#### **Supporting Information**

#### From 2- to 3-Substituted Ferrocene Carboxamides or How to Apply Halogen 'dance' to the Ferrocene Series

Mehdi Tazi,<sup>†</sup> William Erb,<sup>\*,†</sup> Yury S. Halauko,<sup>\*,‡</sup> Oleg A. Ivashkevich,<sup>‡</sup> Vadim E. Matulis,<sup>§</sup> Thierry Roisnel,<sup>¶</sup> Vincent Dorcet<sup>¶</sup> and Florence Mongin<sup>†</sup>

<sup>†</sup> Equipe Chimie Organique et Interfaces, Institut des Sciences Chimiques de Rennes, UMR 6226 CNRS-Université de Rennes 1, Bâtiment 10A, Case 1003, Campus Scientifique de Beaulieu, 35042 Rennes Cedex, France

<sup>‡</sup> UNESCO Chair of Belarusian State University, 14 Leningradskaya Str., Minsk 220030, Belarus

<sup>§</sup> Research Institute for Physico-Chemical Problems of Belarusian State University, 14 Leningradskaya Str., Minsk 220030, Belarus

<sup>¶</sup> Centre de Diffractométrie X, Institut des Sciences Chimiques de Rennes, UMR 6226 CNRS-Université de Rennes 1, Bâtiment 10B, Campus de Beaulieu, 35042 Rennes Cedex, France

william.erb@univ-rennes1.fr, hys@tut.by

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#### Optimization of the halogen 'dance' reaction

Table A. Optimization of the halogen 'dance' reaction time from 5 using LiTMP (1.1 equiv) in THF at -50  $^{\circ}$ C.



Table B. Optimization of the halogen 'dance' base from 5 using LiTMP or LiDA (1.1 equiv) in THF at -50 °C for 14 h.



Table C. Optimization of the halogen 'dance' reaction time using LiDA (1.1 equiv) in THF at -50 °C.







Table E. Optimization of the halogen 'dance' temperature from 7 using LiTMP (1.1 equiv) in THF for14 h.



 Table F. Halogen 'dance' reaction at 1.0 and 4.0 mmol scale from the 2-iodoferrocenecarboxamides 6 and 8.



Table G. Halogen 'dance' reaction from 5 using LiTMP in THF at -50 °C for 6 h before hydrolysis or iodolysis.







#### Experimental data on the compounds 1-4 and 6

*N*,*N*-Diisopropylferrocenecarboxamide (1). The general procedure 2 using diisopropylamine (8.5 mL) gave 1 (eluent: heptane-AcOEt 80:20;  $R_f = 0.33$ ) in 94% yield as an orange powder: mp 74 °C; IR (ATR): 806, 824, 1026, 1042, 1369, 1463, 1473, 1627, 2972, 3085 cm<sup>-1</sup>; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, 298 K)  $\delta$  1.20 (br s, 6H, CH<sub>3</sub>), 1.46 (br s, 6H, CH<sub>3</sub>), 3.41 (br s, 1H, *CH*Me<sub>2</sub>), 4.21 (s, 5H, Cp-H), 4.25 (t, 2H, *J* = 1.9 Hz, Cp-H), 4.54 (t, 2H, J = 1.9 Hz, Cp-H), 4.59 (br s, 1H, *CH*Me<sub>2</sub>); <sup>1</sup>H NMR (500 MHz, (CD<sub>3</sub>)<sub>2</sub>SO, 303 K)  $\delta$  1.28 (br s, 12H, CH<sub>3</sub>), 3.49 (br s, 1H, *CH*Me<sub>2</sub>), 4.23 (s, 5H, Cp-H), 4.33 (br s, 2H, Cp-H), 4.48 (br s, 2H, Cp-H), 4.65 (br s, 1H, *CH*Me<sub>2</sub>); <sup>1</sup>H NMR (500 MHz, (CD<sub>3</sub>)<sub>2</sub>SO, 383 K)  $\delta$  1.31 (d, 12H, *J* = 6.7 Hz, CH<sub>3</sub>), 4.04 (sept, 2H, *J* = 6.9 Hz, *CH*Me<sub>2</sub>), 4.23 (s, 5H, Cp-H), 4.31 (br s, 2H, Cp-H), 4.51 (br s, 2H, Cp-H); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>, 298 K)  $\delta$  21.3 (4CH<sub>3</sub>), 46.3 (*C*HMe<sub>2</sub>), 49.6 (*C*HMe<sub>2</sub>), 68.9 (2CH), 69.8 (5CH), 70.0 (2CH), 81.3 (*C*-C=O), 169.4 (C=O). The spectral data were found similar to those already reported.<sup>1</sup>

