

Monitoring Neonicotinoid Exposure for Bees in Rural and Peri-urban Areas of the UK during the Transition from Pre- to Post-moratorium

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Ecotoxicology and Human Environmental Health

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 during the transition from pre- to post-moratorium.

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ABSTRACT: Concerns regarding the impact of neonicotinoid exposure on bee populations recently led to an EU-wide moratorium on the use of certain neonicotinoids on flowering crops. Currently evidence regarding the impact, if any, the moratorium has had on bees' exposure is limited. We sampled pollen and nectar from bumblebee colonies in rural and peri-urban



19 habitats in three UK regions; Stirlingshire, Hertfordshire and Sussex. Colonies were sampled over 20 three years; prior to the ban (2013), during the initial implementation when some seed-treated winter-21 sown oilseed rape was still grown (2014), and following the ban (2015). To compare species-level 22 differences, in 2014 only, honeybee colonies in rural habitats were also sampled. Over half of all 23 samples were found to be contaminated (n=408), with thiamethoxam being the compound detected at 24 the highest concentrations in honeybee- (up to 2.29 ng/g in nectar in 2014, median<0.1 ng/g, n=79) 25 and bumblebee-collected pollen and nectar (up to 38.77 ng/g in pollen in 2013, median ≤ 0.12 ng/g, 26 n=76). Honeybees were exposed to higher concentrations of neonicotinoids than bumblebees in 2014. 27 While neonicotinoid exposure for rural bumblebees declined post-ban (2015), suggesting a positive 28 impact of the moratorium, the risk of neonicotinoid exposure for bumblebees in peri-urban habitats 29 remained largely the same between 2013 and 2015.

30

31 INTRODUCTION

Neonicotinoids are the most commonly used insecticides worldwide¹. Their systemic nature 32 33 means that, following seed-application to crops such as oilseeds or cereals, neonicotinoids become 34 incorporated into the tissues of a plant as it grows, including pollen and nectar, the main source of food for economically important pollinators, such as honeybees and bumblebees². Multiple studies 35 have raised concerns regarding the negative impacts of neonicotinoid exposure on bees³. Whitehorn et 36 al. $(2012)^4$ found that exposure of bumblebees to pollen and nectar containing 6 ng/g and 0.7 ng/g of 37 38 imidacloprid respectively, resulted in slower colony growth and the production of fewer new queens, 39 relative to unexposed colonies. Other studies have observed detrimental impacts on foraging and navigation^{5,6}, immunity⁷ and worker mortality⁸. Based on these findings, in 2013 the European 40 41 Commission instated a EU-wide moratorium on the use of three types of neonicotinoid, thiamethoxam, clothianidin and imidacloprid on bee-attractive flowering crops such as oilseed rape⁹. 42 In 2018, this ban was subsequently expanded to include all field crops $^{10-12}$. 43

44 Criticism has been levied against studies cited in support of the moratorium, mainly for using neonicotinoid concentrations purported to exceed those routinely experienced by foraging bees¹³, 45 sparking demand for further evidence as to what constitutes a 'field-realistic' dose. Several studies 46 have screened bee-collected pollen and nectar¹⁴⁻¹⁹ for neonicotinoid residues, quantifying the 47 48 'exposure landscape' by incorporating multiple chemicals from several forage sources. 49 Concentrations have been shown to vary considerably across studies, depending on location, time of 50 year and species. Pollen sampled from rural bumblebee colonies in Sussex, England, prior to the 51 implementation of the moratorium in 2013, was found to contain 18 ng/g of thiamethoxam on 52 average, with pollen collected from nests in nearby peri-urban areas containing up to 20 ng/g imidacloprid¹⁵ (mean=6.5 ng/g), well above the 6 ng/g used by Whitehorn *et al.*⁹. A large scale 53 Swedish field study found clothianidin concentrations averaging 5.4 ng/g in nectar sampled from 54 bumblebees foraging in fields of seed-treated oilseed rape (range 1.4-14 ng/g)¹⁶. In contrast, a study 55 conducted in Germany found considerably lower average concentrations (0.88 ng/g) in pollen 56 collected from bumblebee nests adjacent to neonicotinoids treated winter-sown oilseed rape²⁰, and a 57

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more recent study conducted across the UK, Hungary and Germany reported that concentrations detected in pollen and nectar collected by honeybees, bumblebees and the solitary bee *Osmia bicornis* rarely exceeded 1.5 ng/g^{21} . The wide ranging values reported by these studies highlights the need for further data to determine the actual exposure risk, particularly for wild bees.

62 Here we monitored bees' risk of neonicotinoid exposure during the period from pre- to post-63 moratorium, by screening pollen and nectar collected from bumblebee colonies located in several 64 regions; Sussex (2013-2015) and Hertfordshire (2014 only) in the south of England and Stirling, 65 Scotland (2013 only) in the north of the UK. Given the total weight of neonicotinoids applied in Scotland is much lower compared to the south of England (FERA PUS STATS database²²), we 66 67 expected the exposure risk to be lowest for bees in this region. There is currently limited data on the 68 exposure risk for wild bees from foraging on ornamental plants grown using neonicotinoids^{15,23,24} and 69 the use of neonicotinoid-based garden sprays, therefore we monitored bumblebees in both rural and 70 peri-urban habitats (Sussex and Stirling only), the latter consisting of domestic gardens located on the 71 outskirts of urban areas. For bees in rural areas, we expected neonicotinoid concentrations in pollen 72 and nectar collected in 2015 to be lower than those collected in 2013, before the implementation of 73 the moratorium. In 2014, the impact of the ban may not have fully come into effect, as any winter-74 sown oilseed crops would have been drilled prior to the implementation of the ban in December 2013 75 and therefore may still have been seed-treated with neonicotinoids. To compare species-level 76 differences in exposure risk during this transitional year (2014), we also screened pollen and nectar 77 from rural honeybee colonies located in Sussex and Hertfordshire.

