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Sanctis A–C: Three Racemic Procyanidin Analogues From The Lichen *Parmotrema sancti-angelii*

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Abstract: The phytochemical investigation of the lichen *Parmotrema sancti-angelii* afforded three racemic compounds featuring an original dibenzo-2,8-dioxabicyclo[3.3.1]nonane scaffold, sanctis A–C. These compounds were structurally characterized by extensive NMR analyses, comparison between experimental and theoretical NMR data and X-ray crystallography. These procyanidin A-like metabolites display a methyl group instead of a pendant aromatic ring at C-9, a so far unprecedented structural feature. A biosynthetic route to sanctis A–C is proposed herein.

In the course of our systematic research on lichen substances from Viet Nam, we have examined *Parmotrema sancti-angelii* (Lyngé) Hale, a widely distributed species in the southern part of central Viet Nam. Previous studies on the chemical constituents of *P. sancti-angelii* reported on the presence of two depsides, atranorin and lecanoric acid, and one depsidone, α -collatolic acid.^[2] The current phytochemical investigation of *P. sancti-angelii* led to the isolation of three racemic pairs of new metabolites bearing an unprecedented carbon skeleton comprising a dibenzo-2,8-dioxabicyclo[3.3.1]nonane core, namely sanctis A–C (**1–3**) (Figure 1), for which a putative biosynthetic scheme is proposed herein alongside eight known metabolites, the structures of which are collated in Figure S1 (Supporting Information).

Introduction

Lichens are symbiotic associations comprised of a fungus and a photobiont partner (green alga and/or cyanobacteria). This original consortium produces unique secondary metabolites endowed with various biological activities, paving the way for the phytochemical investigation of these fascinating organisms.^[1]

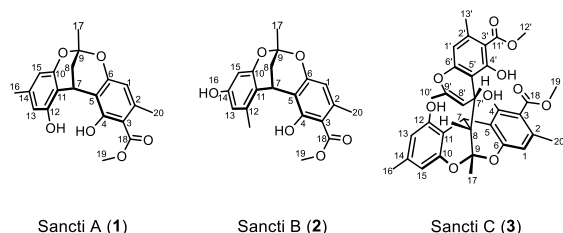


Figure 1. Structures of sanctis A–C (**1–3**). Only one enantiomer is represented per racemic pair.

Results and Discussion

Compound **1** was obtained as a colorless amorphous solid. The molecular formula was deduced to be $C_{20}H_{20}O_6$ by HRESIMS at m/z 379.1153 $[M+Na]^+$ (calcd. 379.1152 for $C_{20}H_{20}O_6Na$) (11 units of unsaturation). 1H NMR and HSQC spectra of **1** showed a hydrogen-bonded hydroxyl group (δ_H 13.03), one phenolic hydroxyl group (δ_H 7.58), three aromatic protons (δ_H 6.37, 6.34 and 6.31), one methoxy group (δ_H 3.93), one deshielded methine proton (δ_H 4.47, 1H, t, $J = 3.0$ Hz), one diastereotopic methylene group (δ_H 2.11, 1H, dd, $J = 13.0, 3.0$ Hz and δ_H 2.16, 1H, dd, $J = 13.0, 3.0$ Hz) and three methyl groups (δ_H 2.43, 2.19 and 1.85, each 3H, s) (Table 1). The ^{13}C NMR spectrum revealed twenty carbons including one methyl ester group [δ_C 172.8 (-CO), 52.3 (-OCH₃)], three aromatic methine carbons (δ_C 112.8, 110.9 and 109.4), four oxygenated aromatic quaternary carbons (δ_C 159.1, 156.9, 153.3 and 152.1), six quaternary carbons (δ_C 141.3, 138.5, 112.1, 110.9, 105.4 and 98.2), one upfield shifted methine carbon (δ_C 21.4), one methylene carbon (δ_C 30.9), and three methyl groups (δ_C 27.3, 24.0 and 20.4). The HMBC spectrum showed the correlations of the chelated hydroxyl group (δ_H 13.03, 4-OH) to signals at δ_C 105.4 (C-3), 112.1 (C-5), and 159.1 (C-4). A first methyl group (δ_H 2.43, H₃-20) correlated to signals at δ_C 105.4 (C-3), 112.8 (C-1), 141.3 (C-2) and also showed a 4J coupling to a signal at δ_C 172.8 (C-18). A first aromatic proton (δ_H 6.34, H-1) correlated to this methyl carbon (δ_C 24.0), as well as to signals at δ_C 105.4 (C-3) and 156.9 (C-6). HMBC cross peaks revealed the correlations of two aromatic methine protons at δ_H 6.37 (H-13) and 6.34 (H-15) to a second methyl group at δ_C 20.4 (C-16). This methyl group also correlated to signals at δ_C 138.5 (C-14), 110.9 (C-13) and 109.4 (C-15). The H-13 proton correlated to a signal at δ_C 153.3 (C-12) and the H-15 proton revealed cross peaks to signals at δ_C 152.1 (C-10) and 110.9 (C-13). HMBC connectivities comprised H-7 methine that resonated at δ_H 4.47 and δ_C 21.4. Although H-7 displays a low chemical shift, the high field shift of the corresponding carbon indicates that it might be directly