*N*,*N*-Diethylferrocenecarboxamide (2). The general procedure 1 using ClCONEt<sub>2</sub> (1.6 mL, 7.5 mmol) gave **2** (eluent: heptane-AcOEt 90:10;  $R_f = 0.09$ ) in 86.5% yield as an orange powder: mp 68 °C (lit.<sup>2</sup> 66-67 °C); IR (ATR): 761, 818, 1001, 1106, 1115, 1280, 1378, 1413, 1451, 1462, 1481, 1607, 2932, 2971 cm<sup>-1</sup>; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, 298 K)  $\delta$  1.17 (t, 6H, *J* = 6.4 Hz, CH<sub>3</sub>), 3.46 (br s, 4H, CH<sub>2</sub>), 4.17 (s, 5H, Cp-H), 4.25 (t, 2H, *J* = 2.0 Hz, Cp-H), 4.57 (t, 2H, *J* = 2.0 Hz, Cp-H). These data correspond to those already published.<sup>2</sup> <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>, 298 K)  $\delta$  12.9 (CH<sub>3</sub>), 14.8 (CH<sub>3</sub>), 40.9 (CH<sub>2</sub>), 42.6 (CH<sub>2</sub>), 69.3 (2CH), 69.7 (5CH), 70.3 (2CH), 78.9 (*C*-C=O), 169.5 (C=O).

*N*,*N*-Dimethylferrocenecarboxamide (3). The general procedure 1 using ClCONMe<sub>2</sub> (1.2 mL, 12.5 mmol) gave **3** (eluent: heptane-AcOEt 70:30;  $R_f = 0.12$ ) in 77% yield as an orange powder: mp 112-113 °C (lit.<sup>3</sup> 114-115 °C); IR (ATR): 682, 763, 818, 845, 861, 940, 1000, 1035, 1062, 1103, 1230, 1264, 1390, 1455, 1501, 1611, 2922, 3081 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 291 K)  $\delta$  3.09 (br s, 6H, CH<sub>3</sub>), 4.20 (s, 5H, Cp-H), 4.28 (t, 2H, *J* = 1.9 Hz, Cp-H), 4.60 (t, 2H, *J* = 1.9 Hz, Cp-H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>, 291 K)  $\delta$  36.8 (CH<sub>3</sub>), 38.8 (CH<sub>3</sub>), 69.4 (2CH), 69.9 (5CH), 70.7 (2CH), 78.6 (*C*-C=O), 170.9 (C=O). These data correspond to those already published.<sup>4</sup>

(4-Morpholinocarbonyl)ferrocene (4). The general procedure 1 using 4-morpholinocarbonyl chloride (1.75 mL, 7.5 mmol) gave 4 (eluent: heptane-AcOEt 70:30;  $R_f = 0.12$ ) in 76% yield as a yellowish powder: mp 126 °C (lit.<sup>2</sup> 127-128 °C); IR (ATR): 674, 757, 825, 911, 941, 1001, 1021, 1047, 1063, 1106, 1175, 1254, 1281, 1301, 1411, 1473, 1615, 2050, 2971 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 291 K)  $\delta$  3.66-3.69 (m, 4H, CH<sub>2</sub>), 3.72-3.75 (m, 4H, CH<sub>2</sub>), 4.24 (s, 5H, Cp-H), 4.31 (t, 2H, *J* = 1.9 Hz, Cp-H), 4.54 (t, 2H, *J* = 1.9 Hz, Cp-H). These data correspond to those already published.<sup>2</sup> <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>, 291 K)  $\delta$  45.5 (br s, 2CH<sub>2</sub>), 67.0 (2CH<sub>2</sub>), 69.3 (2CH), 69.9 (5CH), 70.3 (2CH), 78.4 (*C*-C=O), 169.8 (C=O). The <sup>13</sup>C NMR data are similar to those reported previously.<sup>5</sup>

