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**Regioselective galactofuranosylation for the synthesis of disaccharide patterns found in pathogenic microorganisms**

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Vincent Ferrières<sup>\* a</sup>

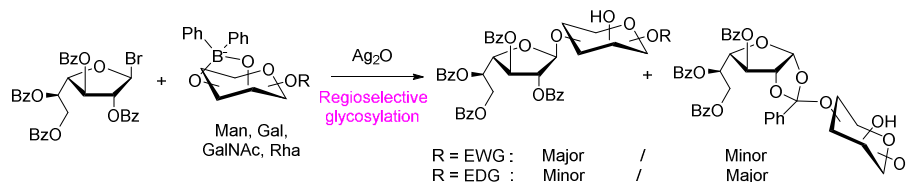
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**Abstract:** Koenigs-Knorr glycosylation of acceptors with more than one free hydroxyl group by 2,3,5,6-tetrabenzoyl galactofuranosyl bromide was performed using diphenylborinic acid 2-aminoethyl ester (DPBA) as inducer of regioselectivity. High regioselectivity for the glycosylation on the equatorial hydroxyl group of the acceptor was obtained thanks to the transient formation of a borinate adduct of the corresponding 1,2-*cis* diol. Nevertheless formation of orthoester by-products hampered the efficiency of the method. Interestingly electron-withdrawing groups on *O*-6 or on *C*-1 of the acceptor displaced the reaction in favour of the desired galactofuranosyl containing disaccharide. The best yield was obtained for the furanosylation of *p*-nitrophenyl 6-*O*-acetyl mannopyranoside. Precursors of other disaccharides, found in the glycocalix of some pathogens, were synthesized according to the same protocol with yields ranging from 45 to 86%. This is a good alternative for the synthesis of biologically relevant glycoconjugates.

## Introduction

The significant discoveries on the organization of the cell, either from prokaryote or eukaryote origin, have pointed out the key role of the glycocalix, the complex layer of hetero- and oligosaccharides that surrounds the cell wall.<sup>1-4</sup> Even if most cells use common sugars to build this glycocalix, they manage to differentiate from each other thanks to the infinite possibilities of sequence and the nature of the linkages involved. Intriguingly some microorganisms incorporate carbohydrates in their furanose form rather than in their pyranose one in order to further differentiate themselves from other organisms.<sup>5,6</sup> In particular D-galactofuranose (D-Galf) containing glycoconjugates are found in large amount in pathogenic microorganisms like *Mycobacteria* or *Leishmania* but are absent in the mammalian kingdom.<sup>7</sup> Owing to the key role played by such motifs in the virulence or survival of the parasite or bacteria, numerous groups have developed evolved synthetic pathways to access to such Galf-containing oligosaccharides.<sup>8-13</sup> Alternatively galactofuranosyl-transferases isolated from mycobacteria or furanosylhydrolases involved in the degradation of biomass have been used to obtain either oligomers of Galf<sup>14,15</sup> or heterodisaccharides.<sup>16,17</sup> These last strategies involve a minimum of protecting group manipulation as biocatalysts are able to selectively transfer a furanosyl entity on a specific position of an acceptor.

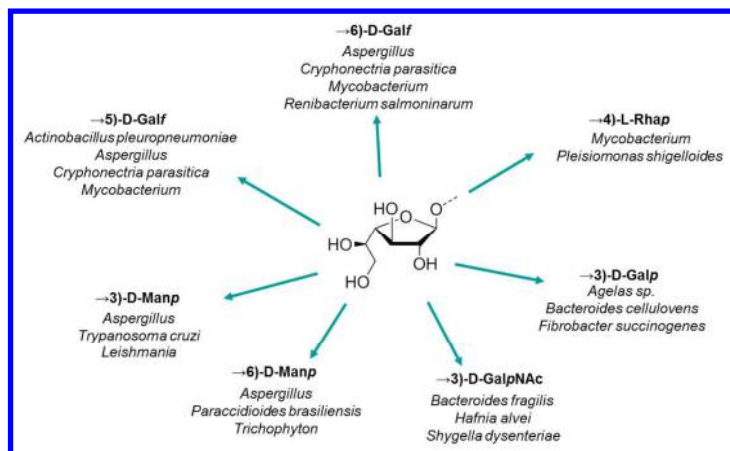
Attempt to mimic such selectivity remains the grail for glycochemists. Already, regioselective glycosylation of naked acceptors could be performed thanks to transient selective activation of one hydroxyl group against the others using for example dibutyltin(IV) oxide<sup>18,19</sup> or aryl borinic acid<sup>20,21</sup> as inducers of regioselectivity. More recently, Taylor and coworkers developed new diarylborinic acid derivatives for the regioselective acylation,<sup>22</sup> alkylation<sup>23</sup> or tosylation<sup>24</sup> of the secondary alcohol of various alkyl glycosides. They postulated that diarylborinic acid could form tetracoordinate adducts with 1,3-diols and 1,2-*cis* diols thus increasing the nucleophilicity of either the primary hydroxyl group or the least hindered

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3 equatorial one. The methodology was recently extended to the glycosylation of thiophenyl  
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5 mannopyranoside using D-arabinofuranosyl methanesulfonate as a donor.<sup>25</sup> It is the first  
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7 example of the synthesis of furanosyl containing disaccharides obtained by this methodology  
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9 and works remain to be done to apply such process to the synthesis of hexofuranosyl-  
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11 containing conjugates.  
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14 After having proposed to work with unprotected thioimidates as donors,<sup>26</sup> we now report on  
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16 the use of diphenylborinic acid 2-aminoethyl ester (DPBA) to induce regioselectivity for  
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18 galactofuranosylation of various glycosidic acceptors: D-mannopyranosides (D-Man<sub>p</sub>), D-  
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20 galactopyranoside (D-Galp), N-acetyl-D-galactopyranoside (D-GalpNAc), L-  
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22 rhamnopyranoside (L-Rhap) and D-Galf. The target compounds are disaccharidic parts of  
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24 biomolecules anchored to the cell wall of pathogenic microorganisms (Figure 1).<sup>5,6</sup>  
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26 Representative examples include β-(1→3) and (1→6)-D-Man<sub>p</sub> linkage found in *Aspergillus*,  
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28 *Trypanosoma* or *Leishmania* sp.; β-(1→3)-D-Galp or β-(1→3)-D-GalpNAc linkage identified  
29  
30 in *Bacteroides*, *Fibrobacter* or *Agelas* sp. for the first one or *Bacteroides* and *Shigella* for the  
31  
32 second one; β-(1→4)-L-Rhap bond presented by *Mycobacterium*.<sup>6,27</sup> Wide ligation diversity  
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34 was also established between two D-Galf entities: β-(1→5) and β-(1→6) found in particular in  
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36 *Mycobacterium tuberculosis*, *Cryphonectria parasitica* and *Aspergillus*.<sup>28</sup>  
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40 Some of these disaccharides linked the Galf non-reducing end to the most nucleophilic  
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42 equatorial oxygen of the sugar at the reducing end. This oxygen is in addition in a 1,2-*cis*  
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44 configuration with one of the proximal hydroxyl group. Such glycosidic bonds could therefore  
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46 be selectively obtained thanks to glycosylation in presence of diarylborinic inducer. First the  
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48 conditions of glycosylation were optimized with mannopyranosidic acceptor using  
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50 peracylated galactofuranosyl bromide as donor. Different groups were introduced on the  
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52 acceptor either on the primary hydroxyl group or at the anomeric position in order to  
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54 investigate the influence of the inductive effect on the glycosylation. An attempt of  
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rationalization of the results was performed thanks to *ab initio* calculation. Finally the methodology was extended to the other relevant acceptors.

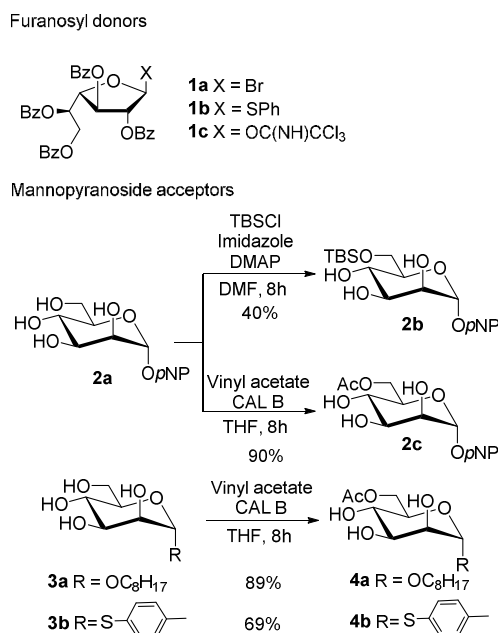


**FIGURE 1. Natural occurrence of galactofuranose in some glyocalix of selected organisms.**

## Results and discussion

Our aim is to develop a regio- and stereoselective glycosylation to obtain Galf-containing disaccharides. The conditions developed by Taylor<sup>29</sup> were first applied on a model reaction between various Galf donors and Manp acceptors (Figure 2). Different activating groups were introduced from peracylated derivatives of galactofuranose, namely the bromide,<sup>30</sup> the thiophenyl<sup>31</sup> or the trichloroacetimidate<sup>32</sup> according to available protocols to give **1a**, **1b** and **1c** respectively. As for the acceptors, we decided to evaluate the influence of the protecting group on both positions *C*-1 and *O*-6. The primary position has to be masked anyway in order to avoid glycosylation at this position. Starting from commercially available *p*-nitrophenyl mannopyranoside (*p*NP Manp) **2a**, the electron-donating group *tert*-butyldimethylsilyl was first introduced on position *O*-6 by action of the corresponding silylating agent TBSCl in presence of imidazole and DMAP with a moderate 40% yield. The electron-withdrawing group acetyl on the other hand was added using vinyl acetate in THF in presence of the supported lipase from *Candida antarctica* (CAL-B). This last strategy was also applied to the

octyl and thiotolyl  $\alpha$ -Manp **3a** and **3b** to obtain the corresponding 6-*O*Ac mannoside **4a** and **4b** with 89% and 69% yields respectively.



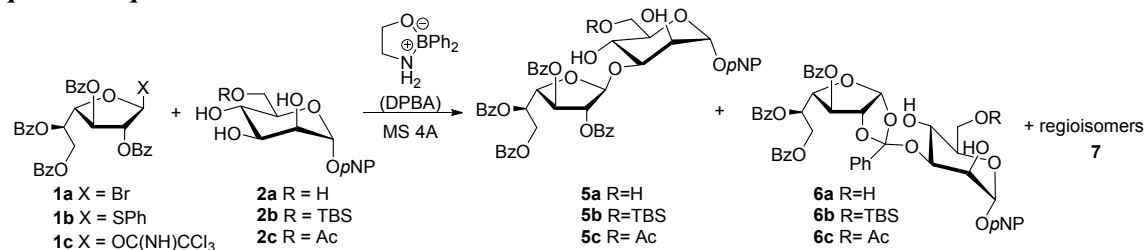
**FIGURE 2. Building blocks used in the regioselective glycosylation.**

With the donors **1** and acceptors **2** in hand, different attempts of glycosylation were implemented (Table 1). Following the findings of Taylor and coworkers, a catalytic amount of diphenylborinic acid 2-aminoethyl ester (DPBA) was used first (Entries 1, 3 to 7). The reactions proceeded in acetonitrile in presence of molecular sieves 4Å in order to limit the hydrolysis of the donors **1**. These donors were used also in excess in the case where the hydrolysis is too high. With TBS Manp **2b** as acceptor and bromide **1a** as donor, the reaction proceeded quickly in presence of both silver oxide and the borinic acid to give the target disaccharide **5b** in 36% yield (Entry 1). It was unambiguously characterized thanks to <sup>1</sup>H and <sup>13</sup>C NMR spectroscopy. The isolated disaccharide presented an anomeric proton at 5.55 ppm associated with a carbon at 103.8 ppm. Coupling constant  $J_{H1,H2}$  was almost zero, a typical value for an anomeric proton of a furanoside in 1,2-*trans* configuration. The regioisomery of the disaccharide was further confirmed by 2D-NMR HMBC experiments. A long range correlation between C<sub>Gal</sub>-1 and H<sub>Man</sub>-3 indicated a (1→3)-glycosidic bond. The other product