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connected to carbon atoms in both aromatic rings. Likewise, the low field shift of H-7 is consistent with its being constrained within the deshielding shift of two aromatic systems.^[3,4] Its COSY correlation with a methylene group (H₂-8, δ_{H} 2.11 and 2.16) indicated that these groups were adjacent to one another, which was further supported by the multiplicity of the methine signal. The AB system of this methylene group implied that the mentioned methine was an asymmetric carbon. Furthermore, the HMBC spectrum showed that these methine and methylene protons correlated with carbons C-5, and C-11 and also with an oxygenated quaternary carbon C-9 at δ_{C} 98.2. Finally, a third methyl group at δ_{C} 27.3 (C-17) revealed long range heteronuclear cross peaks to the oxygenated quaternary carbon C-9, to the methylene carbon C-8 and finally to carbons C-6 at δ_{C} 156.9 and C-10 at δ_{C} 152.1 (Figure 2). Combined, these data account for 9 degrees of unsaturation, requiring two additional rings to be present. Overall, NMR data indicated the presence of eight carbon-oxygen bonds.

Owing to both the molecular formula and constraints on connectivities among the different substructures, C-7–C-9 were deduced to be involved in a bicyclic 2,8-dioxabicyclo[3.3.1]nonane framework that bridges the two aromatic rings by incorporating two ether functions. Besides fulfilling hydrogen deficiency index, this proposed structure was well supported by HMBC cross-peaks between H₃-17 and C-6 and C-10 as well as by NOESY correlations between proton H-7 with protons H₂-8, 4-OH and 12-OH, between H₂-8 and H₃-17 and also between 4-OH and 12-OH. The null optical activity of **1** indicated its racemic nature, necessarily corresponding to a mixture of (7*R*, 9*S*)- and (7*S*, 9*R*)- enantiomers owing to its bridged structure. A single crystal X-ray experiment with Mo K α ($\lambda = 0.71073 \text{ \AA}$) (Figure 3) both corroborated the planar structure deduced from spectroscopic data and validated this relative stereochemistry.

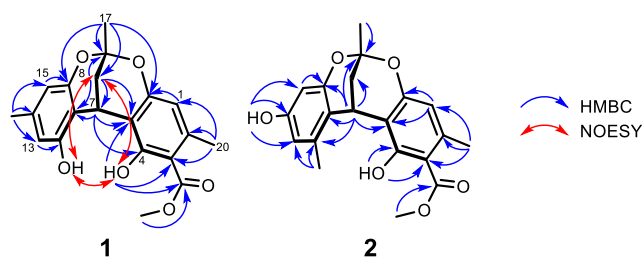


Figure 2. Key HMBC and NOESY correlations for **1** and **2**.

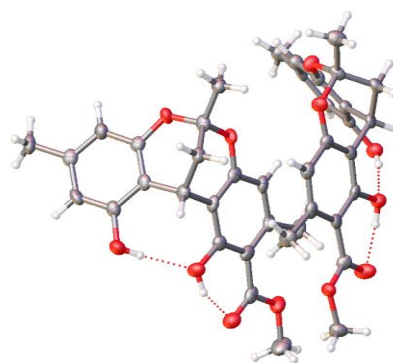


Figure 3. ORTEP diagram of **1**

Compound **2** was isolated as a white amorphous solid with the molecular formula of C₂₀H₂₀O₆ [HRESIMS at m/z 379.1152 [M+Na]⁺ (calcd. 379.1152 for C₂₀H₂₀O₆Na)]. The ¹H and the ¹³C NMR spectral data of **2** (Table 1) were similar to those of **1**. Long-range heteronuclear correlations between H-1 at δ_{H} 6.32 to C-5 (δ_{C} 115.7), C-6 (δ_{C} 155.3), C-3 (δ_{C} 113.0) and C-20 (δ_{C} 19.4) led to define a substitution pattern for a first ring identical to that evidenced through the so-called first spin system of **1** (Figure 2). The 2,8-dioxabicyclo[3.3.1]nonane system was deduced from HMBC correlations of proton H-7 at δ_{H} 4.52 to C-5 (δ_{C} 115.7), to C-10 (δ_{C} 152.2), C-11 (δ_{C} 117.1), C-12 (δ_{C} 138.1), C-8 (δ_{C} 32.8) and C-9 (δ_{C} 98.3). The only difference was the permutation of positions of the hydroxyl and the methyl groups in the second aromatic ring. In **2**, the phenolic hydroxyl group (δ_{H} 8.03) and the methyl moiety (δ_{H} 2.50) were respectively assigned to C-14 and C-12, based on long range heteronuclear connectivities of the phenolic hydroxyl proton with C-14 (δ_{C} 157.2), C-13 (δ_{C} 111.3) and C-15 (δ_{C} 101.7), and of the methyl protons with C-12 (δ_{C} 138.1), C-11 (δ_{C} 117.1) and C-13 (δ_{C} 111.3) (Figure 2). The methyl group anchored at C-12 would indeed be in close spatial proximity to both the methoxycarbonyl and the C-4 phenolic hydroxyl groups. The steric hindrance caused by the bulky methyl group could compromise the coplanarity of the hydrogen-oxygen bond of the C-4 moiety, precluding the chelation of this phenolic group by the vicinal ester carboxyl carbon as in **1** (4-OH, δ_{H} 13.03). Therefore, in **2**, this signal appeared as a phenolic hydroxyl group resonating at δ_{H} 8.94. This might also account for the much higher polarity of **2** compared to that of **1**. On the basis of the aforementioned data, the planar structure of **2** was determined as depicted in Figure 1. Compound **2** was isolated as an optically inactive compound, having no optical rotation nor Cotton effects in the ECD spectrum, determining it to consist of a racemic mixture of the two (7*S*, 9*R*)- and (7*R*, 9*S*)-enantiomers, as previously reported for compound **1**.

