. Med. Chem. 51, 1981–1990.
- Brodersen, D.E., Clemons Jr, W.M., Carter, A.P., Morgan-Warren, R.J., Wimberly, B.T., Ramakrishnan, V., 2000. The structural basis for the action of the antibiotics tetracycline, pactamycin, and hygromycin B on the 30S ribosomal subunit. Cell 103, 1143–1154.
- Cafini, F., Nguyen le, T.T., Higashide, M., Roman, F., Prieto, J., Morikawa, K., 2016. Horizontal gene transmission of the *cf* gene to MRSA and Enterococcus: role of Staphylococcus epidermidis as a reservoir and alternative pathway for the spread of linezolid resistance. J. Antimicrob. Chemother. 71, 587–592.
- Cai, J., Wang, Y., Schwarz, S., Zhang, G., Chen, S., Gu, D., Shen, Y., Li, D., Fan, R., Zhang, R., 2016. High detection rate of the oxazolidinone resistance gene *optrA* in Enterococcus faecalis isolated from a Chinese anorectal surgery ward. Int. J. Antimicrob. Agents 48, 757–759.
- Casapao, A.M., Kullar, R., Davis, S.L., Levine, D.P., Zhao, J.J., Potoski, B.A., Goff, D.A., Crank, C.W., Segreti, J., Sakoulas, G., Cosgrove, S.E., Rybak, M.J., 2013. Multicenter study of high-dose daptomycin for treatment of enterococcal infections. Antimicrob. Agents Chemother. 57, 4190–4196.
- Cattoir, V., Isnard, C., Cosquer, T., Odhiambo, A., Bucquet, F., Guerin, F., Giard, J.C., 2015. Genomic analysis of reduced susceptibility to tetracycline in Enterococcus faecium. Antimicrob. Agents Chemother. 59, 239–244.
- Cavaco, L.M., Korsgaard, H., Kaas, R.S., Seyfarth, A.M., Leekitcharoenphon, P., Hendriksen, R.S., 2017. First detection of linezolid resistance due to the *optrA* gene in enterococci isolated from food products in Denmark. J. Glob. Antimicrob. Resist. 9, 128–129.
- Cercenado, E., Pachon, J., 2012. [The EUCORE registry: objectives and general results]. Enferm. Infecc. Microbiol. Clin. 30 (Suppl. 1), 3–9.
- Cercenado, E., Marin, M., Bouza, E., 2010. Emerging linezolid resistance: dissemination of the *cf* gene among Staphylococcus aureus, Staphylococcus epidermidis, enterococcus faecium and Enterococcus faecalis and inability of the Etest method for detection. 50th Interscience Conference on Antimicrobial Agents and Chemotherapy.
- Chacko, K., Sullivan, M., Beckford, C., Altman, D., Ciferri, B., Pak, T., Sebra, R., Kasarskis, A., Hamula, C., van Bakel, H., 2018. The genetic basis of emerging vancomycin, linezolid, and daptomycin heteroresistance in a case of persistent Enterococcus faecium bacteremia. Antimicrob. Agents Chemother.
- Chen, H., Wu, W., Ni, M., Liu, Y., Zhang, J., Xia, F., He, W., Wang, Q., Wang, Z., Cao, B., Wang, H., 2013. Linezolid-resistant clinical isolates of enterococci and Staphylococcus cohnii from a multicenter study in China: molecular epidemiology and resistance mechanisms. Int. J. Antimicrob. Agents 42, 317–321.
- Chen, Y.F., Sun, T.L., Sun, Y., Huang, H.W., 2014. Interaction of daptomycin with lipid bilayers: a lipid extracting effect. Biochemistry 53, 5384–5392.
- Chopra, I., Roberts, M., 2001. Tetracycline antibiotics: mode of action, applications, molecular biology, and epidemiology of bacterial resistance. Microbiol. Mol. Biol. Rev. 65, 232–260 second page, table of contents.
- Coburn, B., Low, D.E., Patel, S.N., Poutanen, S.M., Shahinas, D., Eshaghi, A., Willey, B.M., McGeer, A., 2014. Vancomycin-variable Enterococcus faecium: in vivo emergence of vancomycin resistance in a vancomycin-susceptible isolate. J. Clin. Microbiol. 52, 1766–1767.
- Cogo, A., Gonzalez-Ruiz, A., Pathan, R., Hamed, K., 2015. Real-world treatment of complicated skin and soft tissue infections with daptomycin: results from a large European registry (EU-CORE). Infect. Dis. Ther. 4, 273–282.
- Cordina, C., Hill, R., Deshpande, A., Hood, J., Inkster, T., 2012. Tigecycline-resistant Enterococcus faecalis associated with omepazole use in a surgical patient. J. Antimicrob. Chemother. 67, 1806–1807.
- Cotroneo, N., Harris, R., Perlmutter, N., Beveridge, T., Silverman, J.A., 2008. Daptomycin exerts bactericidal activity without lysis of Staphylococcus aureus. Antimicrob. Agents Chemother. 52, 2223–2225.
- Courvalin, P., 2006. Vancomycin resistance in gram-positive cocci. Clin. Infect. Dis. 42 (Suppl 1) pp. S25–34.
- Cui, L., Wang, Y., Lv, Y., Wang, S., Song, Y., Li, Y., Liu, J., Xue, F., Yang, W., Zhang, J., 2016. Nationwide surveillance of novel oxazolidinone resistance gene *optrA* in Enterococcus isolates in China from 2004 to 2014. Antimicrob. Agents Chemother. 60, 7490–7493.
- Dean, C.R., Visalli, M.A., Projan, S.J., Sum, P.E., Bradford, P.A., 2003. Efflux-mediated resistance to tigecycline (GAR-936) in Pseudomonas aeruginosa PAO1. Antimicrob. Agents Chemother. 47, 972–978.
- Debono, M., Abbott, B.J., Molloy, R.M., Fukuda, D.S., Hunt, A.H., Daupert, V.M., Counter, F.T., Ott, J.L., Carrell, C.B., Howard, L.C., et al., 1988. Enzymatic and chemical modifications of lipopeptide antibiotic A21978C: the synthesis and evaluation of daptomycin (LY146032). J. Antibiot (Tokyo) 41, 1093–1105.
- Deshpande, L.M., Ashcraft, D.S., Kahn, H.P., Pankey, G., Jones, R.N., Farrell, D.J., Mendes, R.E., 2015. Detection of a new *cf*-like gene, *cf*(B), in Enterococcus faecium isolates recovered from human specimens in the United States as part of the SENTRY Antimicrobial Surveillance Program. Antimicrob. Agents Chemother. 59, 6256–6261.
- Diaz, L., Kiratisin, P., Mendes, R.E., Panesso, D., Singh, K.V., Arias, C.A., 2012. Transferable plasmid-mediated resistance to linezolid due to *cf* in a human clinical isolate of Enterococcus faecalis. Antimicrob. Agents Chemother. 56, 3917–3922.
- Diaz, L., Tran, T.T., Munita, J.M., Miller, W.R., Rincon, S., Carvajal, L.P., Wollam, A., Reyes, J., Panesso, D., Rojas, N.L.S., Shamoo, Y., Murray, B.E., Weinstock, G.M., Arias, C.A., 2014. Whole-genome analyses of Enterococcus faecium isolates with diverse daptomycin MICs. Antimicrob. Agents Chemother. 58, 4527–4534.
- DiPippo, A.J., Tverdek, F.P., Tarrand, J.J., Munita, J.M., Tran, T.T., Arias, C.A., Shelburne, S.A., Aitken, S.L., 2016. Daptomycin non-susceptible Enterococcus faecium in leukemia patients: role of prior daptomycin exposure. J. Infect. 74, 5.