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3 of the reaction, and the major one, was identified as the orthoester **6b**. It showed a  
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5 characteristic signal in  $^1\text{H}$  NMR at 6.47 ppm associated with a carbon at 105.3 ppm. The  
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7 related coupling constant  $J_{\text{H}_1,\text{H}_2}$  reached 4.4 Hz thus indicating a 1,2-*cis* configuration. In  
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9 addition  $^{13}\text{C}$  NMR showed a carbon at 123.3 ppm assigned to the quaternary carbon of the  
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11 orthoester function.  
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14 For comparative purpose, we tested the absence of DPBA (Entry 2) or of TBS on the primary  
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16 hydroxyl group of the acceptor (Entry 3). Both conditions led to a complex mixture of  
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18 products. As expected when no borinic acid was added the glycosylation occurred on different  
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20 positions of the acceptor and the disaccharide **5b** was isolated in only 10% yield. Also no  
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22 protecting group on *O*-6 resulted in the concomitant glycosylation of both positions 3 and 6,  
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24 the two positions that were activated by the inducers of regioselectivity.<sup>29</sup> In these conditions  
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26 only trisaccharide 3,6-di-*O*-( $\beta$ -D-Galf)-D-Manp could be isolated.  
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30 Then we tried to isomerize the orthoester **6b** into **5a** by heating the reaction media or by the  
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32 addition of a Lewis acid but only the degradation of the orthoester occurred. Alternatively  
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34 another halogenophile, the silver triflate was used as promoter as it is known to avoid  
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36 orthoester formation. However in these acidic conditions, the TBS group has been cleaved  
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38 and again the trisaccharide 3,6-di-*O*-( $\beta$ -D-Galf)-D-Manp was obtained. Therefore the silyl  
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40 group was swapped by an acetyl one. After reaction between **1a** and **2c** (Entry 4), the major  
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42 product remained the orthoester **6c** and **5c** was obtained only in 19% yield. As the acetyl  
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44 group is tolerant to acidic condition, silver triflate was this time used successfully as promoter  
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46 (Entry 5). It allowed to access to **5c** with a better yield and no orthoester was isolated.  
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48 However other regioisomers, the *p*NP  $\beta$ -D-Galf-(1 $\rightarrow$ 2)-D-Manp and *p*NP  $\beta$ -D-Galf-(1 $\rightarrow$ 4)-D-  
49  
50 Manp were formed as well. It confirms that acidic conditions are detrimental to the formation  
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52 of the borinate complex.<sup>24</sup> Finally, when other Galf donors **1b** and **1c** were used (Entries 6 and  
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60 7), the yield did not improve and the regioselectivity was poorer as previously reported.<sup>29</sup>

**TABLE 1. Optimization of the conditions for the regioselective galactofuranosylation of *p*NP Manp derivatives.**

Entry	Donor (2 eq.)	Acceptor (1 eq.)	Promoter (1 eq.)	DPBA (eq.)	Solvent	Time (h)	<b>5</b> (Yield %)	Ratio <b>5:6:7</b>
1	<b>1a</b>	<b>2b</b>	Ag <sub>2</sub> O	0.1	CH <sub>3</sub> CN	2	<b>5b</b> (36)	5:6:0
2	<b>1a</b>	<b>2b</b>	Ag <sub>2</sub> O	-	CH <sub>3</sub> CN	28	<b>5b</b> (10)	Mixture
3	<b>1a</b>	<b>2a</b>	Ag <sub>2</sub> O	0.1	CH <sub>3</sub> CN	2	-	Trisaccharide <sup>a</sup>
4	<b>1a</b>	<b>2c</b>	Ag <sub>2</sub> O	0.1	CH <sub>3</sub> CN	19	<b>5c</b> (19)	1:2.4:0
5	<b>1a</b>	<b>2c</b>	AgOTf	0.1	CH <sub>3</sub> CN	19	<b>5c</b> (33)	13:0:2 <sup>b</sup>
6	<b>1b</b>	<b>2c</b>	NIS, AgOTf	0.1	CH <sub>3</sub> CN	19	-	-
7	<b>1c</b>	<b>2c</b>	TMSOTf	0.1	CH <sub>3</sub> CN	19	<b>5c</b> (25)	8:0:2 <sup>c</sup>
8	<b>1a</b>	<b>2b</b>	Ag <sub>2</sub> O	1	CH <sub>3</sub> CN	6	<b>5b</b> (46)	7:3:0
9	<b>1a</b>	<b>2b</b>	Ag <sub>2</sub> O	1	CH <sub>2</sub> Cl <sub>2</sub>	8	<b>5b</b> (40)	1:0:0
10	<b>1a</b>	<b>2b</b>	Ag <sub>2</sub> O	1	THF	8	<b>5b</b> (25)	1:1:0
11	<b>1a</b>	<b>2c</b>	Ag <sub>2</sub> O	1	CH <sub>3</sub> CN	2	<b>5c</b> (85)	1:0:0
12	<b>1a</b>	<b>2c</b>	AgOTf	1	CH <sub>3</sub> CN	19	<b>5c</b> (80)	1:0:tr <sup>c</sup>

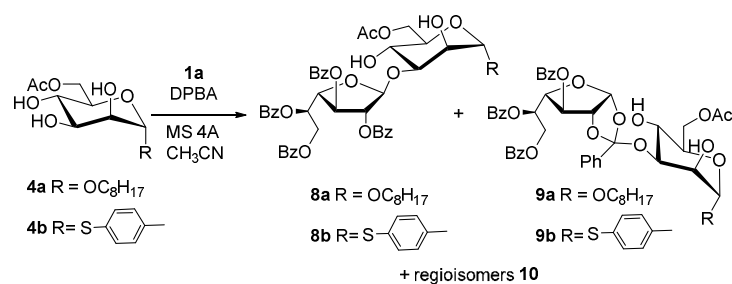
<sup>a</sup> Mainly *p*NP 3,6-di-O-(β-D-Galf)-D-Manp. <sup>b</sup> Mainly *p*NP β-D-Galf-(1→2)-D-Manp and *p*NP β-D-Galf-(1→4)-D-Manp. <sup>c</sup> Mainly *p*NP β-D-Galf-(1→4)-D-Manp

Interestingly, when a stoichiometric amount of DPBA was used, the yield greatly improved and no or trace amount of orthoester was formed (Entries 8 to 12). In addition the use of alternative solvents like THF or dichloromethane decreased either the yields or the selectivity (Entries 9 and 10). Also, the glycosylation of 6-*O*-TBS Manp **2b** proceeded with lower yields than with 6-*O*-Ac Manp **2c**, and a higher amount of orthoester **6b** was isolated (Entries 8 vs. 11). As for the promoter, the acidic AgOTf can be used but trace amount of regioisomers (mainly β-(1→4)-) contaminated again the disaccharide **5c** (Entry 12). The best result was

obtained using the bromide **1a** as donor, activated by one molar ratio of silver oxide, the 6-*O*-acetyl acceptor **2c**, DPBA (one equivalent), in acetonitrile. Under these conditions, the *p*NP β-D-Galf-(1→3)-α-D-Man<sub>p</sub> compound **5c** was obtained in a quite worthy yield of 85% with no contamination by the orthoester (Entry 11).

Following this optimization, the methodology was then applied to 6-*O*-acetylated mannosidic acceptors **4a** and **4b** (Table 2) that differ on the nature of the substituent attached to the anomeric position. On one hand, the presence of the electron-donating group octyl at the anomeric position seemed to favour greatly the formation of the orthoester **9a** (Entry 1). Solely the use of silver trifluoromethanesulfonate instead of silver oxide allowed isolating the target compound **8a** with 57% yield (Entry 2). Once again, contamination by other regioisomers hampered the use of such acidic promoter. On the other hand, a completely different outcome arose with a thiotolyl group on *C*-1 of the mannose as the donor **1a** easily glycosylated the position 3 of the mannoside **4b**. Formation of the orthoester **9b** was low and no other regioisomers were isolated (Entry 3 and 4). It confirms the great influence of the electron-density of the substituent on both position *O*-6 and *C*-1.

**TABLE 2. Extension to octyl and thiotolyl mannoside.**



Entry	Acceptor	Promoter	Time (h)	Disaccharide (Yield %)	Ratio 8:9:10
1	<b>4a</b>	Ag <sub>2</sub> O	2	<b>8a</b> (40)	45:55:0
2	<b>4a</b>	AgOTf	5	<b>8a</b> (57)	4:0:1 <sup>a</sup>
3	<b>4b</b>	Ag <sub>2</sub> O	2	<b>8b</b> (96)	95:5:0
4	<b>4b</b>	AgOTf	6	<b>8b</b> (69)	1:0:0

<sup>a</sup> Mainly octyl β-D-Galf-(1→4)-D-Man<sub>p</sub>.

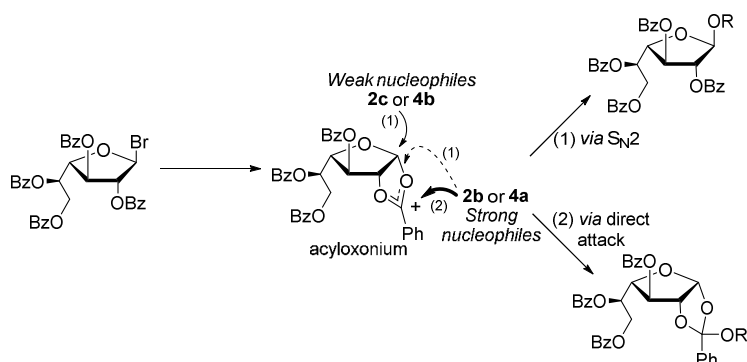
To explain these results, we decided to compare the difference of reactivity of the tested acceptors **2b**, **2c**, **4a** and **4b** in complex with diphenylborinic acid using DFT calculation in the gas-phase (B3LYP/6-31+G\*) (Table 3 and Supporting Information).

**TABLE 3. Calculated Mulliken charges and Fukui indexes of diphenylborinate adduct of **2b**, **2c**, **4a** and **4b** (B3LYP/6-31+G\*).**

Entry	Mannoside acceptor	Mulliken charge at <i>O</i> -3	$f_k$ at <i>O</i> -3
1	<b>2b</b>	-0.35	0.180
2	<b>2c</b>	-0.31	0.115
3	<b>4a</b>	-0.32	0.135
4	<b>4b</b>	-0.33	0.115

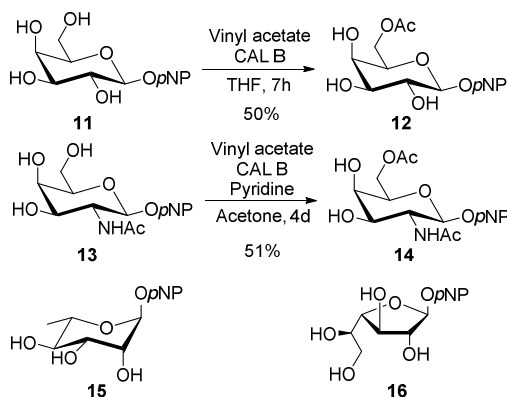
Surprisingly, the estimation of the Mulliken charges on the most reactive equatorial oxygen *O*-3 did not vary much between the different mannosides. Only the Fukui index  $f_k$ , a measure of the strength of the nucleophile,<sup>33</sup> decreased significantly with the presence of electron-withdrawing groups either on *O*-6 or on *C*-1. Interestingly, those molecules **2c** and **4b** also formed no or trace amount of orthoesters. On the contrary, electron-donating groups like the TBS (**2b**) or the alkyl (**4a**) one reinforced the nucleophilicity on *O*-3. Those acceptors generated after glycosylation the largest amount of orthoester. Orthoesters are known to form in various amounts during Koenigs-Knorr glycosylation.<sup>34</sup> According to Taylor *et al*'s work on pyranosyl bromide,<sup>29</sup> the formation of such orthoester is not favoured because the reaction pathway follows a pure S<sub>N</sub>2 mechanism. Here, the presence of the bromide in a 1,2-*trans* configuration and the assistance of the benzoyl group on *O*-2 favour the transient acyloxonium specie and not the oxonium intermediate (Figure 3). The formation of such intermediate is in addition predominant in the furanose series.<sup>35</sup> The attack of the nucleophile could then occur either at the quaternary carbon or at the anomeric one *via* an S<sub>N</sub>2

mechanism. Strong nucleophiles like **4a** or **2b** reacted first at the quaternary carbon and then at the anomeric position leading to a mixture of orthoester and disaccharide. The resulting orthoesters were stable enough to be isolated by silica gel chromatography.<sup>36</sup> On the contrary, if less nucleophilic acceptors **2c** and **4b** were used, the attack occurred only at the anomeric center to give the corresponding disaccharides **5c** and **8b**. For the moment we are not able to decide if such selectivity is under kinetic or thermodynamic control. Nevertheless these results could be compared with the armed/disarmed effect linked with the nature of the substituent on the donor. Such activating/deactivating effects are however rarely reported when dealing with the acceptor and thus pave the way to a new understanding of the acceptor reactivity.