Compound **3** was obtained as a white amorphous solid. Its molecular formula, C₃₃H₃₂O₁₀, was established from the [M+Na]⁺ peak at m/z 611.1926 (calcd. 611.1888 for C₃₃H₃₂O₁₀Na). The comparison of ¹³C NMR spectra of **3** and **1** revealed that the chemical structure of the former was highly reminiscent to that of the latter with an addition of thirteen carbons (Table 1). Among these additional carbons, the ¹³C chemical shift values of nine of them were similar to those of an aromatic ring of **1** (C-10–C-16),

suggesting the presence of a second methyl β -orsellinate moiety in **3**. The four remaining carbons corresponded to one methine (δ_{H} 3.93; δ_{C} 30.9, C-7'), one olefinic methine (δ_{H} 4.48; δ_{C} 100.7, C-8'), one oxygenated quaternary olefinic carbon (δ_{C} 149.1, C-9') and one methyl group (δ_{H} 1.85; δ_{C} 19.0, C-10'). Signal patterns related to the common dibenzo-2,8-dioxabicyclo[3.3.1]nonane substructure only differed for two positions (Figure 4 and Table 1). At first, the triplet methine proton at δ_{H} 4.47 (1H, $J = 3.0$ Hz, H-7) turned into a doublet methine proton at δ_{H} 4.35 ($J = 2.0$ Hz, H-7'). A second difference was the disappearance of the diastereotopic methylene signals of H-8 [pair of doublets methylene protons at δ_{H} 2.11 (1H, $J = 13.0, 3.0$ Hz, H-8a), and 2.16 (1H, $J = 13.0, 3.0$ Hz, H-8b) in **1**] to give rise to a doublet of doublets methine proton at δ_{H} 2.44 [(1H, $J = 2.0, 1.5$ Hz) in **3**]. These differences indicated that the additional core was anchored at C-8 position. COSY cross-peaks between H-8 and H-7' further supported this vicinity. The H-7/H-8/H-7'/H-8' connectivity also appeared throughout both COSY correlations and HMBC cross peaks (Figure 4). Altogether, these spectroscopic data led to establish the planar structure of **3** as displayed in Figure 1. NOESY spectrum could grant some information as to its stereochemistry such as the close spatial proximity between H-7' and H-7.