- Downing, M.A., Xiong, J., Eshaghi, A., McGeer, A., Patel, S.N., Johnstone, J., 2015. Vancomycin-variable enterococcal bacteremia. J. Clin. Microbiol. 53, 3951–3953.
- Eliopoulos, G.M., Thauvin, C., Gerson, B., Moellering Jr, R.C., 1985. In vitro activity and mechanism of action of A21978C1, a novel cyclic lipopeptide antibiotic. Antimicrob. Agents Chemother. 27, 357–362.
- Fiedler, S., Bender, J.K., Klare, I., Halbedel, S., Grohmann, E., Szwedzyk, U., Werner, G., 2016. Tigecycline resistance in clinical isolates of Enterococcus faecium is mediated by an upregulation of plasmid-encoded tetracycline determinants tet(L) and tet(M). J. Antimicrob. Chemother. 71, 871–881.
- Fines, M., Leclercq, R., 2000. Activity of linezolid against Gram-positive cocci possessing genes conferring resistance to protein synthesis inhibitors. J. Antimicrob. Chemother. 45, 797–802.
- Flamm, R.K., Mendes, R.E., Hogan, P.A., Streit, J.M., Ross, J.E., Jones, R.N., 2016. Linezolid Surveillance Results for the United States (LEADER Surveillance Program 2014). Antimicrob. Agents Chemother. 60, 2273–2280.
- Forsberg, K.J., Patel, S., Wencewicz, T.A., Dantas, G., 2015. The Tetracycline Destructases: A Novel Family of Tetracycline-Inactivating Enzymes. Chem. Biol. 22, 888–897.
- Franz, C.M., Huch, M., Abriouel, H., Holzapfel, W., Galvez, A., 2011. Enterococci as probiotics and their implications in food safety. Int. J. Food Microbiol. 151, 125–140.
- Freitas, A.R., Novais, C., Correia, R., Monteiro, M., Coque, T.M., Peixe, L., 2011. Non-susceptibility to tigecycline in enterococci from hospitalised patients, food products and community sources. Int. J. Antimicrob. Agents 38, 174–176.
- Freitas, A.R., Novais, C., Read, A., Alves, V., Peixe, L., 2016. Co-infection with three linezolid-resistant Enterococcus faecium ST117 strain variants: what are we missing in diagnosis? Int. J. Antimicrob. Agents 47, 500–501.
- Gawryszywska, I., Zabicka, D., Hryniewicz, W., Sadowy, E., 2017. Linezolid-resistant enterococci in Polish hospitals: species, clonality and determinants of linezolid resistance. Eur. J. Clin. Microbiol. Infect. Dis. 36, 1279–1286.
- Gomez-Gil, R., Romero-Gomez, M.P., Garcia-Arias, A., Ubeda, M.G., Busselo, M.S., Cisterna, R., Gutierrez-Altes, A., Mingorance, J., 2009. Nosocomial outbreak of linezolid-resistant Enterococcus faecalis infection in a tertiary care hospital. Diagn. Microbiol. Infect. Dis. 65, 175–179.
- Gonzales, R.D., Schreckenberger, P.C., Graham, M.B., Kelkar, S., DenBesten, K., Quinn, J.P., 2001. Infections due to vancomycin-resistant Enterococcus faecium resistant to linezolid. Lancet 357, 1179.
- Gonzalez-Ruiz, A., Gargalianos-Kakolyris, P., Timerman, A., Sarma, J., Jose Gonzalez Ramallo, V., Bouylout, K., Trostmann, U., Pathan, R., Hamed, K., 2015. Daptomycin in the clinical setting: 8-year experience with Gram-positive bacterial infections from the EU-CORE(SM) registry. Adv. Ther. 32, 496–509.
- Grossman, T.H., 2016. Tetracycline antibiotics and resistance. Cold Spring Harb. Perspect. Med. 6, a025387.
- Guo, D., Liu, Y., Han, C., Chen, Z., Ye, X., 2018. Phenotypic and molecular characteristics of methicillin-resistant and methicillin-susceptible Staphylococcus aureus isolated from pigs: implication for livestock-association markers and vaccine strategies. Infect. Drug Resist. 11, 1299–1307.
- Hachmann, A.B., Angert, E.R., Helmann, J.D., 2009. Genetic analysis of factors affecting susceptibility of Bacillus subtilis to daptomycin. Antimicrob. Agents Chemother. 53, 1598–1609.
- Hall, A.D., Steed, M.E., Arias, C.A., Murray, B.E., Rybak, M.J., 2012. Evaluation of standard- and high-dose daptomycin versus linezolid against vancomycin-resistant Enterococcus isolates in an in vitro pharmacokinetic/pharmacodynamic model with simulated endocardial vegetations. Antimicrob. Agents Chemother. 56, 3174–3180.
- Hansen, T.A., Pedersen, M.S., Nielsen, L.G., Ma, C.M.G., Soes, L.M., Worning, P., Ostergaard, C., Westh, H., Pinholt, M., Schonning, K., 2018. Emergence of a vancomycin-variable Enterococcus faecium ST1421 strain containing a deletion in vanX. J. Antimicrob. Chemother. 73, 2936–2940.
- Hayakawa, K., Marchaim, D., Pogue, J.M., Ho, K., Parveen, S., Nanjireddy, P., Sunkara, B., Singla, M., Jagadeesh, K.K., Moshos, J.A., Bommarito, S., Mrour, R., Farhat, M., Obeid, T., Chaudhry, A., Vadlamudi, G., Lephart, P.R., Martin, E.T., Rybak, M.J., Kaye, K.S., 2012. Predictors and outcomes of linezolid-resistant vancomycin-resistant Enterococcus: a case-control study. Am. J. Infect. Control 40 pp. e261–263.
- He, T., Shen, Y., Schwarz, S., Cai, J., Lv, Y., Li, J., Fessler, A.T., Zhang, R., Wu, C., Shen, J., Wang, Y., 2016. Genetic environment of the transferable oxazolidinone/phenicol resistance gene *optrA* in Enterococcus faecalis isolates of human and animal origin. J. Antimicrob. Chemother. 71, 1466–1473.
- Hegstad, K., Mikalson, T., Coque, T.M., Werner, G., Sundsfjord, A., 2010. Mobile genetic elements and their contribution to the emergence of antimicrobial resistant Enterococcus faecalis and Enterococcus faecium. Clin. Microbiol. Infect. 16, 541–554.
- Hegstad, K., Longva, J.A., Hide, R., Aasnaes, B., Lund, T.M., Simonsen, G.S., 2014. Cluster of linezolid-resistant Enterococcus faecium ST117 in Norwegian hospitals. Scand. J. Infect. Dis. 46, 712–715.
- Herrero, I.A., Issa, N.C., Patel, R., 2002. Nosocomial spread of linezolid-resistant, vancomycin-resistant Enterococcus faecium. N. Engl. J. Med. 346, 867–869.
- Ho, S.W., Jung, D., Calhoun, J.R., Lear, J.D., Okon, M., Scott, W.R., Hancock, R.E., Straus, S.K., 2008. Effect of divalent cations on the structure of the antibiotic daptomycin. Eur. Biophys. J. 37, 421–433.
- Hoban, D.J., Reinert, R.R., Bouchillon, S.K., Dowzicky, M.J., 2015. Global in vitro activity of tigecycline and comparator agents: tigecycline Evaluation and Surveillance Trial 2004–2013. Ann. Clin. Microbiol. Antimicrob. 14, 27.