**FIGURE 3. Mechanism of the formation of the orthoester and the disaccharide according to the nucleophilicity of the acceptor.**

The same trend of reactivity was observed when the furanosylation assisted by DPBA was extended to the acceptors *p*NP galactopyranoside **12**, *N*-acetyl-galactosamine **14**, L-rhamnoside **15** and galactofuranoside **16** (Table 4). The building blocks **12** and **14** were obtained through biocatalysed acetylation as before (Figure 4)<sup>21,37,38</sup> while **15** and **16** came from commercial sources.



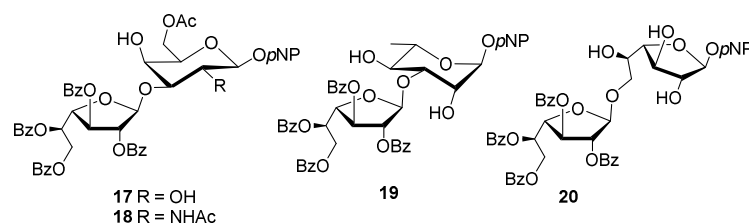
**FIGURE 4. Acceptors used to obtain Galf-containing disaccharides found in various microorganisms.**

On one hand **12** that bears electron-withdrawing groups on *O*-6 and *C*-1 and shows a low Fukui index at *O*-3, gave the corresponding disaccharide **17** with excellent yield and without formation of the orthoester (Entry 1). On the other hand the 6-deoxy-hexose **15** is rather nucleophilic and during the regioselective furanosylation catalysed by silver oxide, more than 30% of orthoester was formed (Entry 4). Alternative silver triflate protocol allowed to increase the yield for disaccharide **19** to 59% but the regioselectivity was decreased and 20% of the regioisomer *p*NP β-Galf-(1→4)-L-Rhap was obtained as well (Entry 5). Surprisingly, *p*NP *N*-acetyl-galactosamine **14** whose calculated Fukui index was very low, gave the corresponding disaccharide **18** with poor yield (Entry 2). The amount of the corresponding orthoester was also low. The low nucleophilicity of *O*-3, due to the presence of the acetamide group on *C*-2 could explain the poor yield for the glycosylation.<sup>39</sup> However 55% yield could be reached with silver triflate as promoter with the downside formation of regioisomers (Entry 3).

The last acceptor tested in the regioselective glycosylation was the *p*NP Galf **16**. In presence of the DPBA, we were expecting to form the diphenylborinate adduct between the *O*-6 and the *O*-5 and thus exacerbating the nucleophilicity of the primary hydroxyl group. Indeed the reaction between the bromide furanosyl **1a** and *p*NP Galf **16** in presence of the borinic acid

and silver oxide led quickly to the total conversion of the acceptor (Entry 6). The minor product of the reaction was then identified as the targeted *p*NP  $\beta$ -Gal $f$ -(1 $\rightarrow$ 6)-Gal $f$  **20** thanks to 2D-NMR experiments. The major one was the orthoester derivative isolated in 56% yield. This was expected as primary hydroxyl groups are much more reactive than secondary hydroxyl groups thus leading to high Fukui index and therefore to a high ratio of orthoester. It is worth to mention nevertheless that no other regioisomer was isolated which means that DPBA indeed formed a complex with *O*-6 and *O*-5 of the Gal $f$  moiety. Finally to prevent from the formation of the orthoester, silver triflate was used and allowed to obtain **20** with 45% yield in mixture with the regioisomer *p*NP  $\beta$ -Gal $f$ -(1 $\rightarrow$ 5)-Gal $f$  (Entry 7).

**TABLE 4. Extension to relevant acceptors to obtain Gal $f$ -containing disaccharides.**



Entry	Acceptor	$f_k$	Promoter	Product (Yield %)	By-product (%)
1	<b>12</b>	0.121 <sup>a</sup>	Ag <sub>2</sub> O	<b>17</b> (86)	None
2	<b>14</b>	0.101 <sup>a</sup>	Ag <sub>2</sub> O	<b>18</b> (10)	Orthoester (37)
3			AgOTf	<b>18</b> (55 <sup>c</sup> )	$\beta$ -Gal $f$ -(1 $\rightarrow$ 4)-Gal $p$ NHAc ( <b>20</b> )
4	<b>15</b>	0.134 <sup>a</sup>	Ag <sub>2</sub> O	<b>19</b> (45)	Orthoester (33)
5			AgOTf	<b>19</b> (59 <sup>c</sup> )	$\beta$ -Gal $f$ -(1 $\rightarrow$ 4)-L-Rhap ( <b>20</b> )
6	<b>16</b>	0.157 <sup>b</sup>	Ag <sub>2</sub> O	<b>20</b> (12)	Orthoester (56)
7			AgOTf	<b>20</b> (45 <sup>c</sup> )	$\beta$ -Gal $f$ -(1 $\rightarrow$ 5)-Gal $f$ (10)

<sup>a</sup> Fukui index at *O*-3. <sup>b</sup> Fukui index at *O*-6. <sup>c</sup> As an inseparable mixture. Yield and ratio determined by <sup>1</sup>H NMR.

## Conclusion

As a conclusion, the high potential of diphenylborinic acid 2-aminoethyl ester as inducer of regioselective galactofuranosylation of various acceptors was confirmed. In this study we have validated the extension of Taylor's methodology from hexopyranosyl bromide to a 1,2-*trans* hexofuranosyl bromide unable to react in a pure S<sub>N</sub>2 glycosylation pathway. When using silver oxide as promoter, only one regioisomer was formed as expected. High amount of orthoester however was formed when the *O*-6 or *C*-1 position of the acceptor possessed an electron-donating group. Silver triflate could be used instead but such an acidic catalyst destabilized the borinate adduct and the resulting regioselectivity dropped. The role of the substituents on the acceptor was partly explained thanks to *ab initio* calculation of the Fukui index of all borinate complexes. Electron-donating groups at primary and anomeric positions significantly increased the Fukui indexes and borinate adducts with high Fukui index on the equatorial oxygen generated the greater amount of orthoester. This is a confirmation that the electron-density of the substituent at the anomeric position of the acceptor can modulate the glycosylation reaction. Finally, despite these limitations, we managed to obtain quickly and efficiently galactofuranosyl containing disaccharides found in the glycocalix of pathogenic microorganisms. The minimum protecting group manipulation and the simplicity of the method represent an attractive alternative to biocatalysed process that still suffer from too strong specificity and low conversion yields.

## Experimental Section

### General Experimental Details

All reactions were carried out in oven-dried glassware. All reagents were purchased from commercial sources and were used without further purification unless noted. Dried acetonitrile, dichloromethane and THF were purchased sealed on molecular sieves. Unless