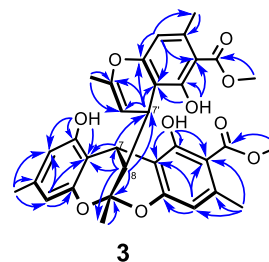


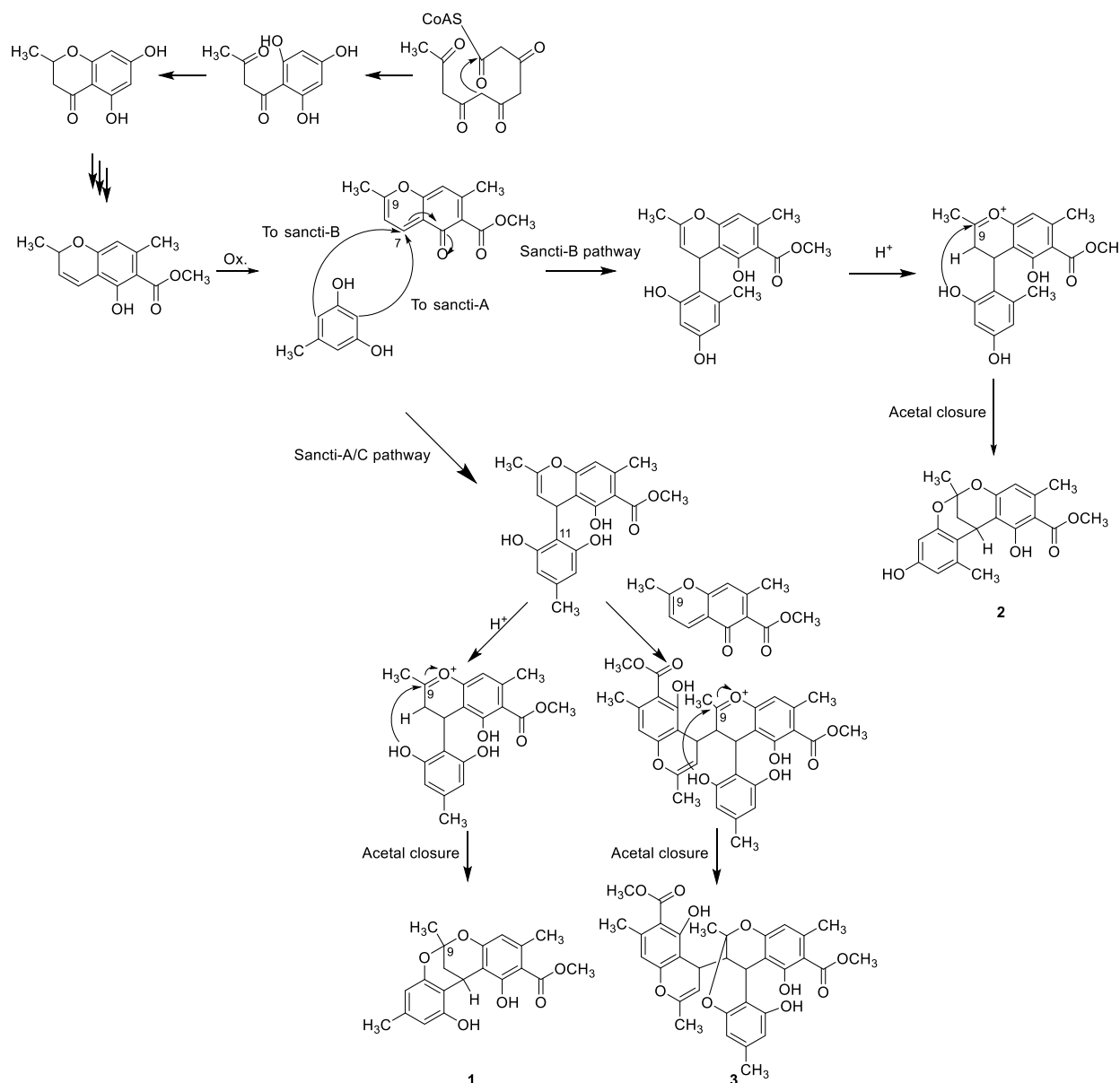
Figure 4. Key HMBC correlations for **3**.

Unfortunately, no spectral evidence could lead to define the relative configuration of C-8, as NOE cross-correlations between H-8 and both H-7' and H-7 might be observed irrespective of the configuration of the former carbon. In an attempt to discriminate between possible diastereoisomers, ^{13}C NMR modeling of the eight possible structures was performed using the well-established calculation of the DP4 probability.^[5] The DP4 application demonstrated the structural equivalence of diastereomer 7*R**, 8*R**, 9*S**, 7'*R** with **3** with 99.80% probability. The lack of optical activity indicated the racemic nature of **3**. Although the minute amount of **3** precluded any attempt of purification, this deduction was confirmed by the observation of two distinct peaks with identical areas by chiral HPLC analysis (Supporting Information; Figure S23).

Table 1. ^1H (500 MHz) and ^{13}C NMR (125 MHz) Data for **1–3**.

position	Sanctis-A (1) (CDCl_3)		Sanctis-B (2) (acetone- d_6)		Sanctis-C (3) (CDCl_3)	
	δ_{H} , mult (J , Hz)	δ_{C}	δ_{H} , mult (J , Hz)	δ_{C}	δ_{H} , mult (J , Hz)	δ_{C}
1	6.34, s	112.8	6.32, s	109.9	6.21, s	111.2
2		141.3		135.9		140.9 ^[a]
3		105.4		113.0		105.2
4		159.1		154.4		159.2
5		112.1		115.7		111.0
6		156.9		155.3		157.3
7	4.47, t (3.0)	21.4	4.52, d (3.0)	24.2	4.35, d (2.0)	24.9
8	2.11, dd (13.0, 3.0) 2.16 dd (13.0, 3.0)	30.9	2.08, m	32.8	2.44, dd (2.0, 1.5)	45.1
9		98.2		98.3		101.0
10		152.1		152.2		151.6
11		110.9		117.1		113.5
12		153.3		138.1		152.7
13	6.37, br d	110.9	6.23, br d	111.3	6.31, br d	111.4
14		138.5		157.2		138.3
15	6.31, br d	109.4	6.15, br d	101.7	6.23, br d	109.1
16	2.19, s	20.4	2.50, s	19.9	2.15, s	21.3
17	1.85, s	27.3	1.72, s	27.3	1.84, s	24.9
18		172.8		168.7		172.5
19	3.93, s	52.3	3.79, s	51.7	3.92, s	52.2
20	2.43, s	24.0	2.10, s	19.4	2.36, s	23.9
1'					6.23, s	112.1
2'						140.7 ^[a]
3'						106.8
4'						162.1
5'						108.5
6'						156.6
7'					3.93, br d	30.9
8'					4.48, br d	100.7
9'						149.1
10'					1.85, s	19.0
11'						172.1
12'					3.89, s	52.2
13'					2.34, s	23.9
4-OH	13.03, s		8.94, s		12.82, s	
12-OH	7.58, s				7.52, s	
14-OH			8.03, s			
4'-OH					11.94, s	