78

79 MATERIALS AND METHODS

Site Information Bumblebee colonies (*B.terrestris audax*) were obtained from Agralan Ltd.,
Swindon, UK (originating from Biobest, Belgium) and in late spring (late May to early June, see
Table 1 for exact dates) were placed into the field:

i) to monitor exposure risk over the course of the implementation of the ban for both rural and
peri-urban habitats, bumblebee colonies were placed in rural (n=135, n=32-47/year) and peri-urban
(n=42, 12-15/year) locations across Sussex each year between 2013 and 2015. While the UK granted
a derogation to use neonicotinoids on oilseed rape in 2015, this was limited to a portion of East
England and did not affect the study area;

ii) to assess regional differences in neonicotinoid exposure between the north and south of the
UK, prior to the implementation of the ban (2013), bumblebee colonies were also placed in rural
(n=10) and peri-urban (n=20) locations in Stirling. In 2014 only, bumblebees were also placed in rural
locations across Hertfordshire (n=30) for comparison with Sussex colonies;

92 iii) to compare species-level differences in exposure risk, 15 rural bumblebee colonies were 93 each paired with a honeybee colony (located within 10m distance and placed into the field at the 94 beginning of April) in both Sussex and Hertfordshire in 2014 only. Queenright honeybee colonies 95 were obtained from experimental stocks at the University of Sussex and Rothamsted Research, which 96 at the beginning of the experiment consisted of a single brood box and a super containing frames of 97 fresh foundation wax, with additional space for bees to store pollen and nectar added as necessary. 98 We also mapped which crops were grown in ten, 5 km^2 surrounding the experimental colonies in 99 Sussex and Hertfordshire in 2014 (Fig. S4) and, where possible, asked farmers growing winter-sown 100 oilseed rape which seed treatments they had used (Table S4).

Sampling Pollen and nectar was collected from bumblebee colonies following four, eight and ten weeks of foraging in the field. Pollen was scraped out of the colony using a stainless steel microspoon, which was cleaned using methanol to avoid cross-contamination. From each colony, we aimed to collect enough pollen to fill a 1.5 ml micro-centrifuge tube, to ensure enough material for chemical analysis. Concurrently, 1.5 ml of nectar was obtained from nectar pots using disposable glass pipettes. However, care was taken not to completely deplete bumblebee colony stores. Where stores were low, no sample was collected (Table 2).

108 For honeybees, samples were collected once per month in April, May and June 2014, with the 109 last two sampling dates coinciding with sample collection from adjacent bumblebee colonies. Samples 110 were obtained from freshly drawn comb, where possible, to minimise contamination from previous 111 years. Enough pollen to fill a 1.5 ml micro-centrifuge tube was scraped out of ~ 10 cells using a 112 stainless steel micro-spoon as described above, and 1.5 ml of recently stored nectar was obtained from 113 uncapped and newly drawn comb using disposable glass pipettes. Freshness was determined by first 114 shaking the frame to ensure nectar dripped easily out of the comb. All pollen and nectar samples were 115 stored in individually labelled tubes and put on ice during transport back to the lab, and were then 116 frozen at -20°C until residue analysis was performed.

117 **Chemical analyses:** Pollen and nectar samples were extracted using the QuEChERS 118 method¹⁴ and screened for five neonicotinoids: thiamethoxam (TMX), clothianidin (CLO), 119 imidacloprid (IMC), acetamiprid (ACT) and thiacloprid (THC), using ultra high-performance liquid 120 chromatography tandem mass spectrometry (UHPLC-MS/MS). Pollen samples collected in Sussex in 121 2013 were not screened for acetamiprid.

122 **Sample preparation:** Pollen samples were extracted as described by Botias *et al.* $(2015)^{14}$. 123 Briefly, 100 mg of pollen was weighed into an Eppendorf tube and 400 pg of deuterated pesticides in 124 ACN were added. The extraction was performed by the addition of 400 μ l of water, 500 μ l of ACN, 125 125 mg of magnesium sulphate: sodium acetate mix (4:1) and 125 mg of PSA/C18/ENVI-Carb for the 126 dispersive solid phase extraction (dSPE) step (QuEChERS method). After the first extraction, the 127 aqueous phase and re-suspended pellet were extracted again with 400 µl of ACN and the supernatants 128 combined. Extracts were mixed with PSA/C18/ENVI-Carb and centrifuged. The supernatant was 129 evaporated to dryness under vacuum, reconstituted with 120 µl ACN:H₂O (10:90) and spin filtered 130 $(0.22 \ \mu m)$.

131 Nectar samples were centrifuged at 13,000 relative centrifugal force (RCF) for 10 min to 132 remove plant debris and the supernatant transferred into a clean eppendorf tube. Nectar samples were 133 very viscous and were therefore weighed for more accuracy (175 ± 50 mg depending on availability) 134 and the volume then increased to 400 µl with water. Four hundred pg of deuterated pesticide standard mixture was added to the nectar and the samples were extracted using the same QuEChERS methoddescribed for pollen.

137 **UHPLC-MS/MS analyses.** The ultra high-performance liquid chromatography tandem mass spectrometry (UHPLC-MS/MS) method described by Botias et al. (2015)¹⁴ was used for the analysis 138 139 of samples. UHPLC-MS/MS analyses were carried out using a Waters Acquity UHPLC system 140 coupled to a Quattro Premier triple quadrupole mass spectrometer from Micromass (Waters, 141 Manchester, UK). Data were acquired using MassLynx 4.1 and the quantification was carried out by 142 calculating the response factor of neonicotinoid compounds to their respective internal standards. 143 Concentrations were determined using a least-square linear regression analysis of the peak area ratio 144 versus the concentration ratio (native to deuterated). Method detection and quantification limits (MDL and MQL, respectively) as well as recoveries were determined as described by Botias et al. (2015)¹⁴ 145 146 (Table S1-3).

Quality control. One blank workup sample (*i.e.* solvent without matrix) per batch of eleven samples was included and injected on the UHPLC-MS/MS to ensure that no contamination occurred during the sample preparation. Solvent samples were also injected between sample batches to ensure that there was no carryover in the UHPLC system that might affect adjacent results in analytical runs. Samples were analysed in a random order and quality control samples (*i.e.* standards) were injected during runs every ten samples to check the sensitivity of the machine. Identities of detected neonicotinoids were confirmed by comparing ratio of MRM transitions in samples and pure standards.

154 Statistical Analysis. All analyses were performed using R-3.3.3. Residue concentrations that were 155 above the MDL but below the MQL were assigned the MDL (Tables 2-3, range 0.03-0.10 ng/g). Concentrations below the MDL were assumed to be zero¹⁴. Shapiro-Wilk tests, combined with 156 157 inspection of q-q plots, confirmed that residue data were not normally distributed. Therefore we 158 compared the frequency of neonicotinoid contamination using contingency tables and either χ^2 or 159 Fisher's exact tests (where expected frequencies were <5). To compare total neonicotinoid 160 concentrations between regions (Sussex vs. Stirling; Sussex vs. Herts), habitats (Rural vs. Peri-Urban) 161 and years of the study (2013 vs. 2015) we used non-parametric Mann-Whitney tests. For honeybee

data, where frequencies of contamination and residue concentrations were compared between samples from the same hive over several months, we used Cochran's Q test (with McNemar's test for post-hoc comparisons) and the Wilcoxon Signed-Rank test, with Bonferroni corrections to account for multiple comparisons. Given the relatively small number of pollen and nectar samples collected from each bumblebee colony, for analyses involving bumblebees we pooled samples collected after four and eight weeks in the field.