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2  
3 otherwise stated, all reactions were monitored by TLC on Silica Gel 60 F<sub>254</sub>. TLC spots were  
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5 detected under 254 nm UV-light or by staining with cerium ammonium molybdate solution.  
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7 Column chromatography was performed on Silica Gel (25 or 50  $\mu$ m). Optical rotations were  
8  
9 measured at 20 °C on a Perkin-Elmer 341 polarimeter. NMR spectra were recorded at 400  
10  
11 MHz for <sup>1</sup>H and 100 MHz for <sup>13</sup>C. Chemical shifts are given in  $\delta$  units (ppm) and referenced  
12  
13 to either CDCl<sub>3</sub> or CD<sub>3</sub>OD. Coupling constants *J* were calculated in Hertz (Hz). Proton and  
14  
15 carbon NMR peaks were unambiguously assigned by COSY (double quantum filtered with  
16  
17 gradient pulse for selection), HSQC (gradient echo-anti echo selection and shape pulse) and  
18  
19 HMBC (echo-anti echo gradient selection, magnitude mode) correlation experiments. For  
20  
21 each isolated oligosaccharide, the reducing end (bearing *p*NP or alkyl chain) was quoted as  
22  
23 “a”, and the letter increased toward the non-reducing end (for example the sugar after was  
24  
25 quoted as “b”). High Resolution Masses were ) were recorded in positive mode using direct  
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27 Electrospray ionization on a Waters Q-ToF 2 spectrometer.  
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33 **4-Nitrophenyl 6-*O*-*tert*-butyldimethylsilyl- $\alpha$ -D-mannopyranoside (2b):** To a solution of 4-  
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35 Nitrophenyl  $\alpha$ -D-mannopyranoside **2a** (1.5 g, 4.98 mmol) in DMF (30 mL) were added  
36  
37 imidazole (1.02 g, 14.94 mmol), a catalytic amount of DMAP (30 mg, 0.25 mmol) followed  
38  
39 by TBSCl (901 mg, 5.98 mmol). The reaction mixture was stirred at room temperature until  
40  
41 no evolution was observed through TLC monitoring (8 h). The reaction mixture was diluted  
42  
43 into CH<sub>2</sub>Cl<sub>2</sub> (100 mL) and washed with water (100 mL). The organic layer was washed twice  
44  
45 with a saturated aq NH<sub>4</sub>Cl solution, water and brine. The resulting organic layer was dried  
46  
47 with MgSO<sub>4</sub>, filtered and concentrated under reduced pressure. The remaining silane was  
48  
49 removed by submitting the crude material to vacuum during several hours. The 6-*O*-protected  
50  
51 mannopyranoside **2b** was obtained as a white solid (829 mg, 40%) and could be used without  
52  
53 further purification.  $[\alpha]_D^{20} + 115$  (c 1, MeOH). <sup>1</sup>H NMR (CD<sub>3</sub>OD):  $\delta_H$  8.20 (2H, d, *J* = 9.3 Hz,  
54  
55 H<sub>Ar</sub>), 7.28 (2H, d, *J* = 9.3 Hz, H<sub>Ar</sub>), 5.64 (1H, d, *J*<sub>1,2</sub> = 1.8 Hz, H-1), 4.04 (1H, dd, *J*<sub>2,3</sub> = 3.4  
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3 Hz, H-2), 3.92 (1H, dd,  $J_{6,5} = 2$  Hz,  $J_{6,6'} = 11.2$  Hz, H-6), 3.87 (1H, dd,  $J_{3,4} = 9.3$  Hz, H-3),  
4  
5 3.74 (1H, dd,  $J_{6',5} = 6.8$  Hz, H-6'), 3.65 (1H, app. t,  $J_{4,5} = 9.5$  Hz, H-4), 3.50 (1H, ddd, H-5),  
6  
7 0.8 (9H, s, C(CH<sub>3</sub>)<sub>3</sub>), 0.02 (3H, s, OSi(CH<sub>3</sub>)<sub>2</sub>), 0.00 (3H, s, OSi(CH<sub>3</sub>)<sub>2</sub>). <sup>13</sup>C NMR (CD<sub>3</sub>OD):  
8  
9  $\delta_C$  162.7 (C<sub>Ar</sub>), 143.8 (C<sub>Ar</sub>), 126.7 (C<sub>Ar</sub>), 118.0 (C<sub>Ar</sub>), 100.0 (C-1), 76.6 (C-5), 72.4 (C-3), 71.5  
10 (C-2), 68.4 (C-4), 64.3 (C-6), 26.3 [SiC(CH<sub>3</sub>)<sub>3</sub>], 19.1 [SiC(CH<sub>3</sub>)<sub>3</sub>], -5.15 [OSi(CH<sub>3</sub>)<sub>2</sub>], -5.17  
11 [OSi(CH<sub>3</sub>)<sub>2</sub>]. HRMS (ESI/Q-TOF) m/z: [M+Na]<sup>+</sup> Calcd for C<sub>18</sub>H<sub>29</sub>NO<sub>8</sub>SiNa 438.1560; Found  
12  
13 438.1560.  
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19 **General procedure for the regioselective acetylation of pyranosides:** A solution (0.1 M) of  
20 4-nitrophenyl glycopyranoside in THF was heated to 45 °C until complete dissolution. Then  
21 vinyl acetate (3 equiv.) and Novozym®435 were added (Lipase acrylic resin *Candida*  
22 *antartica* from Novozymes) in a w/w ratio 1:1.2 [glycoside : immobilized enzyme]. The  
23 reaction mixture was stirred at 45 °C until complete conversion of starting material (≈ 7 h).  
24 The enzyme was filtered off and rinsed with MeOH. The resulting filtrate was then  
25 concentrated under reduced pressure and purified by column chromatography on silica gel  
26 (DCM/MeOH ratio 95:5).  
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38 **4-Nitrophenyl 6-O-acetyl- $\alpha$ -D-mannopyranoside (2c):** Synthesized according to general  
39 procedure from *p*NP  $\alpha$ -D-mannopyranoside **2a** (1.0 g, 3.32 mmol), vinyl acetate (918  $\mu$ L, 9.96  
40 mmol) and Novozym®435 (1.2 g) to afford **2c** as a pale yellow solid (1.03 g, 90%).  $[\alpha]_D^{20} +$   
41 128 (c 1, MeOH). <sup>1</sup>H NMR (CD<sub>3</sub>OD):  $\delta_H$  8.24 (2H, d,  $^3J = 9.3$  Hz, H<sub>Ar</sub>), 7.27 (2H, d, H<sub>Ar</sub>),  
42 5.63 (1H, d,  $J_{1,2} = 1.7$  Hz, H-1), 4.33 (1H, dd,  $J_{6,6'} = 11.8$  Hz,  $J_{6a,5} = 2$  Hz, H-6), 4.19 (1H, dd,  
43  $J_{6',5} = 6.4$ , H-6'), 4.05 (1H, dd,  $J_{2,3} = 3.4$  Hz, H-2), 3.88 (1H, dd,  $J_{3,4} = 9$  Hz, H-3), 3.73 (1H,  
44 app. t,  $J_{4,5} = 9.8$  Hz, H-4), 3.61 (1H, ddd, H-5), 1.94 (3H, s, CH<sub>3</sub>CO). <sup>13</sup>C NMR (CD<sub>3</sub>OD):  $\delta_C$   
45 172.7 (CO), 162.6 (C<sub>Ar</sub>), 144.0 (C<sub>Ar</sub>), 126.7 (C<sub>Ar</sub>), 117.9 (C<sub>Ar</sub>), 100 (C-1); 73.5 (C-4), 72.2 (C-  
46 3), 71.5 (C-2), 68.3 (C-5), 64.8 (C-6), 20.7 (CH<sub>3</sub>CO). HRMS (ESI/Q-TOF) m/z: [M+Na]<sup>+</sup>  
47 Calcd for C<sub>14</sub>H<sub>17</sub>NO<sub>9</sub>Na 366.0801 ; Found 366.0797.  
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3 **Octyl 6-*O*-acetyl- $\alpha$ -D-mannopyranoside (4a):** Synthesized according to general procedure  
4  
5 from octyl  $\alpha$ -D-mannopyranoside **3a**<sup>40</sup> (300 mg, 1 mmol), vinyl acetate (284  $\mu$ L, 3.31 mmol)  
6  
7 and Novozym<sup>®</sup>435 (400 mg) to afford **2c** as a colorless oil (305 mg, 90%).  $[\alpha]_D^{20} + 43$  (c 1,  
8  
9 MeOH). <sup>1</sup>H NMR (CD<sub>3</sub>OD):  $\delta_H$  4.70 (1H, d,  $J_{1,2} = 1.6$  Hz, H-1), 4.39 (1H, dd,  $J_{6,6'} = 11.7$  Hz,  
10  
11  $J_{6,5} = 2.0$  Hz, H-6), 4.20 (1H, dd,  $J_{6',5} = 6.4$  Hz, H-6'), 3.79 (1H, dd,  $J_{2,3} = 3.2$  Hz, H-2), 3.73–  
12  
13 3.65 (3H, m, H-5, H-3, OCH<sub>2</sub>), 3.60 (1H, t,  $J_{4,5} = 9.4$  Hz, H-4), 3.42 (1H, dt,  $^2J = 9.6$  Hz,  $^3J =$   
14  
15 6.3 Hz, OCH<sub>2</sub>), 2.06 (3H, s, COCH<sub>3</sub>), 1.66–1.54 (2H, m, OCH<sub>2</sub>CH<sub>2</sub>), 1.45–1.27 (10H, m,  
16  
17 (CH<sub>2</sub>)<sub>5</sub>CH<sub>3</sub>), 0.91 (3H, t,  $^3J = 6.8$  Hz, CH<sub>2</sub>CH<sub>3</sub>). <sup>13</sup>C NMR (CD<sub>3</sub>OD):  $\delta_C$  172.8 (CO), 101.7  
18  
19 (C-1), 72.6 (C-3), 72.1, 72.1 (C-2, C-5), 68.7, 68.7 (C-4, OCH<sub>2</sub>), 65.3 (C-6), 33.0, 30.6, 30.5,  
20  
21 30.4, 27.4, 23.7[(CH<sub>2</sub>)<sub>5</sub>CH<sub>3</sub>], 20.8 (COCH<sub>3</sub>), 14.4 (CH<sub>2</sub>CH<sub>3</sub>). HRMS (ESI/Q-TOF) m/z:  
22  
23 [M+Na]<sup>+</sup> Calcd for C<sub>16</sub>H<sub>30</sub>O<sub>7</sub>Na 357.18837 ; Found 357.1882.  
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28 **Thiotolyl 6-*O*-acetyl- $\alpha$ -D-mannopyranoside (4b):** Synthesized according to general  
29  
30 procedure from thiotolyl  $\alpha$ -D-mannopyranoside **3b**<sup>41</sup> (780 mg, 2.7 mmol), vinyl acetate (750  
31  
32  $\mu$ L, 8.2 mmol) and Novozym<sup>®</sup>435 (1 g) to afford **4b** as a colorless oil (624 mg, 69%).  $[\alpha]_D^{20}$   
33  
34 +244 (c 1.25, MeOH). <sup>1</sup>H NMR (CD<sub>3</sub>OD):  $\delta_H$  7.40 (2H, d,  $^3J = 8.1$  Hz, H<sub>Ar</sub>), 7.14 (2H, d,  
35  
36 H<sub>Ar</sub>), 5.35 (1H, d,  $J_{1,2} = 1.5$  Hz, H-1), 4.43-4.35 (1H, m, H-6), 4.29-4.20 (2H, m, H-5, H-6'),  
37  
38 4.07 (1H, dd,  $J_{2,3} = 2.9$  Hz, H-2), 3.72-3.64 (2H, m, H-3, H-4), 2.32 (3H, s, CH<sub>3</sub>), 2.00 (3H, s,  
39  
40 COCH<sub>3</sub>). <sup>13</sup>C NMR (CD<sub>3</sub>OD):  $\delta_C$  172.7 (CO), 139.0, 133.5, 131.7, 130.7 (C<sub>Ar</sub>), 90.3 (C-1),  
41  
42 73.4 (C-2), 73.0, 72.9 (C-4, C-5), 69.0 (C-3), 65.0 (C-6), 21.1 (CH<sub>3</sub>), 20.8 (COCH<sub>3</sub>). HRMS  
43  
44 (ESI/Q-TOF) m/z: [M+Na]<sup>+</sup> Calcd for C<sub>15</sub>H<sub>20</sub>O<sub>6</sub>SNa 351.08728 ; Found 351.0872.  
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49 **4-Nitrophenyl 6-*O*-acetyl- $\beta$ -D-galactopyranoside (12):** Synthesized according to general  
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51 procedure from *p*NP  $\beta$ -D-galactopyranoside **11** (100 mg, 0.3 mmol), vinyl acetate (100  $\mu$ L,  
52  
53 1mmol) and Novozym<sup>®</sup>435 (120 mg) to afford **12** as a colorless oil (56 mg, 50%). <sup>1</sup>H NMR  
54  
55 (CD<sub>3</sub>OD+CDCl<sub>3</sub>):  $\delta_H$  8.20 (2H, d,  $^3J = 9.3$  Hz, H<sub>Ar</sub>), 7.19 (2H, d, H<sub>Ar</sub>), 4.99 (1H, d,  $J_{1,2} = 7.7$   
56  
57 Hz, H-1), 4.34 (1H, dd,  $J_{6,6'} = 11.5$  Hz,  $J_{6,5} = 7.8$  Hz, H-6), 4.25 (1H, dd,  $J_{6',5} = 4.5$  Hz, H-6'),  
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3 3.99–3.91 (1H, m, H-5), 3.90 (1H, dt,  $J_{4,3} = 3.5$  Hz,  $J_{4,5} = 0.9$  Hz, H-4), 3.85 (1H, dd,  $J_{2,3} = 9.7$   
4 Hz, H-2), 3.61 (1H, dd, H-3), 2.07 (3H, s, CH<sub>3</sub>). <sup>13</sup>C NMR (CD<sub>3</sub>OD): δ<sub>C</sub> 172.2 (CO), 163.4,  
5 143.5, 126.3, 117.4 (C<sub>Ar</sub>), 101.6 (C-1), 74.1 (C-3, C-5), 71.4 (C-2), 69.5 (C-4), 64.4 (C-6),  
6 20.9 (CH<sub>3</sub>). HRMS (ESI/Q-TOF) m/z: [M+Na]<sup>+</sup> Calcd for C<sub>14</sub>H<sub>17</sub>NO<sub>9</sub>Na 366.0801 ; Found  
7 366.0804.  
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14 **4-Nitrophenyl 6-O-acetyl-2-deoxy-β-D-N-acetylgalactosamine (14)**: Synthesized according  
15 to described procedure<sup>38</sup> from *p*NP β-D-N-acetylgalactosamine **13** (280 mg, 0.8 mmol), vinyl  
16 acetate (10 mL) Novozym<sup>®</sup> 435 (350 mg) in a mixture of acetone/pyridine (29/21 mL) to  
17 afford **14** as a white solid (161 mg, 51%). Spectroscopic data were in accordance with  
18 previous report.  
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27 **General procedure for the 2-ADPB-assisted glycosidic coupling**: Bromide donor **1a**<sup>30</sup> (2  
28 equiv.), acceptor (1 equiv.) and 2-aminoethyl diphenylborinate (2-DPBA, 1 equiv.) were  
29 suspended in dry CH<sub>3</sub>CN (40 mM) with activated 4 Å molecular sieves (100 mg / mL) and  
30 under a nitrogen atmosphere. The mixture was stirred for 30 min and Ag<sub>2</sub>O (1 equiv.) was  
31 added. Alternatively, the mixture was protected from light and AgOTf (1 equiv.) was used.  
32 After stirring at room temperature, when no evolution was observed through TLC monitoring,  
33 the reaction mixture was diluted with EtOAc and filtered through a plug of Celite. In the case  
34 of AgOTf-promoted reactions, a few drops of Et<sub>3</sub>N were added to neutralize the reaction prior  
35 to dilution and filtration. The filtrate was concentrated under reduced pressure and the  
36 resulting crude material was purified by column chromatography on silica gel (cyclohexane/  
37 ethyl acetate ratio 4:1).  
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52 **4-Nitrophenyl 2,3,5,6-tetra-O-benzoyl-β-D-galactofuranosyl-(1→3)-6-O-tert-**  
53 **butyldimethylsilyl-α-D-mannopyranoside (5b)**: Synthesized according to general procedure  
54 starting from 6-O-TBS-mannopyranoside **2b** (63 mg, 0.15 mmol), galactofuranosyl bromide  
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**1a** (200 mg, 0.30 mmol), 2-DPBA (34 mg, 0.15 mmol) and Ag<sub>2</sub>O (35 mg, 0.15 mmol). Reaction reached completion after 6 h. Purification by column chromatography on silica gel afforded **5b** (70 mg, 46%) as a white solid. Further elution allowed isolating the corresponding orthoester **6b** (37 mg, 24%).