[a] Interchangeable



Scheme 1. Proposed biosynthetic pathway for the formation of **1–3**. Note that the functionalization of the aromatic core of the chromene intermediate may occur prior to the cyclization giving rise to the pyrone ring as well.

The similarities between compounds **1–3** suggest that they share common biosynthetic precursors. Sanctis A–C bear striking similarities to co-isolated compounds methyl- β -orsellinate **9** and orcinol **11**. The biosynthesis of structurally related A-type proanthocyanidols originates from flavanols as early precursors.^[6,7] Due to the lack of a phenyl substituent on C-9, a similar biosynthetic scenario would rely on a chromene precursor (Scheme 1), which is quite interestingly encountered in the structure of **3**. A dozen of chromones are known from lichen sources, being assumed to arise from the polyketide pathway upon condensation of an acetyl CoA with four malonyl CoA units.^[8] While the occurrences of both 2-methyl and 5-phenol groups are straightforward to delineate from a biosynthetic perspective, the origins of the 6-methylester and 7-methyl substituents are conversely difficult to decipher. Nevertheless, some formerly described lichen chromones also reveal some such additional methyl moieties whose origin is not

established neither, at different oxidation states.^[9] As to our specific example, the substitution pattern of the benzene ring of the putative chromone is identical to that of the co-isolated methyl- β -orsellinate **9**. Tailoring enzymes might be responsible for the introduction of these substituents as recently evidenced for the methylation of the closely related orsellinic acid.^[10] Later on, the biosynthetic route to sancti derivatives might rely on the oxidation of the chromone into the corresponding *ortho*-quinone methide.^[6] The subsequent 1,4-Michael addition of orcinol **11** to this *ortho*-quinone methide intermediate would then afford a structure displaying a nucleophilic vinyl ether moiety that is likely to react with an array of electrophiles. According to this putative scheme, reaching aromatic ring substitution pattern of **1/3** or of **2** would depend on the side of the orcinol moiety triggering the nucleophilic attack on C-7 position of the chromene precursor. A nucleophilic attack initiated by the carbon located between the two phenolic groups might give rise to **1/3** regioisomery, as

displayed in Scheme 1. Conversely, if the reactive site would be either of the carbons comprised between one phenolic moiety and the methyl group (due to orcinol being a meso compound), the methyl and phenol groups would be inverted to give rise to **2**. The addition of a proton to the nucleophilic vinyl ether moiety (C-9) might yield an oxocarbenium ion giving access to either **1** or **2** upon acetal closure. This process is not catalyzed by an enzyme and could give rise to the two enantiomers, depending on the face of attack in the first step.^[6,11] Alternatively, the enol ether moiety could react with a second chromene-derived *ortho*-quinone methide as an electrophilic molecule to afford a second oxocarbenium ion that could lead to **3** upon acetal closure. This second nucleophilic attack scenario is deemed to occur with a high degree of diastereoselectivity *trans* to the existing orcinyl substituent, consistently with the determined stereochemistry of **3**. Recent biomimetic syntheses of trimeric flavonoids dragonbloodins A1/A2 described a highly similar mechanism.^[11]

While the 2,8-dioxabicyclo[3.3.1]nonane was seldom encountered in the structure of polyketide-derived fungal metabolites such as averufin,^[12] the molecular context of this core comprising benzene rings on both sides of it makes it an unprecedented report not only among lichen metabolites but also in the wider realm of fungi. Structural homologies with flavanol-derived A-type proanthocyanidols account for both the plant-specific distribution of this structural series so far and for the consensual occurrence of an aromatic ring at the equivalent of position C-9. One exception to this being the example of myristicyclins, isolated from the Myristicaceae plant *Horsfieldia spicata*, containing a hydrogen atom at this specific position.^[13] Instead, the presence of a methyl group in our metabolites makes our carbon skeleton unique, also being consistent with the fact that filamentous fungus produce no flavonoid-related compounds.^[14] A further structural originality reported herein is the occurrence of a cyclic substituent on C-8 position of **3**. It must however be pointed out that such analogues were reported to step in the biomimetic synthesis of trimeric flavonoids from *Daemonorops drcao* although later rearrangements occur so that the final products do not display this scaffold any longer.^[11] The first isolation of these metabolites in the frame of this phytochemical investigation might be explained by the large amount of lichen presently investigated.