168 RESULTS

169 Bumblebees: In total, 233 pollen and nectar samples were collected from bumblebee colonies placed 170 in rural and peri-urban habitats in the regions of Stirling, Sussex and Hertfordshire between 2013 and 171 2015. Forty percent of all samples screened were found to be contaminated with neonicotinoids, 172 predominantly thiamethoxam (23%), thiacloprid (15%) and imidacloprid (10%). Pollen samples were 173 more often contaminated (62% samples) than nectar (25% samples) and the mean combined total 174 residues detected in pollen (Pollen N=132, 62% samples, mean \pm standard deviation (SD) =1.44 \pm 5.44 175 ng/g, median \leq MDL, max= 38.77 ng/g) were more than ten times higher (Nectar N=101, mean \pm SD= 176 0.12±0.44 ng/g, median <MDL, max=3.58 ng/g).

177 Differences in exposure by habitat and year: In 2013, the frequency of neonicotinoid contamination was similar for pollen (Table 1, $\chi^2_1=0$, p=1.000, Rural =58%; Peri-urban= 59%) and 178 nectar ($\chi^2_1=0$, p=1.000, Rural=14%, Peri-urban =14%) sampled from peri-urban (PU) and rural (R) 179 180 bumblebee colonies across the regions of Sussex (SU) and Stirling (ST) (Table 1). Concentrations of 181 neonicotinoids were very similar in nectar (Mann-Whitney $U_{21, 21}=225$, p=0.867, mean_{PU} ≤ 0.10 , 182 median_{PU} \leq 0.10, mean_R \pm SD=0.22 \pm 0.55 ng/g, median_R \leq MDL), and though higher in pollen from rural 183 colonies, this difference was not significant ($U_{36, 32}$ =603.5, p=0.73; mean_R=3.37±9.36 ng/g, 184 $median_R \le 0.12$, $mean_{PU} = 1.28 \pm 3.62$ ng/g, $median_{PU} \le 0.12$). While nectar from both habitats contained 185 only one type of neonicotinoid, predominantly thiamethoxam, over a quarter of pollen samples from 186 bumblebee colonies in rural (28%) and peri-urban (26%) habitats contained more than one residue. 187 Thiamethoxam (up to 38.77 ng/g, median < 0.12, mean \pm SD= $2.08\pm7.47 \text{ ng/g}$) and clothianidin (up to 188 2.08 ng/g, mean ≤ 0.12 ng/g, median < 0.12 ng/g) were present at the highest concentrations in rural

colonies. While thiamethoxam was also present in a high percentage of pollen samples collected from
peri-urban colonies in Sussex (79% samples), thiacloprid was found at the highest concentration in
these samples (up to 14.8 ng/g, mean ≤0.04 ng/g, median <0.04 ng/g).

In 2014, less than 10% of pollen (n=13) and nectar (n=13) samples from rural bumblebee colonies in Sussex contained neonicotinoids, all thiamethoxam and below the method quantification limit, whereas a significantly higher proportion of both pollen (85%, $\chi^2_1=8.987$, p=0.003, n=7) and nectar samples (80%, Nectar $\chi^2_1=6.152$, p=0.013, n=5) from peri-urban nests were contaminated (N=12), frequently with multiple residues (40% nectar samples, 29% of pollen). Again, thiacloprid (up to 9.32 ng/g in pollen, mean= 1.34 ± 3.52 ng/g, median ≤ 0.04 ng/g) and thiamethoxam (up to 3.48 ng/g in pollen, mean= 0.76 ± 1.52 , median=0.10 ng/g) and were detected at the highest concentrations.

199 In 2015, the frequency of neonicotinoid detection was similar for nectar collected from rural and peri-urban bumblebee colonies in Sussex ($\chi^2_1=0.158$, p=0.691, Rural=47%, Peri-urban=33%) as 200 201 were the concentrations present (Mann-Whitney $U_{19, 12}=130.5$, p=0.469, mean_R=0.10±0.15 ng/g, 202 median_R \leq MDL, mean_{PU}=0.08±0.17 ng/g, median_{PU} \leq MDL). While the frequency of detection 203 (Rural=35%, Peri-urban=64%), proportion of samples with multiple residues (Rural=9% vs. Peri-204 urban=18%) and mean concentration of neonicotinoids were higher in pollen from peri-urban nests, the difference was not significant (χ^2_1 =1.238, p=0.266, U_{22, 11}= 75.5, p=0.06, mean_R=0.06\pm0.14 ng/g, 205 206 median_R \leq MDL, mean_{PU}=1.29±3.30 ng/g, median_{PU} \leq MDL). Both habitats were contaminated predominantly with thiacloprid (up to 0.44 ng/g, mean±SD=0.04±0.11 ng/g, median <MDL), and 207 208 imidacloprid (up to 11.16 ng/g in peri-urban nests, mean \pm SD=0.21 \pm 1.40 ng/g, median <0.14), though 209 a small proportion of peri-urban samples also contained acetamiprid (4% up to 1.4 ng/g, mean ≤ 0.03 210 ng/g, median <MDL).

To compare the changing risk of exposure to peri-urban and rural bees over the transitional period from pre- to post- moratorium, we compared residue concentrations in 2013 and 2015 for Sussex bumblebee colonies only. For pollen collected from rural colonies there was a significant decrease in overall combined residue concentrations between years (Mann-Whitney $U_{23, 22}=385$, p=0.002, mean₂₀₁₃= 5.10±11.40 ng/g, median ≤ 0.12 ng/g, mean₂₀₁₅=0.06±0.14 ng/g, median \leq MDL), but not for 216 nectar ($U_{14, 19}=98$, p=0.134; mean₂₀₁₃= 0.20±0.51 ng/g, median <MDL, mean₂₀₁₅=0.10±0.15 ng/g, 217 median <MDL). When considering just those neonicotinoids affected by the moratorium 218 (thiamethoxam, clothianidin and imidacloprid), the same effect is observed, with a significant 219 decrease in residue concentrations in pollen ($U_{23, 22} = 389$, p <0.001, mean₂₀₁₃= 5.02±11.32 ng/g, 220 median ≤ 0.12 ng/g, mean₂₀₁₅=0.05±0.14 ng/g, median \leq MDL) but not nectar between 2013 and 2015 221 $(U_{14, 19} = 140, p=0.676; mean_{2013} = 0.20 \pm 0.51 \text{ ng/g}, median < MDL, mean_{2015} < MDL, median < MDL).$ 222 In contrast, concentrations of thiacloprid, which was unaffected by the ban, increased significantly in 223 nectar between 2013 and 2015 ($U_{14, 19}$ = 84, p= 0.013, mean₂₀₁₃ < MDL, median < MDL, 224 mean₂₀₁₅=0.09±0.15 ng/g, median <MDL). Concentrations of thiacloprid in pollen remained 225 unchanged over this period ($U_{23, 22}=267$, p=0.627, mean₂₀₁₃=0.08±0.31 ng/g, median <MDL, mean₂₀₁₅ 226 < MDL, median < MDL).