- Honsa, E.S., Cooper, V.S., Mhaisen, M.N., Frank, M., Shaker, J., Iverson, A., Rubnitz, J., Hayden, R.T., Lee, R.E., Rock, C.O., Tuomanen, E.L., Wolf, J., Rosch, J.W., 2017. RelA mutant Enterococcus faecium with multiantibiotic tolerance arising in an immunocompromised host. MBio 8.
- Hua, R., Xia, Y., Wu, W., Yan, J., Yang, M., 2018. Whole transcriptome analysis reveals

- potential novel mechanisms of low-level linezolid resistance in *Enterococcus faecalis*. *Gene* 647, 143–149.
- Huang, J., Chen, L., Wu, Z., Wang, L., 2017. Retrospective analysis of genome sequences revealed the wide dissemination of *optrA* in Gram-positive bacteria. *J. Antimicrob. Chemother.* 72, 614–616.
- Humphries, R.M., Kelesidis, T., Tewhey, R., Rose, W.E., Schork, N., Nizet, V., Sakoulas, G., 2012. Genotypic and phenotypic evaluation of the evolution of high-level daptomycin nonsusceptibility in vancomycin-resistant *Enterococcus faecium*. *Antimicrob. Agents Chemother.* 56, 6051–6053.
- Humphries, R.M., Pollett, S., Sakoulas, G., 2013. A current perspective on daptomycin for the clinical microbiologist. *Clin. Microbiol. Rev.* 26, 759–780.
- Jenner, L., Starosta, A.L., Terry, D.S., Mikolajka, A., Filonava, L., Yusupov, M., Blanchard, S.C., Wilson, D.N., Yusupova, G., 2013. Structural basis for potent inhibitory activity of the antibiotic tigecycline during protein synthesis. *Proc Natl Acad Sci U S A* 110, 3812–3816.
- Johnson, A.P., Uttley, A.H., Woodford, N., George, R.C., 1990. Resistance to vancomycin and teicoplanin: an emerging clinical problem. *Clin. Microbiol. Rev.* 3, 280–291.
- Jung, D., Rozek, A., Okon, M., Hancock, R.E., 2004. Structural transitions as determinants of the action of the calcium-dependent antibiotic daptomycin. *Chem. Biol.* 11, 949–957.
- Jung, D., Powers, J.P., Straus, S.K., Hancock, R.E., 2008. Lipid-specific binding of the calcium-dependent antibiotic daptomycin leads to changes in lipid polymorphism of model membranes. *Chem. Phys. Lipids* 154, 120–128.
- Jung, Y.H., Lee, Y.S., Lee, S.Y., Yoo, J.S., Yoo, J.I., Kim, H.S., Kim, O., Yu, J.Y., 2014. Structure and transfer of the *vanA* cluster in *vanA*-positive, vancomycin-susceptible *Enterococcus faecium*, and its revertant mutant. *Diagn. Microbiol. Infect. Dis.* 80, 148–150.
- Kaatz, G.W., McAleese, F., Seo, S.M., 2005. Multidrug resistance in *Staphylococcus aureus* due to overexpression of a novel multidrug and toxin extrusion (MATE) transport protein. *Antimicrob. Agents Chemother.* 49, 1857–1864.
- Kainer, M.A., Devasia, R.A., Jones, T.F., Simmons, B.P., Melton, K., Chow, S., Broyles, J., Moore, K.L., Craig, A.S., Schaffner, W., 2007. Response to emerging infection leading to outbreak of linezolid-resistant enterococci. *Emerg Infect Dis* 13, 1024–1030.
- Klare, I., Fleige, C., Geringer, U., Thurmer, A., Bender, J., Mutters, N.T., Mischnik, A., Werner, G., 2015. Increased frequency of linezolid resistance among clinical *Enterococcus faecium* isolates from German hospital patients. *J. Glob. Antimicrob. Resist.* 3, 128–131.
- Kloss, P., Xiong, L., Shinabarger, D.L., Mankin, A.S., 1999. Resistance mutations in 23 S rRNA identify the site of action of the protein synthesis inhibitor linezolid in the ribosomal peptidyl transferase center. *J. Mol. Biol.* 294, 93–101.
- Klupp, E.M., Both, A., Belmar Campos, C., Buttner, H., König, C., Christopeit, M., Christner, M., Aepfelbacher, M., Rohde, H., 2016. Tedizolid susceptibility in linezolid- and vancomycin-resistant *Enterococcus faecium* isolates. *Eur. J. Clin. Microbiol. Infect. Dis.* 35, 1957–1961.
- Kolonen, A., Sinisalo, M., Huttunen, R., Syrjänen, J., Aittoniemi, J., Huhtala, H., Sankelo, M., Rintala, H., Raty, R., Jantunen, E., Nousiainen, T., Saily, M., Kauppila, M., Itälä-Remes, M., Ollikainen, H., Rauhala, A., Koistinen, P., Elonen, E., Finnish Leukemia G., 2017. Bloodstream infections in acute myeloid leukemia patients treated according to the Finnish Leukemia Group AML-2003 protocol - a prospective nationwide study. *Infect. Dis. Lond. (Lond)* 49, 799–808.
- Krawczyk, B., Samet, A., Bronk, M., Hellmann, A., Kur, J., 2004. Emerging linezolid-resistant, vancomycin resistant *Enterococcus faecium* from a patient of a haematological unit in Poland. *Pol. J. Microbiol.* 53, 193–196.
- Kresken, M., Leitner, E., Seifert, H., Peters, G., von Eiff, C., 2009. Susceptibility of clinical isolates of frequently encountered bacterial species to tigecycline one year after the introduction of this new class of antibiotics: results of the second multicentre surveillance trial in Germany (G-TEST II, 2007). *Eur. J. Clin. Microbiol. Infect. Dis.* 28, 1007–1011.
- Kresken, M., Becker, K., Seifert, H., Leitner, E., Korber-Irrgang, B., von Eiff, C., Loschmann, P.A., Study, G., 2011. Resistance trends and in vitro activity of tigecycline and 17 other antimicrobial agents against Gram-positive and Gram-negative organisms, including multidrug-resistant pathogens, in Germany. *Eur. J. Clin. Microbiol. Infect. Dis.* 30, 1095–1103.
- Krull, M., Klare, I., Ross, B., Trenchel, R., Beelen, D.W., Todt, D., Steinmann, E., Buer, J., Rath, P.M., Steinmann, J., 2016. Emergence of linezolid- and vancomycin-resistant *Enterococcus faecium* in a department for hematologic stem cell transplantation. *Antimicrob. Resist. Infect. Control* 5, 31.
- Kuch, A., Willems, R.J., Werner, G., Coque, T.M., Hammerum, A.M., Sundsfjord, A., Klare, I., Ruiz-Garbajosa, P., Simonsen, G.S., van Luit-Asbroek, M., Hryniewicz, W., Sadowy, E., 2012. Insight into antimicrobial susceptibility and population structure of contemporary human *Enterococcus faecalis* isolates from Europe. *J. Antimicrob. Chemother.* 67, 551–558.
- Kumaraswami, M., Schuman, J.T., Seo, S.M., Kaatz, G.W., Brennan, R.G., 2009. Structural and biochemical characterization of MepR, a multidrug binding transcription regulator of the *Staphylococcus aureus* multidrug efflux pump MepA. *Nucleic Acids Res.* 37, 1211–1224.
- Lebreton, F., van Schaik, W., McGuire, A.M., Godfrey, P., Griggs, A., Mazumdar, V., Corander, J., Cheng, L., Saif, S., Young, S., Zeng, Q., Wortman, J., Birren, B., Willems, R.J., Earl, A.M., Gilmore, M.S., 2013. Emergence of epidemic multidrug-resistant *Enterococcus faecium* from animal and commensal strains. *MBio* 4.