Compounds **1**, **2** and **4** were evaluated for their cytotoxic potential against cervical HeLa, breast MCF-7 and lung NCI-H460 cell lines using the sulforhodamine B method^[15] with camptothecin as the positive control (Supporting Information; Table S1). While **2** revealed a mild cytotoxic activity, **1** and **4** were inactive.

Conclusions

Chemical investigation of the methanolic extract of the lichen *Parmotrema sancti-angelii* led to the isolation and structure elucidation of three racemic pairs of procyanidin A analogues for which a biosynthesis pathway is proposed from readily available precursors. Their structure and configurations were assigned on the basis of an extensive set of spectroscopic techniques (1D and 2D NMR, HRMS) along with Goodman probabilities and X-ray crystallography. The carbon skeleton of our newly reported compounds is unique given the pendent methyl group occurring at C-9, indicating their arising from chromene precursors. Instead, flavonoid-derived A-type procyanidins display an aromatic ring at this specific position. Likewise, the occurrence

of an aryl substituent on C-8 represents a further structural novelty.

Experimental Section

General Experimental Procedures: NMR spectra were recorded on a Bruker Avance III 500 MHz spectrometer using residual solvent signal as internal reference: chloroform-*d* δ_{H} 7.26, δ_{C} 77.2 and acetone-*d*₆ δ_{H} 2.1, δ_{C} 206.3, 30.6. The HR-ESI-MS data were recorded on a MicroTOF-Q mass spectrometer. Optical rotations were measured on a Kruss digital polarimeter. Electronic circular dichroism spectra were recorded at room temperature using a Jasco-J815 ECD spectrometer. Thin Layer Chromatography (TLC) analyses were performed on silica gel 60F254 plates with a thickness of 0.25 mm (Merck Millipore, Billerica, Massachusetts, USA). Spots were visualized either by spraying with 10% aqueous H₂SO₄ or with 5% ferric chloride solutions followed by heating. Open column chromatographies were carried out on silica gel 60 (40-63 μm) (HiMedia, Mumbai, India). Crystallographic analyses were carried out with convenient single crystals on an automatic diffractometer (Bruker Model D8 Quest) with Mo K α radiation and graphite monochromatization ($\lambda = 0.71073 \text{ \AA}$) at 100 K. The HPLC system (Prominence Shimadzu, Marne-La-Vallée, France) was equipped with a Chiralpak IA column (250 x 4.6 mm, 5 μm) and consisted of a quaternary pump (LC20ADSP) and a diode array detector (SPD-M20A). The elution was performed under isocratic conditions using a 98/2 (v/v) heptane/isopropanol mixture as the mobile phase at a flow rate of 0.5 mL/min.

Lichen material: Thalli of *Parmotrema sancti-angelii* (Lyngé) Hale were separated from the bark of tea trees *Camelia sinensis* (L.) O. Ktze at Bao Loc city, Lam Dong province, Vietnam in July 2010 (11°29'–11°31' north latitude and 107°44'–107°46' east longitude). The lichen species was identified by Dr. Harrie J.M. Sipman (Berlin Museum). A voucher specimen (No US-B021) was deposited in the herbarium of the Department of Organic Chemistry, University of Science.

Extraction and isolation: The thallus material was washed overnight under a flow of tap water, prior to being rinsed with distilled water to thoroughly cleanse out of all extraneous material. The lichen was then air-dried at ambient temperature to obviate thermally induced decomposition prior to being ground into a fine powder. The powdered lichen thallus (700 g) was macerated with methanol at room temperature to afford a crude methanol extract (120 g). This crude extract was applied to silica gel solid phase extraction, being successively eluted with the following solvents: petroleum ether (60–90 °C) (PE), chloroform (C), ethyl acetate (EA) and methanol (M) to afford corresponding extracts: extract PE (24 g), extract C (20 g), extract EA (14 g) and extract M (30 g). Extract PE (24 g) was applied to silica gel column chromatography to be eluted with PE–EA (stepwise 10:0–0:10), giving 5 fractions (E1–E5). Fraction E4 was chromatographed, eluted with PE–EA (stepwise 9:1–7:3) to afford **8** (40 mg), and **9** (300 mg). Extract C (20 g) was applied to silica gel column chromatography, eluted with PE–EA (95:5–0:10) to give 4 fractions (C1–C4). Fraction C1 (100 mg) was rechromatographed, eluted with PE–C (8:2) to yield three compounds **1** (11 mg), **3** (7 mg) and **10** (8 mg). Fraction C2 (10.1 g) was also selected for further chromatography, eluted with C–EA (9: 1) to provide **5** (300 mg), **6** (900 mg), and **7** (30 mg). Fraction C3 (1.1 g) was rechromatographed, eluted with C–EA (9: 1) to afford **2** (4 mg) and **11** (15.5 mg). Extract EA (14 g) was applied to silica gel column chromatography, eluted with PE–EA (5:5–0:10) to give 2 fractions (EA1–EA2). Fraction EA2 (190 mg) was rechromatographed, eluted with chloroform–acetone (9: 1) to give compound **4** (20 mg).