**5b**: [ $\alpha$ ]<sub>D</sub><sup>20</sup> + 12 (c 1, CH<sub>2</sub>Cl<sub>2</sub>). <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$ <sub>H</sub> 8.19 (2H, d, *J* = 9.3 Hz, H<sub>Ar</sub>), 8.10, 8.04, 7.99, 7.93 (8H, 4dd, *J* = 8.3 Hz, *J* = 1.2 Hz, H<sub>Ar</sub>), 7.61-7.29 (12H, m, H<sub>Ar</sub>), 7.15 (2H, d, H<sub>Ar</sub>), 6.03-5.98 (1H, m, H-5b), 5.76 (1H, dd, *J*<sub>3b,4b</sub> = 5.8 Hz, *J*<sub>3b,2b</sub> = 2 Hz, H-3b), 5.70 (1H, d, *J*<sub>1a,2a</sub> = 1.5 Hz, H-1a), 5.55 (1H, bs, H-1b), 5.49 (1H, dd, *J*<sub>2b,1b</sub> = 0.7 Hz, H-2b), 4.95 (1H, dd, *J*<sub>4b,5b</sub> = 3.6 Hz, H-4b), 4.80 (2H, d, *J*<sub>6b,5b</sub> = 4.9 Hz, H-6b), 4.28 (1H, bs, H-2a), 4.14 (1H, dd, *J*<sub>3a,2a</sub> = 3.3 Hz, *J*<sub>3a,4a</sub> = 9.3 Hz, H-3a), 4.02 (1H, app. td, *J*<sub>4a,5a</sub> = 9.6 Hz, *J*<sub>4a,OH</sub> = 2.7 Hz, H-4), 3.87 (1H, dd, *J*<sub>6a,6'a</sub> = 11 Hz, *J*<sub>6a,5a</sub> = 4.2 Hz, H-6a), 3.81 (1H, dd, *J*<sub>6'a,5a</sub> = 5.4 Hz, H-6'a), 3.65-3.58 (1H, m, H-5a), 3.17 (1H, d, OH), 2.94 (1H, d, *J*<sub>OH,2a</sub> = 2.5 Hz, OH), 0.84 (9H, s, *t*Bu-Si), 0.04 [6H, s, OSi(CH<sub>3</sub>)<sub>2</sub>]. <sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta$ <sub>C</sub> 166.3, 166.2, 165.7, 165.6 (COPh), 160.8, 142.6 (C<sub>pNP</sub>), 133.8, 133.7, 133.4, 133.2, 130.2, 129.9, 129.8, 129.4, 129.3, 128.8, 128.5, 128.4 (C<sub>Ph</sub>), 125.8, 116.5 (C<sub>pNP</sub>), 103.8 (C-1b), 97.7 (C-1a), 83.1 (C-2b), 81.2 (C-4b), 77.3 (C-3a), 76.8 (C-3b), 73.0 (C-5a), 70.1 (C-5b), 67.5 (C-2a), 66.8 (C-4a), 63.6 (C-6a), 63.3 (C-6b), 25.8 [SiC(CH<sub>3</sub>)<sub>3</sub>], 18.2 [SiC(CH<sub>3</sub>)<sub>3</sub>], -5.5, -5.4 [OSi(CH<sub>3</sub>)<sub>2</sub>]. HRMS (ESI/Q-TOF) *m/z*: [M+Na]<sup>+</sup> Calcd for C<sub>52</sub>H<sub>55</sub>NO<sub>17</sub>SiNa 1016.3137 ; Found 1016.3134.

**6b**: [ $\alpha$ ]<sub>D</sub><sup>20</sup> + 65 (c 1, CH<sub>2</sub>Cl<sub>2</sub>). <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$ <sub>H</sub> 8.16 (2H, d, *J* = 9.3 Hz, H<sub>Ar</sub>), 7.97, 7.93, 7.86, 7.76 (8H, 4dd, *J* = 8.5 Hz, *J* = 1.3 Hz, H<sub>Ar</sub>), 7.61-7.30 (12H, m, H<sub>Ar</sub>), 7.06 (2H, d, *J* = 9.3 Hz, H<sub>Ar</sub>), 6.47 (1H, d, *J*<sub>1b,2b</sub> = 4.4 Hz, H-1b), 5.56 (1H, dd, *J*<sub>3b,4b</sub> = 3.5 Hz, *J*<sub>3b,2b</sub> = 0.9 Hz, H-3b), 5.47 (1H, d, *J*<sub>2a,1a</sub> = 1.8 Hz, H-1a), 5.37-5.32 (1H, m, H-5b), 5.30 (1H, dd, H-2b), 4.69 (1H, dd, *J*<sub>4b,5b</sub> = 8.0 Hz, H-4b), 4.56 (1H, dd, *J*<sub>6b,6'b</sub> = 12.5 Hz, *J*<sub>6b,5b</sub> = 3.9 Hz, H-6b), 4.38 (1H, dd, *J*<sub>6'b,5b</sub> = 4.9 Hz, H-6'b), 4.08 (1H, dd, *J*<sub>3a,2a</sub> = 3.3 Hz, *J*<sub>3a,4a</sub> = 9.4 Hz, H-3a), 3.93 (1H, app. td, *J*<sub>4a,5a</sub> = 9.5 Hz, *J*<sub>4a,OH</sub> = 2 Hz, H-4a), 3.78 (2H, d, *J*<sub>6a,5a</sub> = 4.3 Hz, H-6a), 3.64 (1H, br s,

H-2a), 3.58-3.53 (1H, m, H-5a), 3.04 (1H, d, OH), 2.37(1H, d,  $J_{\text{OH},2a} = 2.8$  Hz, OH), 0.83 (9H, s, *t*Bu-Si), 0.02 [6H, s, OSi(CH<sub>3</sub>)<sub>2</sub>]. <sup>13</sup>C NMR (CDCl<sub>3</sub>): δ<sub>C</sub> 165.6, 165.3, 165.2 (COPh), 160.8, 142.6 (C<sub>pNP</sub>), 136.1, 133.7, 133.4, 133.2, 133.1, 130.0, 129.9, 129.8, 129.6, 129.5, 129.4, 128.6, 128.5, 128.3, 128.2, 125.8 (C<sub>Ph</sub>), 125.7 (C<sub>pNP</sub>), 123.3 (PhCO<sub>3</sub>), 116.3 (C<sub>pNP</sub>), 105.3 (C-1b), 97.4 (C-1a), 85.7 (C-2b), 83.9 (C-4b), 76.8 (C-3b), 73.7 (C-3a), 72.7 (C-5a), 71.2 (C-5b), 69.0 (C-2a), 67.3 (C-4a), 64.0 (C-6a), 62.8 (C-6b), 25.8 [SiC(CH<sub>3</sub>)<sub>3</sub>], 18.3 [SiC(CH<sub>3</sub>)<sub>3</sub>], -5.5, -5.6 [OSi(CH<sub>3</sub>)<sub>2</sub>]. HRMS (ESI/Q-TOF) m/z: [M+Na]<sup>+</sup> Calcd for C<sub>52</sub>H<sub>55</sub>NO<sub>17</sub>SiNa 1016.3137 ; Found 1016.3130.

**4-Nitrophenyl 2,3,5,6-tetra-O-benzoyl-β-D-galactofuranosyl-(1→3)-6-O-acetyl-α-D-**

**mannopyranoside (5c):** Synthesized according to general procedure starting from 6-O-Ac-mannopyranoside **2c** (52 mg, 0.15 mmol), galactofuranosyl bromide **1a** (200 mg, 0.30 mmol), 2-DPBA (34 mg, 0.15 mmol) and Ag<sub>2</sub>O (35 mg, 0.15 mmol). Reaction reached completion after 2 h. Purification by column chromatography on silica gel afforded **5c** (119 mg, 85%) as a white solid.  $[\alpha]_{\text{D}}^{20} + 37.5$  (c 0.8, CH<sub>2</sub>Cl<sub>2</sub>). <sup>1</sup>H NMR (CDCl<sub>3</sub>): δ<sub>H</sub> 8.20 (2H, d,  $J = 9.3$  Hz, H<sub>Ar</sub>), 8.11, 8.02, 7.99, 7.93 (8H, 4 dd,  $J = 1.3$  Hz,  $J = 8.4$  Hz, H<sub>Ar</sub>), 7.60-7.32 (12H, m, H<sub>Ar</sub>), 7.14 (2H, d, H<sub>Ar</sub>), 5.99-5.94 (1H, m, H-5b), 5.77 (1H, dd,  $J_{3b,2b} = 2.2$  Hz,  $J_{3b,4b} = 6.0$  Hz, H-3b), 5.71 (1H, d,  $J_{1a,2a} = 1.5$  Hz, H-1a), 5.54 (1H, br s, H-1b), 5.49 (1H, dd,  $J_{2b,1b} = 0.7$  Hz, H-2b), 4.95 (1H, dd,  $J_{4b,5b} = 3.7$  Hz, H-4b), 4.83 (1H, dd,  $J_{6b,6'b} = 11.9$  Hz,  $J_{6b,5b} = 4.8$  Hz, H-6b), 4.78 (1H, dd,  $J_{6'b,5b} = 6.1$  Hz, H-6'b), 4.39 (1H, dd,  $J_{6a,5a} = 5.2$  Hz,  $J_{6a,6'a} = 12.2$  Hz, H-6a), 4.31 (1H, br s, H-2a), 4.27 (1H, dd,  $J_{6'a,5a} = 2.2$  Hz, H-6'a), 4.13 (1H, dd,  $J_{3a,2a} = 3.3$  Hz,  $J_{3a,4a} = 9.3$  Hz, H-3), 3.96 (1H, app. td,  $J_{4a,\text{OH}} = 3.4$  Hz,  $J_{4a,5a} = 9.7$  Hz, H-4a), 3.74 (1H, ddd, H-5a), 3.24 (1H, d, OH), 3.08 (1H, d,  $J_{2a,\text{OH}} = 1.9$  Hz, OH), 2.04 (3H, s, CH<sub>3</sub>CO). <sup>13</sup>C NMR (CDCl<sub>3</sub>): δ<sub>C</sub> 171.3 (COCH<sub>3</sub>), 166.3, 165.6 (COPh), 160.6, 142.8, 133.8, 133.7, 133.5, 133.3, 130.0, 129.9, 129.8, 129.4, 129.3, 128.7, 128.6, 128.5, 128.4, 125.7, 116.4 (C<sub>Ar</sub>), 103.9 (C-1b), 97.8 (C-1a), 83.3 (C-2b), 81.0 (C-4b), 77.0 (C-3a), 76.7 (C-3b), 71.7 (C-5a), 70.0 (C-5b), 67.6 (C-2a), 65.3

(C-4a), 63.1, 63.0 (C-6a, C-6b), 20.8 (CH<sub>3</sub>CO). HRMS (ESI/Q-TOF) m/z: [M+Na]<sup>+</sup> Calcd for C<sub>48</sub>H<sub>43</sub>NO<sub>18</sub>Na 944.23723 ; Found 944.2391.

**Octyl 2,3,5,6-*O*-benzoyl-β-D-galactofuranosyl)-(1→3)-6-*O*-acetyl-α-D-mannopyranoside**

**(8a):** Synthesized according to general procedure starting from 6-*O*-Ac-mannopyranoside **4a** (51 mg, 0.15 mmol), galactofuranosyl bromide **1a** (200 mg, 0.30 mmol), 2-DPBA (34 mg, 0.15 mmol) and Ag<sub>2</sub>O (35 mg, 0.15 mmol). Reaction reached completion after 2 h. Purification by column chromatography on silica gel afforded **8a** (56 mg, 40%) as a white solid. Further elution allowed isolating the corresponding orthoester **9a** (68 mg, 49%).

Alternative protocol using **4a** (44 mg, 0.13 mmol), **1a** (176 mg, 0.27 mmol), 2-DPBA (30 mg, 0.13 mmol) and AgOTf (34 mg, 0.13 mmol) as promoter gave **8a** (70 mg, 57%) in mixture with the octyl β-D-Galf-(1→4)-D-Manp regioisomer (ratio 4:1).