- Lellek, H., Franke, G.C., Ruckert, C., Wolters, M., Wolschke, C., Christner, M., Buttner, H., Alawi, M., Kroger, N., Rohde, H., 2015. Emergence of daptomycin non-susceptibility in colonizing vancomycin-resistant *Enterococcus faecium* isolates during daptomycin therapy. *Int. J. Med. Microbiol.* 305, 902–909.
- Lester, C.H., Sandvang, D., Olsen, S.S., Schonheyder, H.C., Jarlov, J.O., Bangsborg, J., Hansen, D.S., Jensen, T.G., Fridmodt-Møller, N., Hammerum, A.M., Group, D.S., 2008. Emergence of ampicillin-resistant *Enterococcus faecium* in Danish hospitals. *J. Antimicrob. Chemother.* 62, 1203–1206.
- Lewis, J.D., Enfield, K.B., Cox, H.L., Mathers, A.J., Sifri, C.D., 2016. A single-center experience with infections due to daptomycin-nonsusceptible *Enterococcus faecium* in liver transplant recipients. *Transpl. Infect. Dis.* 18, 341–353.
- Li, D., Wang, Y., Schwarz, S., Cai, J., Fan, R., Li, J., Fessler, A.T., Zhang, R., Wu, C., Shen, J., 2016. Co-location of the oxazolidinone resistance genes *optrA* and *cfr* on a multiresistance plasmid from *Staphylococcus sciuri*. *J. Antimicrob. Chemother.* 71, 1474–1478.
- Linkevicius, M., Sandegren, L., Andersson, D.I., 2016. Potential of tetracycline resistance proteins to evolve tigecycline resistance. *Antimicrob. Agents Chemother.* 60, 789–796.
- Liu, Y., Wang, Y., Wu, C., Shen, Z., Schwarz, S., Du, X.D., Dai, L., Zhang, W., Zhang, Q., Shen, J., 2012. First report of the multidrug resistance gene *cfr* in *Enterococcus faecalis* of animal origin. *Antimicrob. Agents Chemother.* 56, 1650–1654.
- Liu, Y., Wang, Y., Schwarz, S., Li, Y., Shen, Z., Zhang, Q., Wu, C., Shen, J., 2013. Transferable multiresistance plasmids carrying *cfr* in *Enterococcus* spp. from swine and farm environment. *Antimicrob. Agents Chemother.* 57, 42–48.
- Liu, Y., Wang, Y., Dai, L., Wu, C., Shen, J., 2014a. First report of multiresistance gene *cfr* in *Enterococcus* species *casseliflavus* and *gallinarum* of swine origin. *Vet. Microbiol.* 170, 352–357.
- Liu, Y., Wang, Y., Schwarz, S., Wang, S., Chen, L., Wu, C., Shen, J., 2014b. Investigation of a multiresistance gene *cfr* that fails to mediate resistance to phenolics and oxazolidinones in *Enterococcus faecalis*. *J. Antimicrob. Chemother.* 69, 892–898.
- Locke, J.B., Zurenko, G.E., Shaw, K.J., Bartizal, K., 2014. Tedizolid for the management of human infections: in vitro characteristics. *Clin. Infect. Dis.* 58 (Suppl. 1) pp. S35–42.
- Loens, K., Verkroost, S., Ieven, M., Goossens, H., 2016. First 2 cases of vancomycin variable *enterococcus faecium* in Belgium. 26th European Congress of Clinical Microbiology and Infectious Diseases.
- Long, K.S., Poehlsgaard, J., Kehrenberg, C., Schwarz, S., Vester, B., 2006. The Cfr rRNA methyltransferase confers resistance to Phenolics, Lincosamides, Oxazolidinones, Pleuromutins, and Streptogramin A antibiotics. *Antimicrob. Agents Chemother.* 50, 2500–2505.
- Lupien, A., Gingras, H., Leprohon, P., Ouellette, M., 2015. Induced tigecycline resistance in *Streptococcus pneumoniae* mutants reveals mutations in ribosomal proteins and rRNA. *J. Antimicrob. Chemother.* 70, 2973–2980.
- Marco, F., Dowdzicky, M.J., 2016. Antimicrobial susceptibility among important pathogens collected as part of the Tigecycline Evaluation and Surveillance Trial (T.E.S.T.) in Spain, 2004–2014. *J. Glob. Antimicrob. Resist.* 6, 50–56.
- Marshall, S.H., Donskey, C.J., Hutton-Thomas, R., Salata, R.A., Rice, L.B., 2002. Gene dosage and linezolid resistance in *Enterococcus faecium* and *Enterococcus faecalis*. *Antimicrob. Agents Chemother.* 46, 3334–3336.
- Matono, T., Hayakawa, K., Hirai, R., Tanimura, A., Yamamoto, K., Fujiya, Y., Mawatari, M., Kutsuna, S., Takeshita, N., Mezaki, K., Ohmagari, N., Miyoshi-Akiyama, T., 2016. Emergence of a daptomycin-non-susceptible *Enterococcus faecium* strain that encodes mutations in DNA repair genes after high-dose daptomycin therapy. *BMC Res. Notes* 9, 197.
- McAleese, F., Petersen, P., Ruzin, A., Dunman, P.M., Murphy, E., Projan, S.J., Bradford, P.A., 2005. A novel MATE family efflux pump contributes to the reduced susceptibility of laboratory-derived *Staphylococcus aureus* mutants to tigecycline. *Antimicrob. Agents Chemother.* 49, 1865–1871.
- Mendes, R.E., Deshpande, L.M., Jones, R.N., 2014. Linezolid update: stable in vitro activity following more than a decade of clinical use and summary of associated resistance mechanisms. *Drug Resist. Updat.* 17, 1–12.
- Mendes, R.E., Castanheira, M., Farrell, D.J., Flamm, R.K., Sader, H.S., Jones, R.N., 2016a. Longitudinal (2001–14) analysis of enterococci and VRE causing invasive infections in European and US hospitals, including a contemporary (2010–13) analysis of oritavancin in vitro potency. *J. Antimicrob. Chemother.* 71, 3453–3458.
- Mendes, R.E., Deshpande, L.M., Castanheira, M., Flamm, R.K., 2016b. Evolving linezolid resistance mechanisms in a worldwide collection of enterococcal clinical isolates: results from the SENTRY antimicrobial surveillance program. *Microbe*. 2016, Boston, MA, USA.
- Mendes, R.E., Hogan, P.A., Jones, R.N., Sader, H.S., Flamm, R.K., 2016c. Surveillance for linezolid resistance via the Zyvox(R) Annual Appraisal of Potency and Spectrum (ZAAPS) programme (2014): evolving resistance mechanisms with stable susceptibility rates. *J. Antimicrob. Chemother.* 71, 1860–1865.
- Miller, C., Kong, J., Tran, T.T., Arias, C.A., Saxer, G., Shamoo, Y., 2013. Adaptation of *Enterococcus faecalis* to daptomycin reveals an ordered progression to resistance. *Antimicrob. Agents Chemother.* 57, 5373–5383.
- Miller, W.R., Munita, J.M., Arias, C.A., 2014. Mechanisms of antibiotic resistance in enterococci. *Expert Rev. Anti. Ther.* 12, 1221–1236.
- Miller, W.R., Bayer, A.S., Arias, C.A., 2016. Mechanism of action and resistance to daptomycin in *Staphylococcus aureus* and enterococci. *Cold Spring Harb. Perspect. Med.* 6.
- Mishra, N.N., Bayer, A.S., Tran, T.T., Shamoo, Y., Mileykovskaya, E., Dowhan, W., Guan, Z., Arias, C.A., 2012. Daptomycin resistance in enterococci is associated with distinct alterations of cell membrane phospholipid content. *PLoS One* 7, e43958.