*N,N*-Diethyl-2-iodoferrocenecarboxamide (6, racemic mixture). As already shown,<sup>6</sup> the general procedure 3 using *N*,*N*-diethylferrocenecarboxamide (2, 0.855 g) gave 6 (eluent: heptane-AcOEt 90:10;  $R_f = 0.55$ ) in 88% yield as an orange powder: mp 57 °C; IR (ATR): 688, 808, 1003, 1360, 1442, 1476, 1612, 2966, 3109 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 291 K)  $\delta$  0.98 (br t, 3H, CH<sub>3</sub>), 1.21 (br t, 3H, CH<sub>3</sub>), 3.09 (br q, 2H, *J* = 6.4 Hz, CH<sub>2</sub>), 3.23 (br q, 1H, *J* = 6.4 Hz, CH<sub>2</sub>), 3.72 (br q, 1H, *J* = 6.3 Hz, CH<sub>2</sub>), 4.20 (t, 1H, *J* = 2.5 Hz, Cp-H), 4.34 (s, 5H, Cp), 4.35 (dd, 1H, *J* = 2.6 and 1.4 Hz, Cp-H), 4.45 (dd, 1H, *J* = 2.4 and 1.3 Hz, Cp-H); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, 298 K)  $\delta$  0.96 (br s, 3H, CH<sub>3</sub>), 1.19 (br s, 3H, CH<sub>3</sub>), 3.07 (br s, 2H, CH<sub>2</sub>), 3.21 (br s, 1H, CH<sub>2</sub>), 3.69 (br s, 1H, CH<sub>2</sub>), 4.18 (t, 1H, *J* = 2.45 Hz, Cp-H), 4.32 (s, 5H, Cp-H), 4.33 (br s, 1H, Cp-H); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>, 298 K)  $\delta$  13.05 (CH<sub>3</sub>), 14.3 (CH<sub>3</sub>), 39.7 (CH<sub>2</sub>), 41.1 (C-I), 43.1 (CH<sub>2</sub>), 67.8 (CH), 68.1 (CH), 72.9 (5CH, Cp), 74.1 (CH), 89.7 (*C*-C=O), 167.0 (C=O). These data correspond to those reported previously.<sup>6</sup> It is worth mentioning that the <sup>1</sup>H NMR spectrum recorded in C<sub>6</sub>D<sub>6</sub> at 340 K shows only one signal for the six protons of the methyl groups and one signal for each CH<sub>2</sub>.<sup>6</sup>

#### X-ray diffraction data for the compounds 1-4, 6 and 13



Figure 1. ORTEP diagrams (30% probability level) of the ferrocenecarboxamides 1 ( $C^6-C^{10}-C^{11}-O^{12}$  torsion angle: -66.62°), 2 ( $C^9-C^{10}-C^{11}-O^1$ : -22.90°), 3 ( $C^9-C^{10}-C^{11}-O^{12}$ : 36.07°), 4 ( $C^6-C^{10}-C^{11}-O^{12}$ : 46.24°), 6 ( $C^9-C^{10}-C^{11}-O^{12}$ : -91.57°) and 13.

**Crystal data for** *N***,***N***-diisopropylferrocenecarboxamide** (1). The obtained crystal structure was found to be similar to that described (CCDC 1298115; XUXKUW).<sup>7</sup>

**Crystal data for** *N*,*N*-**diethylferrocenecarboxamide** (2).  $C_{15}H_{19}FeNO$ , M = 285.16, T = 293(2) K, orthorhombic,  $P \ 2_1 \ 2_1 \ 2_1$ , a = 7.5270(11), b = 9.3585(12), c = 19.310(3) Å, V = 1360.2(3) Å<sup>3</sup>, Z = 4, d = 1.392 g cm<sup>-3</sup>,  $\mu = 1.096$  mm<sup>-1</sup>. A final refinement on  $F^2$  with 3106 unique intensities and 167 parameters converged at  $\omega R(F^2) = 0.0888$  (R(F) = 0.0357) for 2923 observed reflections with  $I > 2\sigma(I)$ . CCDC 1565036.

**Crystal data for** *N*,*N*-dimethylferrocenecarboxamide (3).  $C_{13}H_{15}FeNO$ , M = 257.11, T = 150(2) K, orthorhombic, *P n a*  $2_1$ , *a* = 22.0483(8), *b* = 17.3768(6), *c* = 5.8961(2) Å, *V* = 2258.97(14) Å<sup>3</sup>, *Z* = 8, *d* = 1.512 g cm<sup>-3</sup>,  $\mu = 1.311$  mm<sup>-1</sup>. A final refinement on *F*<sup>2</sup> with 4979 unique intensities and 293 parameters converged at  $\omega R(F^2) = 0.0497$  (R(F) = 0.0228) for 4638 observed reflections with  $I > 2\sigma(I)$ . CCDC 1565037. These structural data were found close to those already described (CCDC 1298114; XUXKOQ).<sup>7</sup>