227 For peri-urban nests, there was no significant difference in overall residue concentrations in 228 either pollen ($U_{19,11}$ =124, p=0.408, mean₂₀₁₃= 2.11±4.56 ng/g, median=0.12 ng/g, mean₂₀₁₅=1.29±0.14 229 ng/g, median ≤ 0.04 ng/g) or nectar ($U_{13, 12}=62.5$, p=0.276, mean₂₀₁₃= 0.02 ± 0.05 ng/g, median \leq MDL, 230 mean₂₀₁₅=0.08±0.17 ng/g, median <MDL), samples collected between 2013 and 2015. When 231 considering either the banned neonicotinoids only (Pollen, $U_{19, 11}=134.5$, p=0.188; mean₂₀₁₃= 232 0.63 ± 1.64 ng/g, median ≤ 0.12 , mean₂₀₁₅= 1.14 ± 3.33 ng/g, median \leq MDL; Nectar $U_{13, 12}$ = 76, p= 0.898, 233 mean₂₀₁₃<MDL, median <MDL, mean₂₀₁₅<MDL, median <MDL) or thiacloprid, which was 234 unaffected by the ban (Pollen $U_{19, 11} = 104$, p= 1, mean₂₀₁₃= 1.47±4.41 ng/g, median <MDL, 235 mean₂₀₁₅<MDL, median <MDL, Nectar U_{13, 12}= 58.5, p= 0.067, mean₂₀₁₃<MDL, median <MDL, 236 $mean_{2015}=0.05\pm0.13$ ng/g, median <MDL), again there was no difference in the concentrations 237 detected in pollen and nectar collected from peri-urban nests between 2013 and 2015.

238 **Regional differences in exposure** In 2013, pollen collected from bumblebee colonies in Sussex (SU)

was more frequently contaminated (χ^2_1 =15.62, p<0.001, Sussex=79%; Stirling=27%), with

significantly higher concentrations of neonicotinoids than pollen collected from colonies in Stirling

- 241 (ST) (Mann-Whitney $U_{42,26}$ =276, p < 0.001; mean_{SU}±SD=3.74±9.01 ng/g, median_{SU}≤0.12 ng/g
- 242 mean_{ST} \pm SD =0.20 \pm 0.49 ng/g, median_{ST} <MDL). Nectar was contaminated at similar frequencies

(Fisher's Exact Test p=1.00, Sussex=14%; Stirling 12.5%) and concentrations ($U_{27,15}=200$, p=0.931;

244 mean_{SU}= 0.11 ± 0.37 ng/g, median_{SU} <MDL, mean_{ST}= 0.13 ± 0.47 ng/g, median_{ST} <MDL).

245 Pollen sampled from Sussex colonies was more frequently contaminated with multiple 246 residues (Peri-urban=37%, Rural=35%) compared to Stirling samples (Peri-urban=8%, Rural=15%), 247 and the concentrations of thiamethoxam detected in pollen were considerably higher 248 $(\text{mean}_{SU}=0.58\pm1.64 \text{ ng/g}, \text{median}=0.12 \text{ ng/g} vs. \text{mean}_{ST} \le 0.12 \text{ ng/g}, \text{median} \le 0.12 \text{ ng/g})$. Sussex peri-249 urban colonies in particular also contained higher concentrations of thiacloprid compared to Stirling 250 $(\text{mean}_{SU} = 1.47 \pm 4.41 \text{ ng/g median} < 0.03 \text{ ng/g vs. } \text{mean}_{ST} = 0.07 \pm 0.22 \text{ ng/g}, \text{ median} < 0.03 \text{ ng/g}).$ 251 Imidacloprid was also frequently detected in pollen from Sussex nests in 2013, but was not detected in 252 any samples from Stirling. Clothianidin was not detected in any Sussex nests, but accounted for the 253 highest residue concentrations detected in nests in Stirling (mean_{ST}= 0.16 ± 0.58 g/g, median <MDL, 254 $max_{ST} = 2.08 \text{ ng/g}$).

255 In 2014, residues detected in pollen and nectar samples collected from bumblebee colonies 256 placed in rural habitats in Hertfordshire (H) and Sussex (SU) were all below the limits of 257 quantification (<0.04-0.1 ng/g). Though there was a higher frequency of contamination of both pollen 258 (H=36%, SU=7%) and nectar (H=20%, SU= 8%) from Hertfordshire colonies, this difference was not 259 significant (Nectar: Fisher's Exact Test p=0.560; N_{SU}=13, N_H=10; Pollen p=0.142, N_{SU}=13, N_H=11). 260 A small proportion of pollen from Sussex (10%), and nectar from both regions was contaminated with 261 thiamethoxam (SU=10%; H=20%). Pollen from Hertfordshire colonies also contained acetamiprid 262 (10%) and, more frequently, this cloprid (40%).

Honeybees: In total, 175 pollen and nectar samples were collected from honeybee hives in Sussex and Hertfordshire between April and June May 2014, with over two thirds (68%) found to be contaminated with neonicotinoids. Total residue concentrations in nectar (N= 85, mean \pm SD = 0.64 \pm 0.84 ng/g, median=0.20 ng/g, max= 4.23 ng/g) were approximately three times the concentrations detected in pollen (N= 90, mean \pm SD = 0.20 \pm 0.32 ng/g, median \leq 0.12 ng/g, max=1.74 ng/g), with 40% of nectar samples containing more than one residue, compared to just 9% of pollen samples. Alongside thiamethoxam, which was highly prevalent in both pollen (61% of samples) and nectar

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(69%), clothianidin was also frequently detected in nectar collected from honeybee hives (40%), but
only once in pollen (Table 2). Imidacloprid and thiacloprid were detected in a very small percentage
of samples (4-5%) and acetamipirid was not detected.