- Montero, C.I., Stock, F., Murray, P.R., 2008. Mechanisms of resistance to daptomycin in *Enterococcus faecium*. *Antimicrob. Agents Chemother.* 52, 1167–1170.
- Moore, I.F., Hughes, D.W., Wright, G.D., 2005. Tigecycline is modified by the flavin-dependent monooxygenase TetX. *Biochemistry* 44, 11829–11835.
- Munita, J.M., Tran, T.T., Diaz, L., Panesso, D., Reyes, J., Murray, B.E., Arias, C.A., 2013. A *liaf* codon deletion abolishes daptomycin bactericidal activity against vancomycin-resistant *Enterococcus faecalis*. *Antimicrob. Agents Chemother.* 57, 2831–2833.
- Muraih, J.K., Pearson, A., Silverman, J., Palmer, M., 2011. Oligomerization of daptomycin on membranes. *Biochim. Biophys. Acta* 1808, 1154–1160.

- Murrah, J.K., Harris, J., Taylor, S.D., Palmer, M., 2012. Characterization of daptomycin oligomerization with perylene excimer fluorescence: stoichiometric binding of phosphatidylglycerol triggers oligomer formation. *Biochim. Biophys. Acta* 1818, 673–678.
- Murray, B.E., 1990. The life and times of the Enterococcus. *Clin. Microbiol. Rev.* 3, 46–65.
- Muthaiyan, A., Silverman, J.A., Jayaswal, R.K., Wilkinson, B.J., 2008. Transcriptional profiling reveals that daptomycin induces the *Staphylococcus aureus* cell wall stress stimulon and genes responsive to membrane depolarization. *Antimicrob. Agents Chemother.* 52, 980–990.
- Mutnick, A.H., Enne, V., Jones, R.N., 2003. Linezolid resistance since 2001: SENTRY antimicrobial surveillance program. *Ann. Pharmacother.* 37, 769–774.
- Mutters, N.T., Werner, G., Tacconelli, E., Mischnick, A., 2015. [Treatment options for serious infections caused by vancomycin-resistant enterococci]. *Dtsch. Med. Wochenschr.* 140, 42–45.
- Navarro-Gonzalez, N., Casas-Diaz, E., Porrero, C.M., Mateos, A., Dominguez, L., Lavin, S., Serrano, E., 2013. Food-borne zoonotic pathogens and antimicrobial resistance of indicator bacteria in urban wild boars in Barcelona, Spain. *Vet. Microbiol.* 167, 686–689.
- Niebel, M., Quick, J., Prieto, A.M., Hill, R.L., Pike, R., Huber, D., David, M., Hornsey, M., Wareham, D., Oppenheim, B., Woodford, N., van Schaik, W., Loman, N., 2015. Deletions in a ribosomal protein-coding gene are associated with tigecycline resistance in *Enterococcus faecium*. *Int. J. Antimicrob. Agents* 46, 572–575.
- Ntokou, E., Stathopoulos, C., Kristo, I., Dimitroulia, E., Labrou, M., Vasdeki, A., Makris, D., Zakynthinos, E., Tsakris, A., Pournaras, S., 2012. Intensive care unit dissemination of multiple clones of linezolid-resistant *Enterococcus faecalis* and *Enterococcus faecium*. *J. Antimicrob. Chemother.* 67, 1819–1823.
- Numminen, E., Gutmann, M., Shubin, M., Marttinen, P., Meric, G., van Schaik, W., Coque, T.M., Baquero, F., Willems, R.J., Sheppard, S.K., Feil, E.J., Hanage, W.P., Corander, J., 2016. The impact of host metapopulation structure on the population genetics of colonizing bacteria. *J. Theor. Biol.* 396, 53–62.
- O'Driscoll, C., Murphy, V., Doyle, O., Wrenn, C., Flynn, A., O'Flaherty, N., Fenelon, L.E., Schaffer, K., FitzGerald, S.F., 2015. First outbreak of linezolid-resistant vancomycin-resistant *Enterococcus faecium* in an Irish hospital, February to September 2014. *J. Hosp. Infect.* 91, 367–370.
- Olson, M.W., Ruzin, A., Feyfant, E., Rush 3rd, T.S., O'Connell, J., Bradford, P.A., 2006. Functional, biophysical, and structural bases for antibacterial activity of tigecycline. *Antimicrob. Agents Chemother.* 50, 2156–2166.
- Palmer, K.L., Daniel, A., Hardy, C., Silverman, J., Gilmore, M.S., 2011. Genetic basis for daptomycin resistance in enterococci. *Antimicrob. Agents Chemother.* 55, 3345–3356.
- Panesso, D., Reyes, J., Gaston, E.P., Deal, M., Londono, A., Nigo, M., Munita, J.M., Miller, W.R., Shamoo, Y., Tran, T.T., Arias, C.A., 2015. Deletion of *liaR* reverses daptomycin resistance in *Enterococcus faecium* independent of the genetic background. *Antimicrob. Agents Chemother.* 59, 7327–7334.
- Pankey, G.A., 2005. Tigecycline. *J. Antimicrob. Chemother.* 56, 470–480.
- Patel, S.N., Memari, N., Shahinas, D., Toye, B., Jamieson, F.B., Farrell, D.J., 2013. Linezolid resistance in *Enterococcus faecium* isolated in Ontario, Canada. *Diagn. Microbiol. Infect. Dis.* 77, 350–353.
- Pericas, J.M., Garcia-de-la-Maria, C., Brunet, M., Armero, Y., Garcia-Gonzalez, J., Casals, G., Almela, M., Quintana, E., Falces, C., Ninot, S., Fuster, D., Llopis, J., Marco, F., Moreno, A., Miro, J.M., Hospital Clinic Endocarditis Study, G., 2017. Early in vitro development of daptomycin non-susceptibility in high-level aminoglycoside-resistant *Enterococcus faecalis* predicts the efficacy of the combination of high-dose daptomycin plus ampicillin in an in vivo model of experimental endocarditis. *J. Antimicrob. Chemother.* 72, 1714–1722.
- Pfaller, M.A., Mendes, R.E., Streit, J.M., Hogan, P.A., Flamm, R.K., 2017a. Five-Year Summary of In Vitro Activity and Resistance Mechanisms of Linezolid against Clinically Important Gram-Positive Cocci in the United States from the LEADER Surveillance Program (2011 to 2015). *Antimicrob. Agents Chemother.* 61.
- Pfaller, M.A., Mendes, R.E., Streit, J.M., Hogan, P.A., Flamm, R.K., 2017b. ZAAPs Program results for 2015: an activity and spectrum analysis of linezolid using clinical isolates from medical centres in 32 countries. *J. Antimicrob. Chemother.* 72, 3093–3099.
- Pioletti, M., Schlunzen, F., Harms, J., Zarivach, R., Gluhmann, M., Avila, H., Bashan, A., Bartels, H., Auerbach, T., Jacobi, C., Hartsch, T., Yonath, A., Franceschi, F., 2001. Crystal structures of complexes of the small ribosomal subunit with tetracycline, edeine and IF3. *EMBO J.* 20, 1829–1839.
- Plosker, G.L., Figgitt, D.P., 2005. Linezolid: a pharmaco-economic review of its use in serious Gram-positive infections. *Pharmacoeconomics* 23, 945–964.
- Pogliano, J., Pogliano, N., Silverman, J.A., 2012. Daptomycin-mediated reorganization of membrane architecture causes mislocalization of essential cell division proteins. *J. Bacteriol.* 194, 4494–4504.