**Crystal data for (4-morpholinocarbonyl)ferrocene (4).**  $C_{15}H_{17}FeNO_2$ , M = 299.14, T = 150(2) K, monoclinic,  $P 2_1$ , a = 5.7178(8), b = 9.4557(13), c = 12.0419(15) Å,  $\beta = 92.803(5)$  °, V = 650.28(15) Å<sup>3</sup>, Z = 2, d = 1.528 g cm<sup>-3</sup>,  $\mu = 1.156$  mm<sup>-1</sup>. A final refinement on  $F^2$  with 2919 unique intensities and 173 parameters converged at  $\omega R(F^2) = 0.0653$  (R(F) = 0.0246) for 2871 observed reflections with  $I > 2\sigma(I)$ . CCDC 1565038. Note that a similar structure to compound **4** was reported in 2005.<sup>8</sup>

**Crystal data for** *N*,*N*-diethyl-2-iodoferrocenecarboxamide (6, racemic mixture). C<sub>15</sub>H<sub>18</sub>FeINO, M = 411.05, T = 150(2) K, monoclinic,  $P 2_1/c$ , a = 11.0250(14), b = 13.6395(16), c = 11.4498(14) Å,  $\beta = 117.212(4)$ °, V = 1531.2(3) Å<sup>3</sup>, Z = 4, d = 1.783 g cm<sup>-3</sup>,  $\mu = 2.993$  mm<sup>-1</sup>. A final refinement on  $F^2$  with 3561 unique intensities and 175 parameters converged at  $\omega R(F^2) = 0.0790$  (R(F) = 0.0297) for 3105 observed reflections with  $I > 2\sigma(I)$ . CCDC 1565039.

#### Table SI-1. Selected <sup>1</sup>H and <sup>13</sup>C NMR data (δ) of 1-4 in CDCl<sub>3</sub>.<sup>*a*</sup>

Compound	$H^{\alpha}$	H <sup>β</sup>	H <sup>Cp</sup>	C <sup>ipso</sup>	C <sup>α</sup>	C <sup>β</sup>	CCp
1	4.54	4.27	4.21	81.3	70.0	68.9	69.8
2	4.57	4.25	4.17	78.9	70.3	69.3	69.7
3	4.60	4.28	4.20	78.6	70.7	69.4	69.9
4	4.54	4.31	4.24	78.4	70.3	69.3	69.9

<sup>*a*</sup> Tentative assignment based on chemical shifts (<sup>1</sup>H)<sup>9</sup> and on HSQC NMR experiment (<sup>13</sup>C).

#### Table SI-2. Selected <sup>1</sup>H and <sup>13</sup>C NMR data ( $\delta$ ) of 5-8 in CDCl<sub>3</sub>.

Compound	$H^3$	$\mathrm{H}^4$	$H^5$	Ср		
$5^{a}$	4.38	4.13	4.24	4.30		
<b>6</b> <sup>b</sup>	4.43	4.18	4.33	4.32		
$7^{b}$	4.37	4.13	4.26	4.21		
$8^{b}$	4.49	4.25	4.36	4.34		
Compound	C	$C^2$	C	$C^4$	C	Ср
$5^{a}$	92.6	40.5	73.6	67.7	66.8	72.7
<b>6</b> <sup>b</sup>	89.7	41.1	74.1	68.1	67.8	72.9
$7^{b}$	87.7	40.3	74.4	68.4	68.3	72.7
$8^{b}$	87.6	40.2	74.8	68.8	67.0	73.1

<sup>*a*</sup> Unambiguous assignment based on advanced NMR studies (see Supporting Information). <sup>*b*</sup> Tentative assignment based on chemical shifts (<sup>1</sup>H)<sup>9</sup> and on HSQC NMR experiment (<sup>13</sup>C).

Table SI-3. Selected <sup>1</sup>H and <sup>13</sup>C NMR data (δ) of ferrocene (Fc-H), iodoferrocene (Fc-I), 1, 5 and 9 in CDCl<sub>3</sub>.