273 **Seasonal differences:** Frequency of neonicotinoid detection in pollen (Cochran's *Q*=24.67, 274 df=2, p<0.001) and nectar (Q=20.38, df=2, p<0.001) sampled from honeybee colonies in 2014 275 changed significantly across the season. The highest frequency and concentration of neonicotinoid 276 residues were detected in April (Fig. 3), when nearly all nectar samples collected from hives in 277 Hertfordshire (H) and Sussex (SU) were contaminated with neonicotinoids (H=100%, mean_H \pm SD 278 $=1.46\pm0.66$ ng/g; median=1.17 ng/g; SU=93%, mean_{SU}=0.95 ±1.13 ng/g, median ≤0.12 ng/g). 279 Likewise, almost all pollen samples contained neonicotinoid residues (H=80%, mean_H=0.41±0.47 280 ng/g, median $\leq 0.12 ng/g$; SU=100%, mean_{SU}=0.23 $\pm 0.19 ng/g$, median $\leq 0.12 ng/g$) in April.

Between April and May, there was a similar frequency of neonicotinoid detection in both pollen (April= 90%, May=73%, McNemar test, p=0.287) and nectar (April=81%, May=80% p=0.760). While the concentration of neonicotinoid residues in pollen remained the same as the previous month (Wilcoxon signed-rank test, $Z_{30}=0.28$, p=0.120, mean_{April}= 0.32 ± 0.37 ng/g, median ≤ 0.12 ng/g mean_{May}= 0.22 ± 0.33 , median ≤ 0.12 ng/g), neonicotinoid concentrations in nectar, previously high in comparison to pollen, declined significantly between April and May ($Z_{26}=0.75$, p<0.001; mean_{April}= 1.20 ± 0.95 ng/g, median=1.06 ng/g, mean_{May}= 0.65 ± 0.72 , median=0.27 ng/g).

At the final sampling point in June, neonicotinoid concentrations detected in samples from both regions were below the limit of quantification, and were significantly lower than in May (Pollen $Z_{30}=0.55$, p=0.003; Nectar $Z_{27}=0.73$, p<0.001). The frequency of neonicotinoid detection in both pollen (30% samples, McNemar test, p=0.002) and nectar (34% samples, p=0.002) was also significantly lower than the previous month (Table 2)

Regional differences: While overall neonicotinoid concentrations in pollen contamination did not differ between Hertfordshire and Sussex (Mann-Whitney $U_{45, 45}=1014$, p=0.100, mean_H=0.23±0.36, median ≤0.12 ng/g, mean_{SU}= 0.17±0.27, median ≤0.12 ng/g), concentrations in nectar were significantly higher in Hertfordshire hives ($U_{44, 42}=1301$, $p \le 0.001$, mean_H=0.88±0.81, median=0.75 ng/g, mean_{SU}=0.40±0.80 ng/g, median ≤ 0.10 ng/g). Crop mapping of the five 5 km² study areas in each region in 2014, showed that arable crops accounted for 55% of land cover in Hertfordshire (9% oilseed rape), and 32% in Sussex (5% oilseed rape, Figure S4).

Species-specific differences: A comparison of residue concentrations in pollen and nectar collected from adjacent honeybee (HB) and bumblebee (BB) nests located in rural habitats in Hertfordshire and Sussex revealed significantly higher concentrations of neonicotinoid exposure for honeybees compared to bumblebees (Table 1, 2, $U_{18, 18}$ = 112, p=0.04; mean_{HB}=0.17±0.39 ng/g, median <MDL, max=1.38 ng/g; mean_{BB}≤0.12 ng/g, median <MDL, max ≤0.12 ng/g).

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306 **DISCUSSION**

307 In December 2013, an EU-wide moratorium on the use of certain neonicotinoids on bee-attractive 308 flowering crops was implemented by the European Commission, which in early 2018 was 309 subsequently expanded to include all field crops. To monitor bees' exposure to neonicotinoids during 310 the intital transitional period from pre- to post-ban, between 2013 and 2015 we collected more than 311 400 pollen and nectar samples from bumblebee and honeybee colonies located in rural and peri-urban 312 habitats in three regions across the UK, finding just over half of all samples to be contaminated with 313 neonicotinoids. While combined total concentrations of neonicotinoids in pollen collected by rural 314 bumblebees declined post-ban from an average of 5.1 ng/g in 2013, to 0.06 ng/g in 2015, suggesting a 315 positive impact of the moratorium, neonicotinoid concentrations detected in samples collected from 316 peri-urban bumblebee colonies remained largely unchanged between 2013 and 2015, indicating that 317 the risk of exposure for peri-urban bees was not altered during the transitional period, and that more 318 could be done to mitigate the risk for bees foraging in such habitats.

Across all samples, the highest neonicotinoid residue concentrations were detected in 2013, in pollen samples collected from rural bumblebee colonies in Sussex. Concentrations of up to 38.77 ng/g of thiamethoxam were detected, with the average total neonicotinoid concentrations of 5.1 ng/g

similar to that detected by previous studies conducted prior to the moratorium^{25,15,26}, and within the 322 range demonstrated to have negative impacts on bumblebee physiology^{27,28}, foraging efficiency²⁹ and 323 324 colony growth²⁸. Pre-ban (2013), the frequency of neonicotinoid contamination was extremely high for pollen sampled from bumblebee colonies in both rural and peri-urban habitats in Sussex (74% and 325 326 84% of pollen samples respectively, mean=3.74 ng/g). As predicted, pollen samples collected from 327 nests near Stirling in 2013 were contaminated to a lesser degree (23-30% of pollen samples), and with 328 lower concentrations (mean=0.20 ng/g). This likely reflects the fact that across Scotland, 329 neonicotinoid use in 2013/2014 was approximately four times lower than in South East England (4, 186 kg, over 78, 345 ha vs. 16, 820 kg, over 197,507 ha²²), though differences in the growth season 330 331 and therefore timing of neonicotinoid application between regions may also have played a role.

332 Pollen and nectar samples collected from honeybee colonies in 2014, post-implementation of 333 the ban, but when any winter-sown oilseed rape may still have been seed-treated with neonicotinoids. 334 also had a high prevalence of neonicotinoid contamination (68% samples). Contamination was highest 335 in April when oilseed rape was flowering (93% samples), and declined throughout the season, a phenomenon observed in several earlier studies^{14,15,23,30}, and hypothesised to arise from temperature 336 337 increases and photo-degradation of neonicotinoid residues in plant tissues as the season progresses³¹. 338 During this early part of the year, concentrations detected in honeybee-collected nectar averaged 1.2 339 ng/g, close to the average maximum concentration detected in seed-treated crop nectar, as reported by Godfray et al.³² (1.9 ng/g, averaged from 20 published studies). Concentrations in pollen were 340 341 considerably lower (0.32 ng/g, average maximum concentration in seed-treated crop pollen=6.1 ng/g³²). likely reflecting honeybees' preference for collecting nectar from oilseed rape. For both 342 343 bumblebees and honeybees, early spring is a period when the colony might be expected to be 344 particularly vulnerable^{33,34}, and levels detected in pollen were within the range known to impair honeybee foraging performance³⁵, immune function⁷ and alter gene expression pathways³⁶. 345 Furthermore, as observed in several previous studies^{15,17,18}, many of the samples we screened were 346 347 found to contain more than one neonicotinoid residue, which gives rise to the potential for additive or synergistic effects. Tosi et al.¹⁷ found when screening honeybee pollen collected from multiple 348

apiaries across Italy for 66 different pesticides, that the frequency of detection actually peaked in summer months. Though here we did not screen for the presence of other chemical classes such as fungicides, there is evidence to suggest that exposure to certain fungicides can make bees more susceptible to the adverse effects of neonicotinoids³⁷.