**8a:** [ $\alpha$ ]<sub>D</sub><sup>20</sup> + 6 (c 1.5, CHCl<sub>3</sub>). <sup>1</sup>H NMR (CDCl<sub>3</sub>): δ<sub>H</sub> 8.13-7.88 (8H, m, H<sub>Ar</sub>), 7.62-7.47 (4H, m, H<sub>Ar</sub>), 7.46-7.28 (8H, m, H<sub>Ar</sub>), 5.96 (1H, dt,  $J_{5b,6'b}$  = 6.2 Hz,  $J_{5b,6b}$  = 4.7 Hz,  $J_{5b,4b}$  = 3.8 Hz, H-5b), 5.73 (1H, dd,  $J_{3b,4b}$  = 5.8 Hz,  $J_{3b,2b}$  = 1.9 Hz, H-3b), 5.51-5.46 (2H, m, H-1b, H-2b), 4.91 (1H, dd, H-4b), 4.89 (1H, d,  $J_{1a,2a}$  = 1.6 Hz, H-1a), 4.80 (1H, dd,  $J_{6b,6'b}$  = 11.9 Hz, H-6b), 4.74 (1H, dd, H-6'b), 4.40 (1H, dd,  $J_{6a,6'a}$  = 11.9 Hz,  $J_{6a,5a}$  = 4.7 Hz, H-6a), 4.36 (1H, dd,  $J_{6'a,5a}$  = 2.8 Hz, H-6'a), 4.07 (1H, dd,  $J_{2a,3a}$  = 3.3 Hz, H-2a), 3.95 (1H, dd,  $J_{3a,4a}$  = 9.2 Hz, H-3a), 3.86 (1H, dd,  $J_{4a,5a}$  = 9.7 Hz, H-4a), 3.77 (1H, ddd, H-5a), 3.66 (1H, dt,  $^2J$  = 9.7 Hz,  $^3J$  = 6.8 Hz, OCH<sub>2</sub>), 3.42 (1H, dt,  $^3J$  = 6.6 Hz, OCH<sub>2</sub>), 2.11 (3H, s, COCH<sub>3</sub>) 1.56 (2H, m, OCH<sub>2</sub>CH<sub>2</sub>), 1.36–1.20 (10H, m, (CH<sub>2</sub>)<sub>5</sub>CH<sub>3</sub>), 0.87 (3H, t,  $^3J$  = 6.9, CH<sub>2</sub>CH<sub>3</sub>). <sup>13</sup>C NMR (CDCl<sub>3</sub>): δ<sub>C</sub> 171.5, 166.4, 166.2, 165.8, 165.7 (CO), 134.8, 133.8, 133.5, 133.3, 130.1, 130.1, 130.0, 129.9, 129.6, 129.5, 128.9, 128.7, 128.6, 128.6, 128.5, 128.0 (C<sub>Ar</sub>), 104.4 (C-1b), 99.5 (C-1a), 83.1 (C-2b), 81.2 (C-4b), 78.8 (C-3a), 77.0 (C-3b), 70.4 (C-5a), 70.3 (C-5b), 68.6 (C-2a), 68.1 (OCH<sub>2</sub>), 66.0 (C-4a), 63.8 (C-6a), 63.2 (C-6b), 32.0, 29.5, 29.5, 29.3, 26.2, 22.8 [(CH<sub>2</sub>)<sub>6</sub>CH<sub>3</sub>],

21.1 (COCH<sub>3</sub>), 14.2 (CH<sub>2</sub>CH<sub>3</sub>). HRMS (ESI/Q-TOF) m/z: [M+Na]<sup>+</sup> Calcd for C<sub>50</sub>H<sub>56</sub>O<sub>16</sub>Na 935.34606 ; Found 935.3467.

**9a:** <sup>1</sup>H NMR (CDCl<sub>3</sub>): δ<sub>H</sub> 7.97-7.91 (2H, m, H<sub>Ar</sub>), 7.91- 7.83 (4H, m, H<sub>Ar</sub>), 7.75-7.70 (2H, m, H<sub>Ar</sub>), 7.60- 7.54 (1H, m, H<sub>Ar</sub>), 7.53-7.46 (2H, m, H<sub>Ar</sub>), 7.42-7.36 (5H, m, H<sub>Ar</sub>), 7.36-7.28 (4H, m, H<sub>Ar</sub>), 6.36 (1H, d, *J*<sub>1b,2b</sub> = 4.4 Hz, H-1b), 5.51 (1H, dd, *J*<sub>3b,4b</sub> = 4.2 Hz, *J*<sub>3b,2b</sub> = 1.5 Hz, H-3b), 5.28 (1H, ddd, *J*<sub>5b,4b</sub> = 7.6 Hz, *J*<sub>5b,6'b</sub> = 5.0 Hz, *J*<sub>5b,6b</sub> = 3.9 Hz, H-5b), 5.23 (1H, dd, H-2b), 4.71 (1H, d, *J*<sub>1a,2a</sub> = 1.7 Hz, H-1a), 4.62 (1H, dd, H-4b), 4.51 (1H, dd, *J*<sub>6b,6'b</sub> = 12.4 Hz, H-6b), 4.42 (1H, dd, *J*<sub>6a,6'a</sub> = 12.1 Hz, *J*<sub>6a,5a</sub> = 4.6 Hz, H-6a), 4.33 (1H, dd, H-6'b), 4.27 (1H, dd, *J*<sub>6'a,5a</sub> = 2.1 Hz, H-6'a), 3.90 (1H, dd, *J*<sub>3a,4a</sub> = 8.8 Hz, *J*<sub>3a,2a</sub> = 3.3 Hz, H-3a), 3.79-3.67 (2H, m, H-4a, H5a), 3.63-3.55 (2H, m, H-2a, OCH<sub>2</sub>), 3.33 (1H, dt, <sup>2</sup>*J* = 9.7 Hz, <sup>3</sup>*J* = 6.6 Hz, OCH<sub>2</sub>), 2.87 (1H, d, *J*<sub>OH,4a</sub> = 2.8 Hz, OH), 2.17 (1H, d, *J*<sub>OH,2a</sub> = 3.2 Hz, OH), 2.09 (3H, s, COCH<sub>3</sub>), 1.56-1.46 (2H, m, OCH<sub>2</sub>CH<sub>2</sub>), 1.31-1.23 (10H, m, (CH<sub>2</sub>)<sub>5</sub>CH<sub>3</sub>), 0.88 (3H, t, <sup>3</sup>*J* = 6.9 Hz, (CH<sub>2</sub>)<sub>5</sub>CH<sub>3</sub>). <sup>13</sup>C NMR (CDCl<sub>3</sub>): δ<sub>C</sub> 171.7 (COCH<sub>3</sub>), 165.8, 165.4, 165.3 (COPh), 136.6, 133.8, 133.2, 133.2, 130.1, 130.0, 129.9, 129.8, 129.6, 128.7, 128.6, 128.5, 128.4, 126.2 (C<sub>Ar</sub>), 123.9 (PhCO<sub>3</sub>), 105.1 (C-1b), 99.3 (C-1a), 86.0 (C-2b), 83.1 (C-4b), 76.7 (C-3b), 74.5 (C-3a), 71.2 (C-5b), 70.6 (C-5a), 70.2 (C-2a), 68.0 (OCH<sub>2</sub>), 65.6 (C-4a), 63.7 (C-6a), 62.9 (C-6b), 31.9, 29.4, 29.4, 29.4, 26.2, 22.8 [(CH<sub>2</sub>)<sub>6</sub>CH<sub>3</sub>], 21.0 (COCH<sub>3</sub>), 14.2 [(CH<sub>2</sub>)<sub>6</sub>CH<sub>3</sub>]. HRMS (ESI/Q-TOF) m/z: [M+Na]<sup>+</sup> Calcd for C<sub>50</sub>H<sub>56</sub>O<sub>16</sub>Na 935.34606 ; Found 935.3465.

**Thiotolyl 2,3,5,6-*O*-benzoyl-β-D-galactofuranosyl-(1→3)-6-*O*-acetyl-α-D-mannopyranoside (8b):** Synthesized according to general procedure starting from thiotolyl 6-*O*-Ac-mannopyranoside **4b** (50 mg, 0.15 mmol), galactofuranosyl bromide **1a** (200 mg, 0.30 mmol), 2-DPBA (34 mg, 0.15 mmol) and Ag<sub>2</sub>O (35 mg, 0.15 mmol). Reaction reached completion after 2 h. Purification by column chromatography on silica gel afforded **8b** (131 mg, 96%) as a white solid.

Alternative protocol using **4b** (50 mg, 0.15 mmol), **1a** (200 mg, 0.30 mmol), 2-DPBA (34 mg, 0.14 mmol) and AgOTf (39 mg, 0.15 mmol) as promoter gave **8b** (94 mg, 69%).

**8b**:  $[\alpha]_D^{20} + 60$  (c 1.15, CH<sub>2</sub>Cl<sub>2</sub>). <sup>1</sup>H NMR (CDCl<sub>3</sub>): δ<sub>H</sub> 8.11 (2H, dd, <sup>3</sup>J = 8.4 Hz, <sup>4</sup>J = 1.3 Hz, H<sub>Ar</sub>), 8.02 (4H, dd, <sup>3</sup>J = 8.4 Hz, <sup>4</sup>J = 1.3 Hz, H<sub>Ar</sub>), 7.93 (2H, dd, <sup>3</sup>J = 8.4 Hz, <sup>4</sup>J = 1.3 Hz, H<sub>Ar</sub>), 7.61-7.51 (4H, m, H<sub>Ar</sub>), 7.46-7.31 (10H, m, H<sub>Ar</sub>), 7.12 (2H, d, <sup>3</sup>J = 8.0 Hz, H<sub>Tol</sub>), 5.94 (1H, td,  $J_{5b,6b} = 5.3$  Hz,  $J_{5b,4b} = 3.6$  Hz, H-5b), 5.73 (1H, dd,  $J_{3b,4b} = 5.9$  Hz,  $J_{3b,2b} = 2.2$  Hz, H-3b), 5.54 (1H, d,  $J_{1a,2a} = 1.4$  Hz, H-1a), 5.48 (1H, d,  $J_{1b,2b} = 0.8$  Hz, H-1b), 5.48 (1H, dd, H-2b), 4.92 (1H, dd, H-4b), 4.80 (2H, d, H-6b), 4.41 (1H, dd,  $J_{6a,6'a} = 12.1$  Hz,  $J_{6a,5a} = 5.9$  Hz, H-6a), 4.37-4.30 (3H, m, H-2a, H-5a, H-6'a), 3.98-3.88 (2H, m, H-3a, H-4a), 3.19 (1H, d,  $J_{OH,4a} = 3.8$  Hz, OH), 2.33 (3H, s, CH<sub>3</sub>), 2.07 (3H, s, COCH<sub>3</sub>). <sup>13</sup>C NMR (CDCl<sub>3</sub>): δ<sub>C</sub> 171.4, 166.5, 166.2, 165.8, 165.7 (CO), 138.1, 133.9, 133.6, 133.4, 132.5, 130.1, 130.0, 129.6, 129.5, 128.9, 128.7, 128.6 (C<sub>Ar</sub>), 104.1 (C-1b), 87.8 (C-1a), 83.2 (C-2b), 81.4 (C-4b), 78.3 (C-3a), 77.0 (C-3b), 71.4 (C-5a), 70.2 (C-5b), 69.6 (C-2a), 66.4 (C-4a), 63.8 (C-6a), 63.2 (C-6b), 21.3 (CH<sub>3</sub>), 21.0 (COCH<sub>3</sub>). HRMS (ESI/Q-TOF) m/z: [M+Na]<sup>+</sup> Calcd for C<sub>49</sub>H<sub>46</sub>O<sub>15</sub>SNa 929.24496 ; Found 929.2455.