Compound	$H^1$	$H^2$	$H^3$	$H^4$	$H^5$	Ср
Fc-H	4.18	4.18	4.18	4.18	4.18	4.18
$\mathbf{Fc}$ - $\mathbf{I}^{a}$	-	4.43	4.17	4.17	4.43	4.21
$1^{a}$	-	4.54	4.27	4.27	4.54	4.21
<b>5</b> <sup>b</sup>	-	-	4.38	4.13	4.24	4.30
<b>9</b> <sup>b</sup>	-	4.73	-	4.51	4.58	4.30
			-		-	
Compound	$C^1$	$C^2$	C <sup>3</sup>	$C^4$	C <sup>5</sup>	Ср
Compound Fc-H	C <sup>1</sup> 68.1	C <sup>2</sup> 68.1	C <sup>3</sup> 68.1	C <sup>4</sup> 68.1	C <sup>5</sup> 68.1	Cp 68.1
Compound Fc-H Fc-I <sup>a</sup>	C <sup>1</sup> 68.1 39.9	C <sup>2</sup> 68.1 74.6	C <sup>3</sup> 68.1 68.9	C <sup>4</sup> 68.1 68.9	C <sup>5</sup> 68.1 74.6	Cp 68.1 71.2
Compound Fc-H Fc-I <sup>a</sup> 1 <sup>a</sup>	C <sup>1</sup> 68.1 39.9 81.3	C <sup>2</sup> 68.1 74.6 70.0	C <sup>3</sup> 68.1 68.9 68.9	C <sup>4</sup> 68.1 68.9 68.9	C <sup>5</sup> 68.1 74.6 70.0	Cp 68.1 71.2 69.8
$\begin{tabular}{c} \hline Compound \\ \hline Fc-H \\ Fc-I^a \\ 1^a \\ 5^b \end{tabular}$	C <sup>1</sup> 68.1 39.9 81.3 92.6	$     \begin{array}{c}       C^2 \\       68.1 \\       74.6 \\       70.0 \\       40.5     \end{array} $	C <sup>3</sup> 68.1 68.9 68.9 73.6	C <sup>4</sup> 68.1 68.9 68.9 67.7	C <sup>5</sup> 68.1 74.6 70.0 66.8	Cp 68.1 71.2 69.8 72.7

<sup>*a*</sup> Tentative assignment based on chemical shifts (<sup>1</sup>H)<sup>9</sup> and on HSQC NMR experiment (<sup>13</sup>C). <sup>*b*</sup> Unambiguous assignment based on advanced NMR studies (see Supporting Information).

Table SI-4. Chemical shift increments for α, β and Cp positions relative to ferrocene at room temperature in CDCl<sub>3</sub>.

Substituent	Η <sup>α</sup>	$\mathrm{H}^{\beta}$	H <sup>Cp</sup>	
CONiPr <sub>2</sub>	+0.36	+0.09	+0.03	_
I	+0.25	-0.01	+0.03	
Substituent	C <sup>α</sup>	$C^{\beta}$	C <sup>Cp</sup>	C <sup>ipso</sup>
CONiPr <sub>2</sub>	+1.9	+0.8	+1.7	+13.2
I	+6.5	+0.8	+3.1	-28.2

## NMR spectra

## **Compound 1**



<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, 298 K)



<sup>1</sup>H NMR (500 MHz, (CD<sub>3</sub>)<sub>2</sub>SO, 383 K)



<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>, 298 K)



## Compound 2

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, 298 K)



CONEt<sub>2</sub>

# <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>, 298 K)



DEPT-135 (126 MHz, CDCl<sub>3</sub>, 298 K)





## HMBC (300 MHz, CDCl<sub>3</sub>, 298 K)



## Compound 3





CONMe<sub>2</sub>

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, 298 K)



<sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>, 291 K)



<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>, 298 K)





## HSQC (300 MHz, CDCl<sub>3</sub>, 298 K)



# HMBC (300 MHz, CDCl<sub>3</sub>, 298 K)



## **Compound 4**





# <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>, 291 K)



## Compound 5





CONiPr<sub>2</sub>

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>, 298 K)



<sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>, 291 K)



<sup>1</sup>H NMR (500 MHz, (CD<sub>3</sub>)<sub>2</sub>SO, 383 K)



#### COSY NMR (500 MHz, CDCl<sub>3</sub>, 298 K)



## HSQC NMR (500 MHz, CDCl<sub>3</sub>, 298 K)



## HMBC NMR (500 MHz, CDCl<sub>3</sub>, 298 K)



## NOESY NMR (500 MHz, CDCl<sub>3</sub>, 298 K)



## **Compound 6**

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, 298 K)



CONEt<sub>2</sub>

# <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>, 298 K)



DEPT-135 (126 MHz, CDCl<sub>3</sub>, 298 K)



COSY (500 MHz, CDCl<sub>3</sub>, 298 K)