353 Although the concentration of neonicotinoids in pollen and nectar sampled from rural 354 bumblebee colonies declined between 2013 and 2015, bumblebees from both rural and peri-urban 355 habitats were nevertheless still exposed to neonicotinoids following the implementation of the ban. 356 Indeed 47% of nectar and 36% of pollen samples collected from rural colonies in 2015 contained 357 neonicotinoid residues, a similar frequency as observed for peri-urban nests (33% nectar, 64%) 358 pollen), albeit at lower concentrations (mean concentration detected in pollen from rural nests = 0.06359 ng/g vs. 1.29 ng/g detected in peri-urban pollen in 2015). This echoes the findings of Woodcock et $al.^{30}$ who screened honey samples submitted by beekeepers across the UK, and found that while 360 361 samples harvested in 2014 were more likely to be contaminated (52% samples), 22.9% of samples 362 harvested post-ban in 2015 also contained neonicotinoids. Similarly, a worldwide study of honey 363 contamination spanning five years between 2012 and 2016, found 75% of 198 samples to contain neonicotinoids, with the highest prevalence in honey from North America, Asia and Europe³⁸. 364

365 Not only did exposure to neonicotinoids change for rural bees between 2013 and 2015, so did 366 the chemical type. Across all samples, thiamethoxam was the most frequently detected, which is 367 unsurprising given that, prior to the moratorium, it was the active ingredient in the mostly commonly 368 used seed dressing on oilseed rape across Great Britain. Indeed, of fifteen farmers growing winter-369 sown oilseed rape within a 5 km radius of our experimental bee colonies that we interviewed in 2014, 370 twelve had used seeds dressed with a thiamethoxam-based formulation (Cruiser®). Clothianidin, a 371 metabolite of thiamethoxam and still in use as a seed-dressing on non-flowering cereal crops, was also 372 frequently detected in honeybee nectar (69% samples), but only once in pollen, and was rarely 373 detected in any samples collected from bumblebee colonies. Post-ban, acetamiprid and thiacloprid, 374 the use of which is unaffected by the moratorium, were detected more often and at higher levels than 375 thiamethoxam. For nectar samples collected from rural bumblebee colonies, thiacloprid

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concentrations actually significantly increased between 2013 and 2015. Thiacloprid is an active
ingredient in many bug sprays sold in garden centres, and a recent study in which ornamental 'beefriendly' plants were screened for multiple pesticide and fungicide residues found more than 70% of
plants contained neonicotinoids, with thiacloprid present in almost half²⁴.

380 Imidacloprid was detected in a moderate proportion (10%) of samples collected from 381 bumblebee nests throughout the duration of the study. Considering that use of imidacloprid in arable 382 farming has dramatically declined in the UK (50% and 90% decline in weight of imidacloprid applied 383 to cereals and oilseeds respectively between 2012 and 2014, PUS Stats database, Table S6), having 384 been replaced by thiamethoxam and clothianidin, it is somewhat concerning that it was detected to such an extent. Woodcock et al.³⁰ also noted that imidacloprid was present in honey samples 385 harvested in 2014 at a rate 'disproportional to its use' and Tosi et al.¹⁷ detected imidacloprid in 9.1% 386 387 of honeybee-collected pollen sampled from multiple apiaries across Italy in 2014 at mean 388 concentrations of 2 ng/g, raising concerns about the persistence of this chemical in agroenvironments. As previously observed when screening pollen from bumblebee colonies¹⁵ and wild 389 bumblebees collected in peri-urban areas²³, the highest concentrations of imidacloprid were detected 390 391 in peri-urban colonies, at levels up to 11.16 ng/g in 2015 (mean=1.13 ng/g). Again, this may originate 392 from use by the horticulture industry, since screening of ornamental plants detected imidacloprid in 38% of samples²⁴. An alternative, yet untested source, is the use of imidacloprid for flea control in 393 394 domestic pets and as ant poison.

Honeybees in Hertfordshire were exposed to significantly higher neonicotinoid concentrations in nectar compared to Sussex honeybees, which is most likely explained by the fact that, in 2014, there was almost double the percentage cover of treated oilseed crops (9% land cover in Hertfordshire *vs.* 5% in Sussex), and generally a higher percentage of arable land cover (55%) compared to Sussex (32%).

400 Overall, honeybee samples had higher concentrations of neonicotinoids compared to 401 bumblebees. This contrasts with findings from an earlier study conducted in 2013 where the reverse 402 was found to be true¹⁵. However in the previous study, colonies of each species were not placed in identical locations, therefore in addition to differences in foraging range and flower preferences^{39,40},
colonies may simply have been in proximity to a different range of plant species. Clearly more paired
sampling of both species is required to establish whether there are consistent differences in exposure.

406 On the basis of evidence published post-2013, the European Food Standards Agency recently concluded that neonicotinoids do indeed pose a risk to bees⁴¹, and in 2017 the EU commission 407 408 proposed extending the moratorium to include all field crops (barring permanent greenhouse crops), which was passed by the European Union in early 2018^{10–12}. Here we have shown for the first time 409 410 how exposure to neonicotinoids has changed for bees foraging in rural and peri-urban areas across the 411 UK, since the initial implementation of the moratorium on their usage in December 2013. The 412 exposure of rural bumblebees appears to have declined post-ban, suggesting that continued limitation 413 of their use on flowering crops could have a positive impact on the risk for bees and other pollinators 414 in rural areas. However, exposure for peri-urban bees remains largely unaffected, presumably as a 415 result of contaminated ornamental plants sold in garden centres and ongoing domestic usage of 416 neonicotinoid-based bug sprays. This is concerning given the growing interest in encouraging 417 pollinators in urban areas; more research is needed to understand the sources of exposure and find 418 ways to reduce it.