- Power, R.A., Parkhill, J., de Oliveira, T., 2017. Microbial genome-wide association studies: lessons from human GWAS. *Nat. Rev. Genet.* 18, 41–50.
- Rahim, S., Pillai, S.K., Gold, H.S., Venkataraman, L., Inglima, K., Press, R.A., 2003. Linezolid-resistant, vancomycin-resistant *Enterococcus faecium* infection in patients without prior exposure to linezolid. *Clin. Infect. Dis.* 36 pp. E146–148.
- Rasmussen, B.A., Gluzman, Y., Tally, F.P., 1994. Inhibition of protein synthesis occurring on tetracycline-resistant, TetM-protected ribosomes by a novel class of tetracyclines, the glycyliclins. *Antimicrob. Agents Chemother.* 38, 1658–1660.
- Raven, K.E., Reuter, S., Reynolds, R., Brodrick, H.J., Russell, J.E., Torok, M.E., Parkhill, J., Peacock, S.J., 2016. A decade of genomic history for healthcare-associated *Enterococcus faecium* in the United Kingdom and Ireland. *Genome Res.* 26, 1388–1396.
- Reyes, J., Panesso, D., Tran, T.T., Mishra, N.N., Cruz, M.R., Munita, J.M., Singh, K.V., Yeaman, M.R., Murray, B.E., Shamoo, Y., Garsin, D., Bayer, A.S., Arias, C.A., 2015. A *liaR* deletion restores susceptibility to daptomycin and antimicrobial peptides in multidrug-resistant *Enterococcus faecalis*. *J. Infect. Dis.* 211, 1317–1325.
- Sader, H.S., Farrell, D.J., Flamm, R.K., Jones, R.N., 2014a. Daptomycin activity tested against 164457 bacterial isolates from hospitalised patients: summary of 8 years of a Worldwide Surveillance Programme (2005–2012). *Int. J. Antimicrob. Agents* 43, 465–469.
- Sader, H.S., Farrell, D.J., Flamm, R.K., Jones, R.N., 2014b. Variation in potency and spectrum of tigecycline activity against bacterial strains from U.S. Medical centers since its approval for clinical use (2006 to 2012). *Antimicrob. Agents Chemother.* 58, 2274–2280.
- Sader, H.S., Farrell, D.J., Flamm, R.K., Jones, R.N., 2015. Analysis of 5-year trends in daptomycin activity tested against *Staphylococcus aureus* and enterococci from European and US hospitals (2009–2013). *J. Glob. Antimicrob. Resist.* 3, 161–165.
- Sanchez-Diaz, A.M., Cuartero, C., Lozano, S., Rodriguez, J.D., Alonso, J.M., Quiles-Melero, I., Lopez, J., Canton, R., Ruiz-Garbajosa, P., 2014. Emergence and long-lasting persistence of linezolid-resistant *Enterococcus faecium*-ST117 in an oncohematologic patient after a nine-day course of linezolid. *Microb. Drug Resist.* 20, 17–21.
- Sanchez-Gomez, J.C., Fraile-Malmierca, F., Valverde-Romero, E.D., Sanchez, M., Garcia-Rodriguez, J.A., Garcia-Sanchez, J.E., 2006. Linezolid-resistant *Enterococcus faecalis*: first report in Spain. *J. Chemother.* 18, 440–442.
- Sandoval, N., Grau, S., Sorli, L., Montero, M., Esteve, E., Horcajada, J.P., 2015. Clinical experience with the use of daptomycin in a tertiary care teaching hospital in Barcelona, Spain. *Future Microbiol.* 10, 1145–1154.
- Santos Arronches, A.F., 2014. Pesquisa de resistência a antibióticos em alguns enterococos intestinais provenientes de diferentes amostras de águas. *Biologica. Technica Lisboa*, Lisboa, pp. 87.
- Scheetz, M.H., Knechtel, S.A., Malczynski, M., Postelnick, M.J., Qi, C., 2008. Increasing incidence of linezolid-intermediate or -resistant, vancomycin-resistant *Enterococcus faecium* strains parallels increasing linezolid consumption. *Antimicrob. Agents Chemother.* 52, 2256–2259.
- Schwarz, S., Werckenthin, C., Kehrenberg, C., 2000. Identification of a plasmid-borne chloramphenicol-florfenicol resistance gene in *Staphylococcus sciuri*. *Antimicrob. Agents Chemother.* 44, 2530–2533.
- Scott, W.R., Baek, S.B., Jung, D., Hancock, R.E., Straus, S.K., 2007. NMR structural studies of the antibiotic lipopeptide daptomycin in DHPC micelles. *Biochim. Biophys. Acta* 1768, 3116–3126.
- Seedat, J., Zick, G., Klare, I., Konstabel, C., Weiler, N., Sahly, H., 2006. Rapid emergence of resistance to linezolid during linezolid therapy of an *Enterococcus faecium* infection. *Antimicrob. Agents Chemother.* 50, 4217–4219.
- Seifert, H., Dowzicky, M.J., 2009. A longitudinal analysis of antimicrobial susceptibility in clinical institutions in Germany as part of the Tigecycline Evaluation and Surveillance Trial (2004–2007). *Chemotherapy* 55, 241–252.
- Sharkey, L.K.R., O'Neill, A.J., 2018. Antibiotic resistance ABC-F proteins: bringing target protection into the limelight. *ACS Infect. Dis.* 4, 239–246.
- Shen, J., Wang, Y., Schwarz, S., 2013. Presence and dissemination of the multiresistance gene *cfr* in Gram-positive and Gram-negative bacteria. *J. Antimicrob. Chemother.* 68, 1697–1706.
- Shukla, B.S., Shelburne, S., Reyes, K., Kamboj, M., Lewis, J.D., Rincon, S.L., Reyes, J., Carvajal, L.P., Panesso, D., Sifri, C.D., Zervos, M.J., Pamer, E.G., Tran, T.T., Adachi, J., Munita, J.M., Hasbun, R., Arias, C.A., 2016. Influence of Minimum Inhibitory Concentration in Clinical Outcomes of *Enterococcus faecium* Bacteremia Treated With Daptomycin: Is it Time to Change the Breakpoint? *Clin. Infect. Dis.* 62, 1514–1520.
- Silverman, J.A., Perlmutter, N.G., Shapiro, H.M., 2003. Correlation of daptomycin bactericidal activity and membrane depolarization in *Staphylococcus aureus*. *Antimicrob. Agents Chemother.* 47, 2538–2544.
- Simner, P.J., Adam, H., Baxter, M., McCracken, M., Golding, G., Karlowsky, J.A., Nichol, K., Lagace-Wiens, P., Gilmour, M.W., 2015. A Canadian Antimicrobial Resistance, D.J. Hoban and G.G. Zhanel, Epidemiology of vancomycin-resistant enterococci in Canadian hospitals (CANWARD study, 2007 to 2013). *Antimicrob. Agents Chemother.* 59, 4315–4317.
- Sinel, C., Cosquer, T., Auzou, M., Goux, D., Giard, J.C., Cattoir, V., 2016a. Sequential steps of daptomycin resistance in *Enterococcus faecium* and reversion to hypersusceptibility through IS-mediated inactivation of the *liaFSR* operon. *J. Antimicrob. Chemother.* 71, 2793–2797.
- Sinel, C., Jaussaud, C., Auzou, M., Giard, J.C., Cattoir, V., 2016b. Mutant prevention concentrations of daptomycin for *Enterococcus faecium* clinical isolates. *Int. J. Antimicrob. Agents* 48, 449–452.