(±)-Sanctis A (1): Colorless crystals (chloroform–ethyl acetate); $[\alpha]_D^{20}$ 0 (c 0.075, chloroform). HR-ESI-MS m/z 379.1153 $[M+Na]^+$ (calcd. for $C_{20}H_{20}O_6Na$, 379.1152). 1H and ^{13}C NMR data ($CDCl_3$) see Table 1. COSY, HMBC, NOESY correlations see Figure 2. Standard TLC^[16] R_F values: R_F (A): 0.84; R_F (C): 0.72; R_F (F): 0.89. Acid spray: yellow-orange. Vanillin/acid spray: purple.

(±)-Sanctis B (2): White amorphous powder; $[\alpha]_D^{20}$ 0 (c 0.075, chloroform). HR-ESI-MS m/z 379.1152 $[M+Na]^+$ (calcd. for $C_{20}H_{20}O_6Na$, 379.1152). 1H and ^{13}C NMR data (Acetone- d_6) see Table 1. HMBC correlations see Figure 2. Standard TLC^[16] R_F values: R_F (A): 0.39; R_F (C): 0.15; R_F (F): 0.67. Acid spray: brown. Vanillin/acid spray: purple.

(±)-Sanctis C (3): White amorphous powder; $[\alpha]_D^{20}$ 0 (c 0.075, chloroform). HR-ESI-MS m/z 611.1926 $[M+Na]^+$ (calcd. for $C_{33}H_{32}O_{10}Na$, 611.1888). 1H and ^{13}C NMR data ($CDCl_3$) see Table 1. COSY, HMBC, NOESY correlations see Figure 4. Standard TLC^[16] R_F values: R_F (A): 0.86; R_F (C): 0.70; R_F (F): 0.87. Acid spray: orange. Vanillin/acid spray: orange-brown.

X-ray crystallographic analysis of (±)-Sanctis A (1): Formula: $C_{20}H_{20}O_6$, $M_r = 356.36$. Colorless crystal 0.05 x 0.30 x 0.40 mm, $a = 7.6701(9)$, $b = 11.2158(14)$, $c = 19.323(3)$ Å, $\alpha = 92.767(3)$, $\beta = 91.849(3)$, $\gamma = 90.152(3)^\circ$. $V = 1659.4(4)$ Å³, $\rho_{calcd} = 1.426$ Mg/m³, $\mu = 0.105$ mm⁻¹, $Z = 4$, triclinic, $P\bar{1}$; $\lambda = 0.71073$ Å, $T = 100.0(2)$ K, ω and ϕ scans, 21113 reflections collected ($\pm h, \pm k, \pm l$) [$I \geq 2\sigma(I)$], 6161 independent reflections ($R_{int} = 0.0626$), 482 refined parameters, $R = 0.1168$, $\omega R^2 = 0.3380$, max residual electron density 0.53 (-0.44) e.Å⁻³. The data set was collected with a Bruker D8 Quest CMOS diffractometer, absorption correction: multi-scan (SADABS) at VNU-Hanoi University of Science. The structure was solved by intrinsic phase methods using SHELXT and refined on F^2 by using SHELXL^[17] with anisotropic parameters for all non-H-atoms. All the hydrogen atoms were refined isotropically in riding model. A twinning refinement with the proposed twin law [-1.000 0.000 0.000 0.010 1.000 0.000 -0.005 -0.177 -1.000] was done. This resulted in a BASF value of 0.097(2) and about 6.0% improvement in R1 value. The asymmetric unit consisted of two independent molecules which were two enantiomers (Figure 3). The two six-membered heterocyclic rings in both molecules adopted distorted envelope conformations. The bond distances C18A–O5A [1.226(8) Å] and C18B–O5B [1.222(9) Å] are in the typical range of C–O double bonds. The labile hydrogen atoms on O3A/O3B and O4A/O4B were refined as rotating groups in riding model. The obtained results are supported by suitable Q peaks found in the difference electron density map and also by the finding of intramolecular hydrogen bonds O3A–H3A...O4A, O4A–H4A...O5A, O3B–H3B...O4B and O4B–H4B...O5B. The other bond lengths and angles are in normal ranges and comparable to those in its analogue 1,5,13-trimethyl-9-phenyl-8,10-dioxatetracyclo [7.7.1.0^{2,7}.0^{11,16}] heptadeca-2,4,6,11, 13,15-hexaene.^[18]

Computational Details: All DFT calculations have been performed using Gaussian 09W. After a conformational analysis of the 8 diastereoisomers realized with MarvinSketch 17.8.0 ChemAxon, a geometry optimization (B3LYP/6-31g(d)) was undertaken on the most stable conformer of each diastereoisomer. The absence of imaginary frequencies was confirmed by a frequency calculation at the same level. Calculation of the chemical shifts was performed using the same method at the 6-311+g(d,p) level in the gas phase. Comparison of the experimental and predicted chemical shifts was realized using an in-house implementation of the DP4 Java code available from Goodman lab (<http://www-jmg.ch.cam.ac.uk/tools/nmr/DP4/>).