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Moratorium Status	Voor	Pagion	Pag Spacios	Habitat	N	Sampling Dates				
	fear	Region	bee species	Habitat	Colonies	Sampling Dates				
Pre-ban	2013	Stirling	Bumblebee	Rural	10	12 th June; 11 th July; 18 th July				
				Peri-urban	20	6 th June; 4 th July; 17 th July				
		Sussex	Bumblebee	Rural	32	30 th May; 9 th June; 23 rd June				
				Peri-urban	12	30 th May; 9 th June; 23 rd June				
During ban	2014	Sussex	Bumblebee	Rural	47	28 th May; 25 th June; 9 th July				
(Winter-sown crops				Peri-urban	15	28 th May; 25 th June; 9 th July				
still seed-treated)			Honeybee	Rural	15	16 th April; 28 th May; 25 th June				
		Herts	Honeybee	Rural	15	16 th April; 28 th May; 25 th June				
			Bumblebee	Rural	30	28 th May; 25 th June; 9 th July				
During ban	2015	Sussex	Bumblebee	Rural	45	15 th June; 13 th July; 27 th July				
				Peri-urban	15	15 th June; 13 th July; 27 th July				

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463 Table 1 Number of honeybee and bumblebee colonies placed in each habitat type (Peri-urban vs.

464 Rural), in each region (Sussex, Stirling, Hertfordshire (Herts)) across the three years of the study

465 (2013-2015). The specific dates colonies were sampled for pollen and nectar are listed.

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						-				NECTA	٨R							POLLEN			
468				Me	thod Quantifica	tion Limit (ng/g)	0.3	0.3	0.4	0.08	0.08			MQL	0.36	0.36	0.48	0.12	0.12		
					Method Deteo	tion Limit (ng/g)	0.1	0.1	0.14	0.03	0.03			MDL	0.12	0.12	0.16	0.04	0.04		
160 Ye	ear	Region	Location	N Colonies	N Samples	ng/g	тмх	CLO	IMC	ACT	THC	TOTAL	% Multi-residue	N	TMX	CLO	IMC	ACT	THC	TOTAL	% Multi-residue
405						Frequency %	12.5%					12.5%	0%		7.7%	7.7%			15.4%	23.1%	8.3%
470			Peri-Urhan	20	8	Mean ±SD	≤0.10					≤0.10		13	≤0.12	≤0.12			0.06±0.22	0.08±0.21	
470				20	0	Median	≤0.10					≤0.10		15	≤0.12	≤0.12			≤0.04	≤0.12	
		STIRLING				Max	≤0.10					≤0.10			≤0.12	≤0.12			0.76	0.76	
4/1						Frequency %	12.5%					12.5%	0%		7.7%	7.7%			30.8%	30.8%	15.3%
			Rural	10	7	Mean ±SD	0.26± 0.68					0.26±0.68		13	≤0.12	0.16±0.58			0.15±0.36	0.32±0.0.6	5
472	-		narai	10		Median	≤0.12					≤0.10		15	≤0.12	≤0.10			≤0.03	≤0.12	
<u> </u>	ב בי					Max	1.81					1.81			≤0.12	2.08			1.15	2.08	
473 č	5					Frequency %	7.7%		7.7%			15.4%	0%		79%	5.26%	26.3%	nt	15.8%	84.2%	36.8%
•	N		Peri-Urban	12	13	Mean ±SD	≤0.10		≤0.14			≤0.10		10	0.58±1.64	≤0.10	≤0.16		1.47±4.41	2.11±4.56	
474					10	Median	≤0.10		≤0.14			≤0.10		15	≤0.12	≤0.10	≤0.16		≤0.04	≤0.12	
		SUSSEX				Max	≤0.10		≤0.14			≤0.14			7.1	≤0.10	≤0.16		14.68	14.8	
475		00002/1				Frequency %	14.3%					14.3%	0%		60.9%	4.35%	39.1%	nt	17.4%	74%	34.8%
.,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,		Rural	32	14	Mean ±SD	0.2±0.51					0.20±0.51		23	4.96± 11.29	≤0.12	≤0.16		0.08±0.31	5.10±11.41	L	
176						Median	≤0.10					≤0.10		20	≤0.12	≤0.12	≤0.16		≤0.04	≤0.12	
470						Max	1.49					1.49			38.77	≤0.12	≤0.16		1.5	38.93	
177						Frequency %	80.0%	40.0%				80.0%	40%	40% 7	57.1%		14.3%	14.3%	42.9%	85.7%	28.6%
477				15	5	Mean ±SD	0.76±1.52	≤0.10				0.80±1.56			≤0.12		0.31±0.82	≤0.04	1.34±3.52	1.73±3.43	
470			Peri-Urban	10		Median	0.10	≤0.10				0.1			≤0.12		≤0.16	≤0.04	≤0.04	≤0.16	
478						Max	3.48	≤0.10				3.58			≤0.12		2.18	≤0.04	9.32	9.32	
		SUSSEX	Rural			Frequency %	8.3%					8.3%	0%		7.7%					7.7%	0.0%
479	Ţ			47	13	Mean ±SD	≤0.10					≤0.10		13	≤0.12					≤0.12	
è	.					Median ≤0.10 ≤0.10	≤0.12					≤0.12	12								
480 r	N _					Max	≤0.10					≤0.10			≤0.12					≤0.12	
						Frequency %	20%					20%	0%					9.1%	36.4%	36.4%	9.1%
481		HERTS	Rural	30	10	Mean ±SD	≤0.10					≤0.10		11				≤0.04	≤0.04	≤0.04	
			narai			Median	≤0.10					≤0.10						≤0.04	≤0.04	≤0.04	
482—						Max	≤0.10					≤0.10						≤0.04	≤0.04	≤0.04	
						Frequency %			16.7%		25%	33.3%	8.3%		9.1%		27.3%	36.35	18.2%	63.6%	18.2%
483			Peri-Urban	15	12	Mean ±SD			≤0.14		0.05±0.13	3 0.08±0.17		11	≤0.12		1.13±3.34	0.14±0.42	2 ≤0.04	1.29±3.30	
.00	•					Median			≤0.14		≤0.03	≤0.10			≤0.12		≤0.16	≤0.04	≤0.04	≤0.04	
/8/		SUSSEX				Max			≤0.14		0.44	0.44			≤0.12		11.16	1.40	≤0.04	11.16	
Ċ	S.					Frequency %	5.3%		5.3%		36.8%	47.4%	0%		13.6%		9.1%	9.1%	13.6%	36.4%	9.1%
195 (N		Rural	45	19	Mean ±SD	≤0.10		≤0.14		0.09±0.15	5 0.10±0.15		22	≤0.12		≤0.16	≤0.04	≤0.04	0.06±0.14	
400					15	Median	≤0.10		≤0.14		≤0.03	≤0.10			≤0.12		≤0.16	≤0.04	≤0.04	≤0.12	
						Max	≤0.10		≤0.14		0.42	0.42			≤0.12		0.60	≤0.04	≤0.04	0.60	

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487Table 2 Frequency of detection (% samples), mean (± standard deviation (SD)), median and maximum concentrations of five neonicotinoids (TMX=thiamethoxam,
CLO= clothianidin, IMC= imidacloprid, ACT=acetamiprid, THC=thiacloprid) and the combined total concentration of neonicotinoids detected in pollen and nectar

⁴⁸⁸ sampled from bumblebee colonies located in rural and peri-urban habitats in three different regions; Stirling, Hertfordshire (Herts) and Sussex. Samples were collected across three years (2013-2015). Multi-residue samples are those where more than one type of neonicotinoid was detected. MQL= Method quantification limit, MDL=Method detection limit, nt= not tested, \leq less than or equal to.