- Sivertsen, A., Pedersen, T., Larssen, K.W., Bergh, K., Ronning, T.G., Radtke, A., Hegstad, K., 2016. A silenced *vanA* gene cluster on a transferable plasmid caused an outbreak of vancomycin-variable enterococci. *Antimicrob. Agents Chemother.* 60, 4119–4127.
- Sivertsen, A., Janice, J., Sjögren, I., Hegstad, K., 2017. Vancomycin variable resistance may occur through creation of a novel promoter after deletions by polymerase slippage. 27th European Congress of Clinical Microbiology and Infectious Diseases.
- Sorlozano, A., Panesso, D., Navarro-Mari, J.M., Arias, C.A., Gutierrez-Fernandez, J., 2015. Characterization of daptomycin non-susceptible *Enterococcus faecium* producing urinary tract infection in a renal transplant recipient. *Rev. Esp. Quimioter.* 28, 207–209.
- Souli, M., Sakka, V., Galani, I., Antoniadou, A., Galani, L., Siafakas, N., Zerva, L., Fytrou, H., Tsiodras, S., Giamarellou, H., 2009. Colonisation with vancomycin- and linezolid-resistant *Enterococcus faecium* in a university hospital: molecular epidemiology and risk factor analysis. *Int. J. Antimicrob. Agents* 33, 137–142.
- Steed, M.E., Vidailac, C., Rose, W.E., Winterfield, P., Kaatz, G.W., Rybak, M.J., 2011. Characterizing vancomycin-resistant *Enterococcus* strains with various mechanisms of daptomycin resistance developed in an in vitro pharmacokinetic/pharmacodynamic model. *Antimicrob. Agents Chemother.* 55, 4748–4754.

- Stefani, S., Dowzicky, M.J., 2016. Assessment of the Activity of Tigecycline against Gram-Positive and Gram-Negative Organisms collected from Italy between 2012 and 2014, as Part of the Tigecycline Evaluation and Surveillance Trial (T.E.S.T.). *Pharmaceuticals Basel (Basel)* 9.
- Stein, G.E., Babinchak, T., 2013. Tigecycline: an update. *Diagn. Microbiol. Infect. Dis.* 75, 331–336.
- Streit, J.M., Flamm, R.K., Ross, J.E., Mendes, R.E., Jones, R.N., Hogan, P.A., 2015. Report of Linezolid Activity From the Linezolid Experience and Accurate Determination of Resistance (LEADER) Program for 2014: Monitoring Trends and Mechanisms. *IDWeek 2015*, San Diego, CA, USA.
- Sun, Y., Cai, Y., Liu, X., Bai, N., Liang, B., Wang, R., 2013. The emergence of clinical resistance to tigecycline. *Int. J. Antimicrob. Agents* 41, 110–116.
- Swaney, S.M., Aoki, H., Ganoza, M.C., Shinabarger, D.L., 1998. The oxazolidinone linezolid inhibits initiation of protein synthesis in bacteria. *Antimicrob. Agents Chemother.* 42, 3251–3255.
- Szakacs, T.A., Kalan, L., McConnell, M.J., Eshaghi, A., Shahinas, D., McGeer, A., Wright, G.D., Low, D.E., Patel, S.N., 2014. Outbreak of vancomycin-susceptible *Enterococcus faecium* containing the wild-type vanA gene. *J. Clin. Microbiol.* 52, 1682–1686.
- Tamang, M.D., Moon, D.C., Kim, S.R., Kang, H.Y., Lee, K., Nam, H.M., Jang, G.C., Lee, H.S., Jung, S.C., Lim, S.K., 2017. Detection of novel oxazolidinone and phenicol resistance gene *optrA* in enterococcal isolates from food animals and animal carcasses. *Vet. Microbiol.* 201, 252–256.
- Taylor, R., Butt, K., Scott, B., Zhang, T., Muraih, J.K., Mintzer, E., Taylor, S., Palmer, M., 2016. Two successive calcium-dependent transitions mediate membrane binding and oligomerization of daptomycin and the related antibiotic A54145. *Biochim. Biophys. Acta* 1858, 1999–2005.
- Tedim, A.P., Ruiz-Garbajosa, P., Corander, J., Rodriguez, C.M., Canton, R., Willems, R.J., Baquero, F., Coque, T.M., 2015. Population biology of intestinal enterococcus isolates from hospitalized and nonhospitalized individuals in different age groups. *Appl. Environ. Microbiol.* 81, 1820–1831.
- Tedim, A.P., Ruiz-Garbajosa, P., Rodriguez, M.C., Rodriguez-Banos, M., Lanza, V.F., Derdoy, L., Cardenas Zurita, G., Loza, E., Canton, R., Baquero, F., Coque, T.M., 2017. Long-term clonal dynamics of *Enterococcus faecium* strains causing bloodstream infections (1995–2015) in Spain. *J. Antimicrob. Chemother.* 72, 48–55.
- Thaker, M., Spanogiannopoulos, P., Wright, G.D., 2010. The tetracycline resistome. *Cell. Mol. Life Sci.* 67, 419–431.
- Thaker, M.N., Kalan, L., Waglechner, N., Eshaghi, A., Patel, S.N., Poutanen, S., Willey, B., Coburn, B., McGeer, A., Low, D.E., Wright, G.D., 2015. Vancomycin-variable enterococci can give rise to constitutive resistance during antibiotic therapy. *Antimicrob. Agents Chemother.* 59, 1405–1410.
- Thilesen, C.M., BJORANG, O., Skrede, T., Aronsen, T., Aasnaes, B., Sundsfjord, A., Hegstad, K., 2014. Emergence of mutation-based linezolid-resistant invasive *Enterococcus faecalis* in a haemodialysis patient in Norway. *APMIS* 122, 83–84.
- Tian, Y., Li, T., Zhu, Y., Wang, B., Zou, X., Li, M., 2014. Mechanisms of linezolid resistance in staphylococci and enterococci isolated from two teaching hospitals in Shanghai, China. *BMC Microbiol.* 14, 292.
- Top, J., Willems, R., Blok, H., de Regt, M., Jalink, K., Troelstra, A., Goorhuis, B., Bonten, M., 2007. Ecological replacement of *Enterococcus faecalis* by multiresistant clonal complex 17 *Enterococcus faecium*. *Clin. Microbiol. Infect.* 13, 316–319.
- Tran, T.T., Panesso, D., Gao, H., Roh, J.H., Munita, J.M., Reyes, J., Diaz, L., Lobos, E.A., Shamoo, Y., Mishra, N.N., Bayer, A.S., Murray, B.E., Weinstock, G.M., Arias, C.A., 2013a. Whole-genome analysis of a daptomycin-susceptible enterococcus faecium strain and its daptomycin-resistant variant arising during therapy. *Antimicrob. Agents Chemother.* 57, 261–268.
- Tran, T.T., Panesso, D., Mishra, N.N., Mileykovskaya, E., Guan, Z., Munita, J.M., Reyes, J., Diaz, L., Weinstock, G.M., Murray, B.E., Shamoo, Y., Dowhan, W., Bayer, A.S., Arias, C.A., 2013b. Daptomycin-resistant *Enterococcus faecalis* diverts the antibiotic molecule from the division septum and remodels cell membrane phospholipids. *MBio* 4.
- Tran, T.T., Munita, J.M., Arias, C.A., 2015. Mechanisms of drug resistance: daptomycin resistance. *Ann. N. Y. Acad. Sci.* 1354, 32–53.
- Uttley, A.H., George, R.C., Naidoo, J., Woodford, N., Johnson, A.P., Collins, C.H., Morrison, D., Gilfillan, A.J., Fitch, L.E., Heptonstall, J., 1989. High-level vancomycin-resistant enterococci causing hospital infections. *Epidemiol. Infect.* 103, 173–181.