***p*-Nitrophenyl (2, 3, 5, 6-*O*-tetrabenzoyl-β-D-galactofuranosyl)-(1→3)-6-*O*-acetyl-β-D-galactopyranoside (17)**: Synthesized according to general procedure starting from *p*NP 6-*O*-Ac-galactopyranoside **12** (52 mg, 0.15 mmol), galactofuranosyl bromide **1a** (200 mg, 0.30 mmol), 2-DPBA (34 mg, 0.15 mmol) and Ag<sub>2</sub>O (35 mg, 0.15 mmol). Reaction reached completion after 2 h. Purification by column chromatography on silica gel afforded **17** (121 mg, 86%) as a white solid.  $[\alpha]_D^{20} - 26.5$  (c 1, CHCl<sub>3</sub>). <sup>1</sup>H NMR (CDCl<sub>3</sub>): δ<sub>H</sub> 8.20 (2H, d,  $J = 9.4$  Hz, H<sub>*p*NP</sub>), 8.12-8.06 (2H, m, H<sub>Bz</sub>), 8.04-7.97 (4H, m, H<sub>Bz</sub>), 7.95-7.89 (2H, m, H<sub>Bz</sub>), 7.62-7.51 (3H, m, H<sub>Bz</sub>), 7.47-7.29 (9H, m, H<sub>Bz</sub>), 7.12 (2H, d, H<sub>*p*NP</sub>), 5.97 (1H, ddd,  $J_{5b,6'b} = 6.3$  Hz,  $J_{5b,6'b} = 5.0$  Hz,  $J_{5b,4b} = 3.8$  Hz, H-5b), 5.75 (1H, dd,  $J_{3b,4b} = 5.8$  Hz,  $J_{3b,2b} = 2.2$  Hz, H-3b), 5.69 (1H, d,  $J_{1b,2b} = 0.9$  Hz H-1b), 5.56 (1H, dd, H-2b), 4.99 (1H, d,  $J_{1a,2a} = 7.8$  Hz, H-1a), 4.83 (1H, dd,

1  
2  
3  $J_{6b,6'b} = 11.8$ , H-6b), 4.80 (1H, dd, H-4b), 4.71 (1H, dd, H-6'b), 4.34 (1H, dd,  $J_{6a,6'a} = 11.7$  Hz,  
4  
5  $J_{6a,5a} = 7.6$  Hz, H-6a), 4.24 (1H, dd,  $J_{6'a,5a} = 4.8$  Hz, H-6'a), 4.22 (1H, dd,  $J_{2a,3a} = 9.4$  Hz, H-  
6  
7 2a), 4.10 (1H, dd,  $J_{4a,3a} = 3.4$  Hz,  $J_{4a,5a} = 1.1$  Hz, H-4a), 3.86 (1H, ddd, H-5a), 3.79 (1H, dd,  
8  
9 H-3a), 2.08 (3H, s, CH<sub>3</sub>). <sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta_C$  170.8, 166.3, 166.3, 165.8, 165.7 (COPh),  
10  
11 161.8 (C<sub>pNP</sub>), 142.9 (C<sub>pNP</sub>), 133.9, 133.6, 133.5, 130.1, 130.0, 129.9, 129.4, 128.7, 128.6,  
12  
13 128.5 (C<sub>Bz</sub>), 125.7 (C<sub>pNP</sub>), 116.7 (C<sub>pNP</sub>), 107.9 (C-1b), 100.2 (C-1a), 83.1 (C-2b), 81.6 (C-4b),  
14  
15 80.7 (C-3a), 77.0 (C-3b), 72.9 (C-5a), 70.2 (C-2a, C-5b), 68.5 (C-4a), 63.0 (C-6a, C-6b), 20.9  
16  
17 (CH<sub>3</sub>). HRMS (ESI/Q-TOF) m/z: [M+Na]<sup>+</sup> Calcd for C<sub>48</sub>H<sub>43</sub>NO<sub>18</sub>Na 944.23723 ; Found  
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19 944.2368.  
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24 ***p*-Nitrophenyl (2, 3, 5, 6-tetra-*O*-benzoyl- $\beta$ -D-galactofuranosyl)-(1 $\rightarrow$ 3)-2-acetamido-6-*O*-**  
25  
26 **acetyl-2-deoxy- $\beta$ -D-galactopyranoside (18):** Synthesized according to general procedure  
27  
28 starting from *p*NP 6-*O*-acetyl- $\beta$ -D-*N*-acetylgalactosamine **14** (58 mg, 0.15 mmol),  
29  
30 galactofuranosyl bromide **1a** (200 mg, 0.30 mmol), 2-DPBA (34 mg, 0.15 mmol) and Ag<sub>2</sub>O  
31  
32 (35 mg, 0.15 mmol). Reaction reached completion after 2 h. Purification by column  
33  
34 chromatography on silica gel afforded **18** (14 mg, 10%) as a light brown solid. Further elution  
35  
36 allowed isolating the corresponding orthoester **18'** (54 mg, 37%).  
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38  
39

40  
41 Alternative protocol using **14** (58 mg, 0.15 mmol), **1a** (200 mg, 0.30 mmol), 2-DPBA (34 mg,  
42  
43 0.15 mmol) and AgOTf (39 mg, 0.15 mmol) as promoter gave **18** (80 mg, 55%) in mixture  
44  
45 with the *p*NP  $\beta$ -D-Galf-(1 $\rightarrow$ 4)-D-GalNHAc regioisomer (ratio 4:1).  
46  
47

48 **18:** <sup>1</sup>H NMR (CD<sub>3</sub>OD+CDCl<sub>3</sub>):  $\delta_H$  8.15 (2H, d, <sup>3</sup>*J* = 9.2 Hz, H<sub>*p*NP</sub>), 8.10-8.03 (2H, m, H<sub>Bz</sub>),  
49  
50 8.02-7.93 (4H, m, H<sub>Bz</sub>), 7.87-7.81 (2H, m, H<sub>Bz</sub>), 7.57-7.50 (3H, m, H<sub>Bz</sub>), 7.44-7.27 (9H, m,  
51  
52 H<sub>Bz</sub>), 7.06 (2H, d, H<sub>*p*NP</sub>), 5.95 (1H, ddd,  $J_{5b,6'b} = 6.7$  Hz,  $J_{5b,6b} = 4.4$  Hz,  $J_{5b,4b} = 4.0$  Hz, H-5b),  
53  
54 5.68 (1H, d,  $J_{1a,2a} = 8.2$  Hz, H-1a), 5.66 (1H, dd,  $J_{3b,4b} = 5.4$  Hz,  $J_{3b,2b} = 1.9$  Hz, H-3b), 5.43  
55  
56 (1H, d, H-2b), 5.37 (1H, s, H-1b), 4.84 (1H, dd, H-4b), 4.76 (1H, dd,  $J_{6b,6'b} = 11.8$  Hz, H-6b),  
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4.67 (1H, dd, H-6'b), 4.49 (1H, dd,  $J_{3a,2a} = 10.8$  Hz,  $J_{3a,4a} = 3.3$  Hz, H-3a), 4.30 (1H, dd,  $J_{6a,6'a} = 11.7$  Hz,  $J_{6a,5a} = 7.9$  Hz, H-6a), 4.16 (1H, dd,  $J_{6'a,5a} = 4.5$  Hz, H-6'a), 4.09 (1H, d, H-4a), 3.89 (1H, dd, H-5a), 3.80 (1H, dd, H-2a), 2.03 (3H, s, CH<sub>3</sub>), 1.93 (3H, s, CH<sub>3</sub>). <sup>13</sup>C NMR (CD<sub>3</sub>OD+CDCl<sub>3</sub>): δ<sub>C</sub> 172.4, 171.0 (COCH<sub>3</sub>), 166.4, 166.2, 165.9, 165.9 (COPh), 161.9 (C<sub>pNP</sub>), 142.8 (C<sub>pNP</sub>), 133.9, 133.6, 133.5, 130.1, 130.0, 129.9, 129.8, 129.4, 128.7, 128.6, 128.5 (C<sub>Bz</sub>), 125.7 (C<sub>pNP</sub>), 116.7 (C<sub>pNP</sub>), 107.9 (C-1b), 97.0 (C-1a), 83.0 (C-2b), 81.2 (C-4b), 77.4 (C-3b), 76.8 (C-3a), 72.8 (C-5a), 70.3 (C-5b), 68.2 (C-4a), 63.3 (C-6a, C-6b), 53.4 (C-2a), 23.3, 20.8 (COCH<sub>3</sub>). HRMS (ESI/Q-TOF) m/z: [M+Na]<sup>+</sup> Calcd for C<sub>50</sub>H<sub>46</sub>N<sub>2</sub>O<sub>18</sub>Na 985.26378 ; Found 985.2646.

**18'**: <sup>1</sup>H NMR (CDCl<sub>3</sub>): δ<sub>H</sub> 8.14 (2H, d, <sup>3</sup>J = 9.3 Hz, H<sub>pNP</sub>), 7.93-7.88 (2H, m, H<sub>Bz</sub>), 7.86-7.80 (4H, m, H<sub>Bz</sub>), 7.73-7.68 (2H, m, H<sub>Bz</sub>), 7.59-7.46 (3H, m, H<sub>Bz</sub>), 7.43-7.27 (9H, m, H<sub>Bz</sub>), 7.04 (2H, d, H<sub>pNP</sub>), 6.21 (1H, d,  $J_{1b,2b} = 4.3$  Hz, H-1b), 5.84 (1H, d,  $J_{1a,2a} = 8.3$  Hz, H-1a), 5.76 (1H, d,  $J = 7.0$ , NH), 5.40 (1H, dd,  $J_{3b,4b} = 5.4$  Hz,  $J_{3b,2b} = 1.8$  Hz, H-3b), 5.19 (1H, ddd,  $J_{5b,4b} = 7.0$  Hz,  $J_{5b,6'b} = 5.3$  Hz,  $J_{5b,6b} = 3.7$  Hz, H-5b), 5.10 (1H, dd, H-2b), 4.63 (1H, dd,  $J_{3a,2a} = 10.6$  Hz,  $J_{3a,4a} = 3.0$  Hz, H-3a), 4.51 (1H, dd, H-4b), 4.42 (1H, dd,  $J_{6b,6'b} = 12.4$  Hz, H-6b), 4.32 (2H, d,  $J_{6a,5a} = 6.2$  Hz, H-6a), 4.24 (1H, dd, H-6'b), 4.10 (1H, t,  $J_{4a,3a} = J_{4a,OH} = 3.0$  Hz, H-4a), 3.92 (1H, t, H-5a), 3.54 (1H, ddd,  $J_{2a,NH} = 7.0$  Hz, H-2a), 2.46 (1H, d, OH), 2.07 (3H, s, OCOCH<sub>3</sub>), 1.98 (3H, s, NHCOCH<sub>3</sub>). <sup>13</sup>C NMR (CDCl<sub>3</sub>): δ<sub>C</sub> 171.9 (NHCO), 170.9 (OCO), 165.8, 165.4 (COPh), 161.9 (C<sub>pNP</sub>), 142.9 (C<sub>pNP</sub>), 137.4, 133.9, 133.3, 133.2, 130.2, 129.9, 129.8, 129.5, 128.7, 128.6, 128.5, 128.4, 126.2 (C<sub>Ar</sub>), 125.8 (C<sub>pNP</sub>), 124.8 (PhCO<sub>3</sub>), 116.7 (C<sub>pNP</sub>), 104.5 (C-1b), 96.6 (C-1a), 86.4 (C-2b), 81.2 (C-4b), 76.4 (C-3b), 72.7 (C-5a), 71.0 (C-5b), 70.0 (C-3a), 68.2 (C-4a), 63.2 (C-6a), 62.7 (C-6b), 53.9 (C-2a), 24.0 (NHCOCH<sub>3</sub>), 21.0 (OCOCH<sub>3</sub>). HRMS (ESI/Q-TOF) m/z: [M+Na]<sup>+</sup> Calcd for C<sub>50</sub>H<sub>46</sub>N<sub>2</sub>O<sub>18</sub>Na 985.26378 ; Found 985.2632.

***p*-Nitrophenyl (2, 3, 5, 6-tetra-*O*-benzoyl-β-D-galactofuranosyl)-(1→3)-α-L-rhamnopyranoside (19)**: Synthesized according to general procedure starting from *p*NP α-L-