Biological assays: Determination of cytotoxic activities against the HepG2 (liver hepatocellular carcinoma), MCF-7 (human breast cancer)

and NCI-H460 (human lung cancer) cell lines of tested samples was performed at the concentration of 100 µg/mL using the Sulforhodamine B (SRB) assay with camptothecin being used as the positive control. All cells were cultured in EMEM medium (Eagle's Minimal Essential Medium) supplemented with 10% foetal bovine serum (FBS), 1% of 2 mM L-glutamine, 50 IU/mL penicillin, 50 µg/mL streptomycin and cultures were fixed by adding 50 µL of cold 50% (w/v) trichloroacetic acid per well and incubated at 4°C for 1 h. The plates were washed five times with distilled water and air dried. The cells were then stained with 50 µL of SRB (0.4% w/v in 1% acetic acid) at room temperature for 30 min. The SRB solution was removed out of plates by rinsing 4 times with a 1% glacial acetic acid solution (200 µL/well). The plates were air-dried for 12–24 h. The bound SRB was dissolved to each well by adding 100 µL of 10 mM Tris Base (pH 10.5). The plates were shaken gently for 20 min and the optical density of each well was read using a scanning multiwell spectrophotometer at a test wavelength of 492 nm and a reference wavelength of 620 nm. The optical density (OD) of SRB in each well is directly proportional to the cell number. Cell survival was measured as the percentage absorbance compared to the control (non-treated cells) maintained at 37 °C in a 5% CO₂ atmosphere with 95% humidity. Viable cells were counted and inoculated in 96-well plate with a density of 104 cells/100 µL/well. After 24 h, the cells were treated with pure compound while the control wells were only added with 100 µL medium. All experiments were performed in triplicate. The plates were incubated in an atmosphere of 5% CO₂, 95% humidity at 37 °C for 48 h. Adherent cell cultures were fixed by adding 50 µL of cold 50% (w/v) trichloroacetic acid per well and incubated at 4°C for 1 h. The plates were washed five times with distilled water and air dried. The cells were then stained with 50 µL of SRB (0.4% w/v in 1% acetic acid) at room temperature for 30 min. The SRB solution was removed out of plates by rinsing 4 times with a 1% glacial acetic acid solution (200 µL/well). The plates were air-dried for 12–24 h. The bound SRB was dissolved to each well by adding 100 µL of 10 mM Tris Base (pH 10.5). The plates were shaken gently for 20 min and the optical density of each well was read using a scanning multiwell spectrophotometer at a test wavelength of 492 nm and a reference wavelength of 620 nm. The optical density (OD) of SRB in each well is directly proportional to the cell number. Cell survival was measured as the percentage absorbance compared to the control (non-treated cells).

Supporting information: CCDC 1581250 contains the supplementary crystallographic data for compound 1. The data can be obtained free of charge via www.ccdc.cam.ac.uk/data_request/cif, or by emailing data_request@ccdc.cam.ac.uk, or by contacting The Cambridge Crystallographic Data Centre, 12 Union Road, Cambridge CB2 1EZ, UK; fax: +44 1223 336033.

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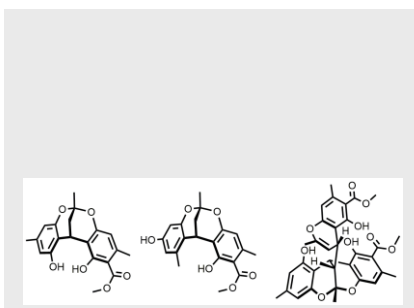
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Keywords: Procyanidin analogues, Lichen, *Parmotrema sancti-angelii*, cytotoxicity, biosynthesis

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FULL PAPER

Three racemic pairs of new procyanidin analogues [sanctis A-C (1-3)] were isolated from the lichen *Parmotrema sancti-angelii* along with 8 other known compounds. These new metabolites all display a novel carbon skeleton.

**Fungal Procyanidin Analogues**

T.-H. Duong, X.-P. Ha, W. Chavasiri, M. A. Beniddir, G. Genta-Jouve, J. Boustie, M. Chollet-Krugler, S. Ferron, H.-H. Nguyen, B. M. Yamin, B.L.C. Huynh, P. Le Pogam, K.-P.-P. Nguyen*.*

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Sanctis A-C: Three racemic procyanidin analogues from the lichen *Parmotrema sancti-angelii*