- Vega, S., Dowzicky, M.J., 2017. Antimicrobial susceptibility among Gram-positive and Gram-negative organisms collected from the Latin American region between 2004 and 2015 as part of the Tigecycline Evaluation and Surveillance Trial. *Ann. Clin. Microbiol. Antimicrob.* 16, 50.
- Villa, L., Feudi, C., Fortini, D., Garcia-Fernandez, A., Carattoli, A., 2014. Genomics of KPC-producing *Klebsiella pneumoniae* sequence type 512 clone highlights the role of RamR and ribosomal S10 protein mutations in conferring tigecycline resistance. *Antimicrob. Agents Chemother.* 58, 1707–1712.
- Visalli, M.A., Murphy, E., Projan, S.J., Bradford, P.A., 2003. AcrAB multidrug efflux pump is associated with reduced levels of susceptibility to tigecycline (GAR-936) in *Proteus mirabilis*. *Antimicrob. Agents Chemother.* 47, 665–669.
- Vorobieva, V., Roer, L., Justesen, U.S., Hansen, F., Frimodt-Moller, N., Hasman, H., Hammerum, A.M., 2017. Detection of the *optrA* gene in a clinical ST16 *Enterococcus faecalis* isolate in Denmark. *J. Glob. Antimicrob. Resist.* 10, 12–13.
- Wang, G., Kamalakaran, S., Dhand, A., Huang, W., Ojaimi, C., Zhuge, J., Yee, L.L., Mayigowda, P., Surendraiah, P.K., Dimitrova, N., Fallon, J.T., 2014. Identification of a novel clone, ST736, among *Enterococcus faecium* clinical isolates and its association with daptomycin nonsusceptibility. *Antimicrob. Agents Chemother.* 58, 4848–4854.
- Wang, Y., Lv, Y., Cai, J., Schwarz, S., Cui, L., Hu, Z., Zhang, R., Li, J., Zhao, Q., He, T., Wang, D., Wang, Z., Shen, Y., Li, Y., Fessler, A.T., Wu, C., Yu, H., Deng, X., Xia, X., Shen, J., 2015. A novel gene, *optrA*, that confers transferable resistance to oxazolidinones and phenicols and its presence in *Enterococcus faecalis* and *Enterococcus faecium* of human and animal origin. *J. Antimicrob. Chemother.* 70, 2182–2190.
- Wang, X., Davlieva, M., Reyes, J., Panesso, D., Arias, C.A., Shamoo, Y., 2017. A novel phosphodiesterase of the GdpP family modulates cyclic di-AMP levels in response to cell membrane stress in daptomycin-resistant enterococci. *Antimicrob. Agents Chemother.*
- Weisser, M., Oostdijk, E.A., Willems, R.J., Bonten, M.J., Frei, R., Elzi, L., Halter, J., Widmer, A.F., Top, J., 2012. Dynamics of ampicillin-resistant *Enterococcus faecium* clones colonizing hospitalized patients: data from a prospective observational study. *BMC Infect. Dis.* 12, 68.
- Weisser, M., Theilacker, C., Tschudin Sutter, S., Babikir, R., Bertz, H., Gotting, T., Dettenkofer, M., Kern, W.V., Widmer, A.F., G. Hospital Infection Surveillance System for Patients With Haematologic/Oncologic Malignancies Study, 2017. Secular trends of bloodstream infections during neutropenia in 15 181 haematopoietic stem cell transplants: 13-year results from a European multicentre surveillance study (ONKO-KISS). *Clin. Microbiol. Infect.* 23, 854–859.
- Werner, G., Coque, T.M., Hammerum, A.M., Hope, R., Hryniewicz, W., Johnson, A., Klare, I., Kristinsson, K.G., Leclercq, R., Lester, C.H., Lillie, M., Novais, C., Olsson-Liljequist, B., Peixe, L.V., Sadowy, E., Simonsen, G.S., Top, J., Vuopio-Varkila, J., Willems, R.J., Witte, W., Woodford, N., 2008a. Emergence and spread of vancomycin resistance among enterococci in Europe. *Euro Surveill.* 13.
- Werner, G., Gfrörer, S., Fleige, C., Witte, W., Klare, I., 2008b. Tigecycline-resistant *Enterococcus faecalis* strain isolated from a German intensive care unit patient. *J. Antimicrob. Chemother.* 61, 1182–1183.
- Werner, G., Coque, T.M., Franz, C.M., Grohmann, E., Hegstad, K., Jensen, L., Schaik, W., Weaver, K., 2013. Antibiotic resistant enterococci-tales of a drug resistance gene trafficker. *Int. J. Med. Microbiol.* 303, 360–379.
- Werth, B.J., Steed, M.E., Ireland, C.E., Tran, T.T., Nonejuie, P., Murray, B.E., Rose, W.E., Sakoulas, G., Pogliano, J., Arias, C.A., Rybak, M.J., 2014. Defining daptomycin resistance prevention exposures in vancomycin-resistant *Enterococcus faecium* and *E. faecalis*. *Antimicrob. Agents Chemother.* 58, 5253–5261.
- Whang, D.W., Miller, L.G., Partain, N.M., McKinnell, J.A., 2013. Systematic review and meta-analysis of linezolid and daptomycin for treatment of vancomycin-resistant enterococcal bloodstream infections. *Antimicrob. Agents Chemother.* 57, 5013–5018.
- Willems, R.J., Top, J., van Schaik, W., Leavis, H., Bonten, M., Siren, J., Hanage, W.P., Corander, J., 2012. Restricted gene flow among hospital subpopulations of *Enterococcus faecium*. *MBio* 3 pp. e00151-00112.
- Wilson, D.N., 2014. Ribosome-targeting antibiotics and mechanisms of bacterial resistance. *Nat. Rev. Microbiol.* 12, 35–48.
- Wilson, D.N., Schluenzen, F., Harms, J.M., Starosta, A.L., Connell, S.R., Fucini, P., 2008. The oxazolidinone antibiotics perturb the ribosomal peptidyl-transferase center and effect tRNA positioning. *Proc Natl Acad Sci U S A* 105, 13339–13344.
- Zhang, T., Muraih, J.K., Mintzer, E., Tishbi, N., Desert, C., Silverman, J., Taylor, S., Palmer, M., 2013. Mutual inhibition through hybrid oligomer formation of daptomycin and the semisynthetic lipopeptide antibiotic CB-182,462. *Biochim. Biophys. Acta* 1828, 302–308.
- Zhang, T., Muraih, J.K., McCormick, B., Silverman, J., Palmer, M., 2014a. Daptomycin forms cation- and size-selective pores in model membranes. *Biochim. Biophys. Acta* 1838, 2425–2430.
- Zhang, T., Muraih, J.K., Tishbi, N., Herskowitz, J., Victor, R.L., Silverman, J., Uwumarenogie, S., Taylor, S.D., Palmer, M., Mintzer, E., 2014b. Cardiolipin prevents membrane translocation and permeabilization by daptomycin. *J. Biol. Chem.* 289, 11584–11591.
- Zhang, Y., Du, M., Chang, Y., Chen, L.A., Zhang, Q., 2017. Incidence, clinical characteristics, and outcomes of nosocomial *Enterococcus* spp. bloodstream infections in a tertiary-care hospital in Beijing, China: a four-year retrospective study. *Antimicrob. Resist. Infect. Control* 6, 73.
- Zhao, M., Liang, L., Ji, L., Chen, D., Zhang, Y., Zhu, Y., Patel, K., 2016. Similar efficacy and safety of daptomycin versus linezolid for treatment of vancomycin-resistant enterococcal bloodstream infections: a meta-analysis. *Int. J. Antimicrob. Agents* 48, 231–238.