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3 rhamnopyranoside **15** (43 mg, 0.15 mmol), galactofuranosyl bromide **1a** (200 mg, 0.30  
4 mmol), 2-DPBA (34 mg, 0.15 mmol) and Ag<sub>2</sub>O (35 mg, 0.15 mmol). Reaction reached  
5 completion after 2 h. Purification by column chromatography on silica gel afforded **19** (58  
6 mg, 45%) as a light brown foam. Further elution allowed isolating the corresponding  
7 orthoester **19'** (43 mg, 33%).  
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14 Alternative protocol using **15** (43 mg, 0.15 mmol), **1a** (200 mg, 0.30 mmol), 2-DPBA (34 mg,  
15 0.15 mmol) and AgOTf (39 mg, 0.15 mmol) as promoter gave **19** (77 mg, 59%) in mixture  
16 with the *p*NP β-D-Galf-(1→4)-L-Rhap regioisomer (ratio 9:1).  
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22 **19**: [ $\alpha$ ]<sub>D</sub><sup>20</sup> -90 (c 0.8, CHCl<sub>3</sub>). <sup>1</sup>H NMR (CDCl<sub>3</sub>): δ<sub>H</sub> 8.20 (2H, d, <sup>3</sup>*J* = 9.3 Hz, H<sub>*p*NP</sub>), 8.10 (2H,  
23 dd, <sup>3</sup>*J* = 8.4 Hz, <sup>4</sup>*J* = 1.3 Hz, H<sub>Bz</sub>), 8.05-7.90 (6H, m, H<sub>Bz</sub>), 7.62-7.48 (4H, m, H<sub>Bz</sub>), 7.47-7.40  
24 (2H, m, H<sub>Bz</sub>), 7.39-7.31 (6H, m, H<sub>Bz</sub>), 7.11 (2H, d, H<sub>*p*NP</sub>), 5.94 (1H, ddd, *J*<sub>5b,6'b</sub> = 6.2 Hz, *J*<sub>5b,6b</sub>  
25 = 5.6 Hz, *J*<sub>5b,4b</sub> = 3.5 Hz, H-5b), 5.78 (1H, dd, *J*<sub>3b,4b</sub> = 6.2 Hz, *J*<sub>3b,2b</sub> = 2.7 Hz, H-3b), 5.65 (1H,  
26 d, *J*<sub>1b,2b</sub> = 1.1 Hz, H-1b), 5.54 (1H, dd, H-2b), 5.49 (1H, d, *J*<sub>1a,2a</sub> = 1.9 Hz, H-1a), 4.87 (1H, dd,  
27 *J*<sub>6b,6'b</sub> = 11.8 Hz, H-6b), 4.84 (1H, dd, H-4b), 4.69 (1H, dd, H-6'b), 4.26 (1H, dd, *J*<sub>2a,3a</sub> = 3.4  
28 Hz, H-2a), 4.05 (1H, dd, *J*<sub>3a,4a</sub> = 9.3 Hz, H-3a), 3.81 (1H, t, *J*<sub>4a,3a</sub> = *J*<sub>4a,5a</sub> = 9.3 Hz, H-4a), 3.71  
29 (1H, qd, *J*<sub>5a,CH<sub>3</sub></sub> = 6.2 Hz, H-5a), 3.12 (1H, s, OH), 2.86 (1H, s, OH), 1.31 (3H, d, CH<sub>3</sub>). <sup>13</sup>C  
30 NMR (CDCl<sub>3</sub>): δ<sub>C</sub> 166.6, 166.5, 165.8, 165.7 (COPh), 161.0 (C<sub>*p*NP</sub>), 142.7 (C<sub>*p*NP</sub>), 134.9,  
31 134.0, 133.9, 133.7, 133.5, 130.2, 130.1, 130.0, 129.9, 129.4, 128.7, 128.7, 128.7, 128.6,  
32 128.5, 128.1 (CPh), 125.9 (C<sub>*p*NP</sub>), 116.4 (C<sub>*p*NP</sub>), 108.6 (C-1b), 97.8 (C-1a), 83.9 (C-2b), 81.0  
33 (C-3a,C-4b), 76.7 (C-3b), 71.3 (C-4a), 70.3 (C-2a), 70.1 (C-5b), 69.4 (C-5a), 63.0 (C-6b),  
34 17.8. (CH<sub>3</sub>). HRMS (ESI/Q-TOF) *m/z*: [M+Na]<sup>+</sup> Calcd for C<sub>46</sub>H<sub>41</sub>NO<sub>16</sub>Na 886.23175 ; Found  
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54 **19'**: <sup>1</sup>H NMR (CDCl<sub>3</sub>): δ<sub>H</sub> 8.14 (2H, d, <sup>3</sup>*J* = 9.2 Hz, H<sub>*p*NP</sub>), 7.96-7.91 (2H, m, H<sub>Bz</sub>), 7.89-7.84  
55 (4H, m, H<sub>Bz</sub>), 7.79-7.73 (2H, m, H<sub>Bz</sub>), 7.61-7.55 (1H, m, H<sub>Bz</sub>), 7.54-7.48 (2H, m, H<sub>Bz</sub>), 7.44-  
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7.30 (9H, m, H<sub>Bz</sub>), 7.05 (2H, d, H<sub>pNP</sub>), 6.36 (1H, d,  $J_{1b,2b} = 4.2$  Hz, H-1b), 5.53-5.49 (2H, m, H-1a, H-3b), 5.28-5.23 (1H, m, H-5b), 5.25 (1H, dd,  $J_{2b,3b} = 1.5$  Hz, H-2b), 4.63 (1H, dd,  $J_{4b,3b} = 7.5$  Hz,  $J_{4b,5b} = 4.6$  Hz, H-4b), 4.51 (1H, dd,  $J_{6b,6'b} = 12.5$  Hz,  $J_{6b,5b} = 3.7$  Hz, H-6b), 4.32 (1H, dd,  $J_{6'b,5b} = 5.3$  Hz, H-6'b), 4.06 (1H, t,  $J_{OH,2a} = J_{2a,3a} = 3.1$  Hz, H-2a), 4.00 (1H, dd,  $J_{3a,4a} = 8.8$  Hz, H-3a), 3.69-3.57 (2H, m, H-4a, H-5a), 2.59 (1H, d, OH), 2.52 (1H, d,  $J_{OH,4a} = 2.4$  Hz, OH), 1.26 (3H, d,  $J_{CH_3,5a} = 1.0$  Hz, CH<sub>3</sub>). <sup>13</sup>C NMR (CDCl<sub>3</sub>): δ<sub>C</sub> 165.8, 165.5, 165.4 (COPh), 161.0 (C<sub>pNP</sub>), 142.7 (C<sub>pNP</sub>), 136.6, 134.0, 133.3, 130.1, 129.9, 129.8, 128.7, 128.6, 128.5, 128.4, 126.2 (C<sub>Bz</sub>), 125.9 (C<sub>pNP</sub>), 124.3 (PhCO<sub>3</sub>), 116.3 (C<sub>pNP</sub>), 104.8 (C-1b), 97.7 (C-1a), 86.2 (C-2b), 82.4 (C-4b), 76.6 (C-3b), 74.4 (C-3a), 71.1 (C-5b), 70.5 (C-4a), 70.0 (C-5a), 69.7 (C-2a), 62.9 (C-6b), 17.8 (CH<sub>3</sub>). HRMS (ESI/Q-TOF) m/z: [M+Na]<sup>+</sup> Calcd for C<sub>46</sub>H<sub>41</sub>NO<sub>16</sub>Na 886.23175 ; Found 886.2318.

**4-Nitrophenyl 2,3,5,6-tetra-O-benzoyl-β-D-galactofuranosyl-(1→6)-β-D-galactofuranoside (20):** Synthesized according to general procedure starting from pNP β-D-galactofuranoside **16** (20 mg, 67 μmol), galactofuranosyl bromide **1a** (88 mg, 0.13 mmol), 2-DPBA (15 mg, 67 μmol) and Ag<sub>2</sub>O (15 mg, 67 μmol). Reaction reached completion after 2 h. Purification by column chromatography on silica gel afforded **20** (7 mg, 12%) as a white solid. Further elution allowed isolating the corresponding orthoester **20'** (35 mg, 56%).

Alternative protocol using **16** (20 mg, 67 μmol), **1a** (88 mg, 0.13 mmol), 2-DPBA (15 mg, 65 μmol) and AgOTf (16 mg, 65 μmol) as promoter yielded a 7:1 inseparable mixture of **20** and pNP β-D-Galf-(1→5)-D-Galf (51% overall yield, 30 mg, 34 μmol).

**20:** <sup>1</sup>H NMR (CD<sub>3</sub>OD): δ<sub>H</sub> 8.11-8.06 (4H, m, H<sub>pNP</sub>, H<sub>Bz</sub>), 7.93-7.81 (6H, 3 dd, <sup>3</sup>J = 8.3 Hz, <sup>4</sup>J = 1.2 Hz, H<sub>Bz</sub>), 7.66 (1H, td, <sup>3</sup>J = 7.3 Hz, <sup>4</sup>J = 1.2 Hz, H<sub>Bz</sub>), 7.56-7.22 (11H, m, H<sub>Bz</sub>), 7.03 (2H, d, <sup>3</sup>J = 9.3 Hz, H<sub>pNP</sub>), 5.77 (1H, ddd,  $J_{5b,6b} = 7.1$  Hz,  $J_{5b,4b} = J_{5b,6'b} = 3.5$  Hz, H-5b), 5.61 (1H, d,  $J_{1a,2a} = 2.4$  Hz, H-1a), 5.51 (1H, d,  $J_{3b,4b} = 4.7$  Hz, H-3b), 5.41 (1H, s, H-1b), 5.39 (1H,

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3 s, H-2b), 4.60 (1H, dd,  $J_{6b,6'b} = 12.1$  Hz, H-6b), 4.42-4.36 (2H, m, H-6'b, H-4b), 4.34 (1H, dd,  
4  $J_{4a,3a} = 7.1$  Hz,  $J_{4a,5a} = 1.9$  Hz, H-4a), 4.27 (1H, dd,  $J_{3a,2a} = 4.9$  Hz, H-3a), 4.15 (1H, dd, H-2a),  
5 4.04 (1H, ddd,  $J_{5a,6'a} = 8.6$  Hz,  $J_{5a,6a} = 6.2$  Hz, H-5a), 3.92 (1H, app. t.,  $J_{6a,6'a} = 8.6$  Hz, H-6a),  
6 3.63 (1H, ddd, H-6'a).  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ )  $\delta_{\text{C}}$  167.6, 167.2, 167.1, 166.8 (COPh), 163.4 ( $\text{C}_{p\text{NP}}$ ),  
7 143.6 ( $\text{C}_{p\text{NP}}$ ), 134.9, 134.7, 134.6, 134.4, 131.1, 130.8, 130.7, 130.4, 130.3, 129.9, 129.7,  
8 129.5 ( $\text{C}_{\text{Bz}}$ ), 126.7, 117.4 ( $\text{C}_{p\text{NP}}$ ), 107.6 (C-1a), 106.3 (C-1b), 84.5 (C-4a), 83.4 (C-2a), 83.3  
9 (C-4b), 83.2 (C-2b), 79.0 (C-3b), 77.1 (C-3a), 71.7 (C-5b), 68.8 (C-5a), 67.4 (C-6a), 64.9 (C-  
10 6b).

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12 **20'**:  $^1\text{H}$  NMR ( $\text{CD}_3\text{OD}$ )  $\delta_{\text{H}}$  8.14 (2H, d,  $^3J = 9.3$  Hz,  $\text{H}_{p\text{NP}}$ ), 7.96, 7.83, 7.62 (8H, 3 d,  $^3J = 7.9$   
13 Hz,  $\text{H}_{\text{Ar}}$ ), 7.54-7.27 (12H, m,  $\text{H}_{\text{Ar}}$ ), 7.18 (2H, d,  $\text{H}_{p\text{NP}}$ ), 6.06 (1H, d,  $J_{1b,2b} = 4.2$  Hz, H-1b), 5.64  
14 (1H, d,  $J_{1a,2a} = 1.8$  Hz, H-1a), 5.51 (1H, d,  $J_{3b,4b} = 3.3$  Hz, H-3b), 5.43-5.38 (1H, m, H-5b),  
15 5.10 (1H, d, H-2b), 4.66-4.59 (2H, m, H-4b, H-6b), 4.43 (1H, dd,  $J_{6'b,6b} = 12.4$  Hz,  $J_{6'b,5b} = 5.5$   
16 Hz, H-6'b), 4.27 (1H, dd,  $J_{2a,3a} = 4.0$  Hz, H-2a), 4.18 (1H, dd,  $J_{3a,4a} = 6.4$  Hz, H-3a), 4.11 (1H,  
17 dd,  $J_{4a,5a} = 2.5$  Hz, H-4a), 3.86 (1H, app. td.,  $J_{5a,6a} = 6.8$  Hz, H-5a), 3.51 (1H, dd,  $J_{6a,6'a} = 9.4$   
18 Hz, H-6a), 3.40 (1H, dd, H-6'a).  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ )  $\delta_{\text{C}}$  167.1, 166.7, 166.7 (COPh), 163.2,  
19 143.5 ( $\text{C}_{p\text{NP}}$ ), 137.2, 134.7, 134.2, 130.7, 130.6, 130.5, 130.5, 130.3, 130.1, 129.6, 129.4,  
20 129.3, 129.1, 127.5 ( $\text{C}_{\text{Ar}}$ ), 126.5 ( $\text{C}_{p\text{NP}}$ ), 124.6 ( $\text{CO}_3\text{Ph}$ ), 117.5 ( $\text{C}_{p\text{NP}}$ ), 107.5 (C-1a), 106.5 (C-  
21 1b), 86.8 (C-2b), 85.2 (C-4a), 84.6 (C-4b), 83.3 (C-2a), 77.9, 77.8 (C-3b, C-3a), 72.7 (C-5b),  
22 69.2 (C-5a), 64.9 (C-6a), 64.0 (C-6b).

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50 0718).  
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3 **Supporting Information Available:** Details of the computational calculations and  $^1\text{H}$  and  
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5  $^{13}\text{C}$  NMR spectra of all new compounds. This material is available free of charge via the  
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7 Internet at <http://pubs.acs.org>.  
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