489								NECTAR								POLLEN																
				Method Quantification Limit (ng/g)	0.3	0.3	0.4	0.08	0.08			MQI	0.36	0.36	0.48	0.12	0.12															
490				Method Detection Limit (ng/g)	0.1	0.1	0.14	0.03	0.03			MDL	0.12	0.12	0.16	0.04	0.04															
	Month	Region	Ν		тмх	CLO	IMC	ACT	THC	TOTAL	% Multi-residue	Ν	тмх	CLO	IMC	ACT	THC	TOTAL	% Multi-residue													
491				Frequency of detection %	100%	73.3%	6.7%			100%	80.0%		80%		6.6%		13.3%	80%	20.0%													
		HERTS	15	Mean ± SD (ng/g)	0.83 ± 0.48	0.63 ± 0.51	≤0.14			1.46±0.66		15	0.26±0.28		≤0.16		0.14±0.42	0.41±0.47														
492			15	Median (ng/g)	0.77	0.66	≤0.14			1.17			0.12		≤0.16		≤0.04	0.12														
102	R			Max (ng/g)	1.83	1.38	≤0.14			1.83			0.94		≤0.16		1.62	1.62														
495	ΑP			Frequency of detection %	93%	47%	7%		7%	93.3%	60.0%		100%					100%	0%													
494	-	SUSSEX	15	Mean ± SD (ng/g)	0.56 ± 0.14	0.37±0.18	≤0.14		≤0.03	0.95 ±1.13		15	0.23±0.19					0.23±0.19														
		SUSSEX	15	Median (ng/g)	0	≤0.1	≤0.14		≤0.03	0.58			0.12					0.12														
495				Max (ng/g)	1.76	2.47	≤0.03		≤0.03	2.47			0.6					0.60														
				Frequency of detection %	86.6%	73.3%				93.3%	66.7%		80%					80%	0%													
496			15	Mean ± SD (ng/g)	0.60±0.16	0.38±0.11				1.04±0.74		15	0.19±0.24					0.19±0.24														
407		HERTS	15	Median (ng/g)	0.45	0.10				1.08			0.12					0.12														
497	Ā			Max (ng/g)	2.29	1.26				2.29			0.92					0.92														
108	Σ		12	Frequency of detection %	66.7%	16.7%			16.70%	66.7%	25.0%		53.3%	6.7%	6.7%		20%	66.7%	20%													
490		SUSSEY		12	12	Mean ± SD (ng/g)	0.12±0.05	≤0.10			≤0.03	0.19±0.34		15	≤0.12	≤0.12	≤0.16		0.16±0.4	0.24±0.4												
499		JUJJLA				12	12	12	12	12	12	12	12	12	12	12	12	12	12	Median (ng/g)	0.10	≤0.10			≤0.03	0.10			≤0.12	≤0.12	≤0.16	
				Max (ng/g)	0.53	0.68			≤0.03	0.68			≤0.12	≤0.12	≤0.16		1.19	1.2														
500				Frequency of detection %	50%	21.4%	7.1%			66.3%	21.4%		26.7%		6.7%			26.7%	8.9%													
		LEPTS	14	Mean ± SD (ng/g)	≤0.10	≤0.10	≤0.14			0.08±0.08		15	≤0.12		≤0.16			0.09±0.26														
501		HENIS	14	Median (ng/g)	≤0.10	≤0.10	≤0.14			0.10			≤0.12		≤0.16			≤0.12														
- 02	Ë			Max (ng/g)	≤0.10	≤0.10	≤0.14			≤0.14			≤0.12		0.88			0.88														
502	S			Frequency of detection %	13.3%					13.3%	0%		26.7%		6.7%		6.7%	33.3%	6.7%													
502	-	SUSSEY	15	Mean ± SD (ng/g)	≤0.10					≤0.10		15	≤0.12		≤0.16		≤0.04	0.05±0.07														
202		JUSSEX	EX 15	Median (ng/g)	≤0.10					≤0.10			≤0.12		≤0.16		≤0.04	≤0.12														
504				Max (ng/g)	≤0.10					≤0.10			≤0.12		≤0.16		≤0.04	≤0.16														

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506 Table 3 Frequency of detection (% samples), mean (± standard deviation (SD)), median and maximum concentrations of five neonicotinoids

507 (TMX=thiamethoxam, CLO= clothianidin, IMC= imidacloprid, ACT=acetamiprid, THC=thiacloprid) and the combined total concentration of neonicotinoids 508 detected in honeybee nectar and pollen sampled from colonies located in in Sussex (N=15) and Hertfordshire (Herts, N=15) between April and June. Multi-

detected in honeybee nectar and pollen sampled from colonies located in in Sussex (N=15) and Hertfordshire (Herts, N=15) between April and June. Multiresidue samples are those where more than one type of neonicotinoid was detected. MQL= Method quantification limit, MDL=Method detection limit, nt= not

510 tested, \leq less than or equal to.



Figure 2 Total neonicotinoid concentrations (Thiamethoxam, clothianidin, imidacloprid, acetamiprid and thiacloprid combined) detected in A) Pollen and B) Nectar samples collected from bumblebee colonies in Rural (White, N Pollen samples=45; Nectar=33) and Peri-urban (Grey, N Pollen samples= 30; Nectar=25) habitats across the region of Sussex in the years 2013 and 2015. Concentrations are plotted on a square-root scale. Black horizontal bars show median values. Box limits denote the first and third quantiles, and boxplot whiskers extend to 1.5 times the interquartile range. Outliers are represented by solid black circles.

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541 ASSOCIATED CONTENT

542 Supporting Information

- 543 The following file is available free of charge.
- Additional figures and tables as mentioned in the text (PDF)

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