HSQC (500 MHz, CDCl<sub>3</sub>, 298 K)



## NOESY (500 MHz, CDCl<sub>3</sub>, 298 K)



## Compound 7





CONMe<sub>2</sub>

# <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, 298 K)



<sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>, 291 K)



<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>, 298 K)





COSY (300 MHz, CDCl<sub>3</sub>, 291 K)





## HMBC (300 MHz, CDCl<sub>3</sub>, 291 K)



## **Compound 8**

<sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 291 K)



<sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>, 291 K)



## **Compound 9**

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, 298 K)



CONiPr<sub>2</sub>

<sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 291 K)



<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>, 298 K)



<sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>, 291 K)



DEPT-135 (126 MHz, CDCl<sub>3</sub>, 298 K)



COSY NMR (500 MHz, CDCl<sub>3</sub>, 298 K)



HSQC NMR (500 MHz, CDCl<sub>3</sub>, 298 K)



## HMBC NMR (500 MHz, CDCl<sub>3</sub>, 298 K)



# NOESY NMR (500 MHz, CDCl<sub>3</sub>, 298 K)



## **Compound 10**

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, 298 K)



CONEt<sub>2</sub>

# <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>, 298 K)



DEPT-135 (126 MHz, CDCl<sub>3</sub>, 298 K)



## COSY NMR (500 MHz, CDCl<sub>3</sub>, 298 K)



## HSQC NMR (500 MHz, CDCl<sub>3</sub>, 298 K)



## HMBC NMR (500 MHz, CDCl<sub>3</sub>, 298 K)



## NOESY NMR (500 MHz, CDCl<sub>3</sub>, 298 K)





## <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>, 298 K)



DEPT-135 (126 MHz, CDCl<sub>3</sub>, 298 K)



COSY NMR (500 MHz, CDCl<sub>3</sub>, 298 K)



HSQC NMR (500 MHz, CDCl<sub>3</sub>, 298 K)



HMBC NMR (500 MHz, CDCl<sub>3</sub>, 298 K)



#### NOESY NMR (500 MHz, CDCl<sub>3</sub>, 298 K)





# <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>, 291 K)

![](_page_41_Figure_2.jpeg)

#### DEPT-135 (126 MHz, CDCl<sub>3</sub>, 298 K)

![](_page_42_Figure_1.jpeg)

COSY NMR (500 MHz, CDCl<sub>3</sub>, 298 K)

![](_page_42_Figure_3.jpeg)

HSQC NMR (500 MHz, CDCl<sub>3</sub>, 298 K)

![](_page_43_Figure_1.jpeg)

HMBC NMR (500 MHz, CDCl<sub>3</sub>, 298 K)

![](_page_43_Figure_3.jpeg)

NOESY NMR (500 MHz, CDCl<sub>3</sub>, 298 K)

![](_page_44_Figure_1.jpeg)

![](_page_45_Figure_1.jpeg)

![](_page_45_Figure_2.jpeg)

∫NiPr₂

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>, 298 K)

![](_page_45_Figure_4.jpeg)

DEPT-135 (126 MHz, CDCl<sub>3</sub>, 298 K)

![](_page_46_Figure_1.jpeg)

COSY NMR (500 MHz, CDCl<sub>3</sub>, 298 K)

![](_page_46_Figure_3.jpeg)

## HSQC NMR (500 MHz, CDCl<sub>3</sub>, 298 K)

![](_page_47_Figure_1.jpeg)

## HMBC NMR (500 MHz, CDCl<sub>3</sub>, 298 K)

![](_page_47_Figure_3.jpeg)

## NOESY NMR (500 MHz, CDCl<sub>3</sub>, 298 K)

![](_page_48_Figure_1.jpeg)

# 

# **Compound 1.** <sup>1</sup>H NMR (500 MHz, (CD<sub>3</sub>)<sub>2</sub>SO, 383 K), bottom to top: 303 K, 308 K, 318 K, 328 K, 338 K, 348 K, 358 K, 368 K, 378 K, 383 K

![](_page_49_Figure_3.jpeg)

**Compound 5.** <sup>1</sup>H NMR (500 MHz, (CD<sub>3</sub>)<sub>2</sub>SO, 383 K), bottom to top: 298 K, 308 K, 318 K, 328 K, 338 K, 348 K, 358 K, 368 K, 378 K, 383 K

![](_page_50_Picture_1.jpeg)

![](_page_50_Figure_2.jpeg)

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