



HAL
open science

Phytochemicals Targeting Estrogen Receptors: Beneficial Rather Than Adverse Effects?

Sylvain Lecomte, Florence Demay, François Ferrière, Farzad Pakdel

► To cite this version:

Sylvain Lecomte, Florence Demay, François Ferrière, Farzad Pakdel. Phytochemicals Targeting Estrogen Receptors: Beneficial Rather Than Adverse Effects?. *International Journal of Molecular Sciences*, 2017, 18 (7), pp.1381. 10.3390/ijms18071381 . hal-01560283

HAL Id: hal-01560283

<https://univ-rennes.hal.science/hal-01560283v1>

Submitted on 20 Jun 2018

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License



Review

Phytochemicals Targeting Estrogen Receptors: Beneficial Rather Than Adverse Effects?

Sylvain Lecomte, Florence Demay, François Ferrière and Farzad Pakdel *

Institut de Recherche en Santé-Environnement-Travail (IRSET), UMR 1085 Inserm, TREC Team, University of Rennes 1, 35000 Rennes, France; sylvain.lecomte@univ-rennes1.fr (S.L.); florence.demay@univ-rennes1.fr (F.D.); francois.ferriere@univ-rennes1.fr (F.F.)

* Correspondence: farzad.pakdel@univ-rennes1.fr; Tel.: +33-223-235-132; Fax: +33-223-235-055

Received: 4 May 2017; Accepted: 24 June 2017; Published: 28 June 2017

Abstract: In mammals, the effects of estrogen are mainly mediated by two different estrogen receptors, ER α and ER β . These proteins are members of the nuclear receptor family, characterized by distinct structural and functional domains, and participate in the regulation of different biological processes, including cell growth, survival and differentiation. The two estrogen receptor (ER) subtypes are generated from two distinct genes and have partially distinct expression patterns. Their activities are modulated differently by a range of natural and synthetic ligands. Some of these ligands show agonistic or antagonistic effects depending on ER subtype and are described as selective ER modulators (SERMs). Accordingly, a few phytochemicals, called phytoestrogens, which are synthesized from plants and vegetables, show low estrogenic activity or anti-estrogenic activity with potentially anti-proliferative effects that offer nutraceutical or pharmacological advantages. These compounds may be used as hormonal substitutes or as complements in breast cancer treatments. In this review, we discuss and summarize the *in vitro* and *in vivo* effects of certain phytoestrogens and their potential roles in the interaction with estrogen receptors.

Keywords: estrogen receptor; ligand; xenoestrogens; selective estrogen receptor modulators; transcription; epigenetic regulation; cell signaling; cancer

1. Introduction

Estrogens, such as 17 β -estradiol (E2), are steroid hormones derived from cholesterol by the successive action of steroidogenic enzymes. They are involved in multiple physiological processes by acting on various tissues. In particular, they participate in the establishment and regulation of the reproductive organs in both males and females, including the gonads or the mammary gland [1]. Furthermore, estrogens participate in many physiological processes in non-reproductive tissues, such as growth and remodeling of bone, differentiation and protection of the central nervous system, vasodilation of cardiovascular systems and lipid metabolism in the liver [1,2]. At the cellular level, E2 has multiple effects, including proliferation, differentiation and survival. E2 is a small, liposoluble molecule that passively enters the cell through the plasma membrane. E2 actions are mainly mediated by their binding to two estrogen receptors, ER α and ER β , which are localized in the cytoplasm and in the nucleus (Figure 1B). These receptors are members of the nuclear receptor superfamily, which also includes receptors for androgens, progesterone, glucocorticoids, thyroids, retinoid acids, and vitamin D, as well as more than twenty orphan receptors (Figure 1A) [3]. Many nuclear receptors are activated by specific ligands and generally act as transcription factors by binding to specific DNA sequences in the genome. Similar to the other nuclear receptors, ERs are modular proteins that consist of distinct structural and functional domains. The N-terminal domain contains the ligand-independent transactivation function (AF1). The central domain contains the conserved

zinc finger DNA-binding-domain, and the C-terminal domain contains the ligand-dependent transactivation function (AF2), as well as the ligand binding and dimerization sequences (Figure 1).

ER-mediated E2 actions at the transcriptional level of the estrogen-sensitive genes are called “genomic” E2 actions. Direct binding of ERs to the chromatin occurs at the estrogen-responsive-element (ERE) at target gene promoters. This induces the mobilization of the transcription coregulators needed to modify chromatin structure and thereby transcriptional regulation of specific gene (Figure 1B). This represents the classical pathway, but many E2-target genes do not contain the ERE. In this case, ERs modulate transcription through DNA-binding sites for Sp1 (stimulating protein 1) or AP1 (activator protein 1) transcription factors [4]. Furthermore, genome-wide studies performed by ChIP (chromatin immunoprecipitation) experiments in breast cancer cell lines specified that ER preferentially regulates its target genes by binding distal regulatory elements [5]. These distal regulatory sites can interact with the promoters of E2 target genes due to chromatin looping. This mechanism of transcriptional regulation represents more than 90% of E2-target genes [5]. Interestingly, these regulatory elements are capable of interacting with several promoters and other enhancers at the same time and are mainly contained in genomic areas called TADs (topologically associating domains) [6,7].

In contrast to the genomic action, the non-genomic actions of estrogens involve cytoplasmic signaling pathways (Figure 1B) and occur rapidly, on the order of seconds or minutes. This leads to the activation of several intracellular signaling pathways such as MAPK (mitogen activated protein kinase) or PI3K (phosphatidylinositide 3-kinase) [8]. Recent studies reported convergence or cross-talk between the genomic and non-genomic actions of ER, enabling a fine regulation of target genes and increasing the complexity of the estrogenic pathways [9–12].

ERs are generated from two different genes that are localized on chromosome 6, for ER α , and chromosome 14, for ER β , in humans. The utilization of different promoters and splicing processes results in multiple ER variants that can interfere with the transcriptional activity of wild type ERs in various cell types [13–16].

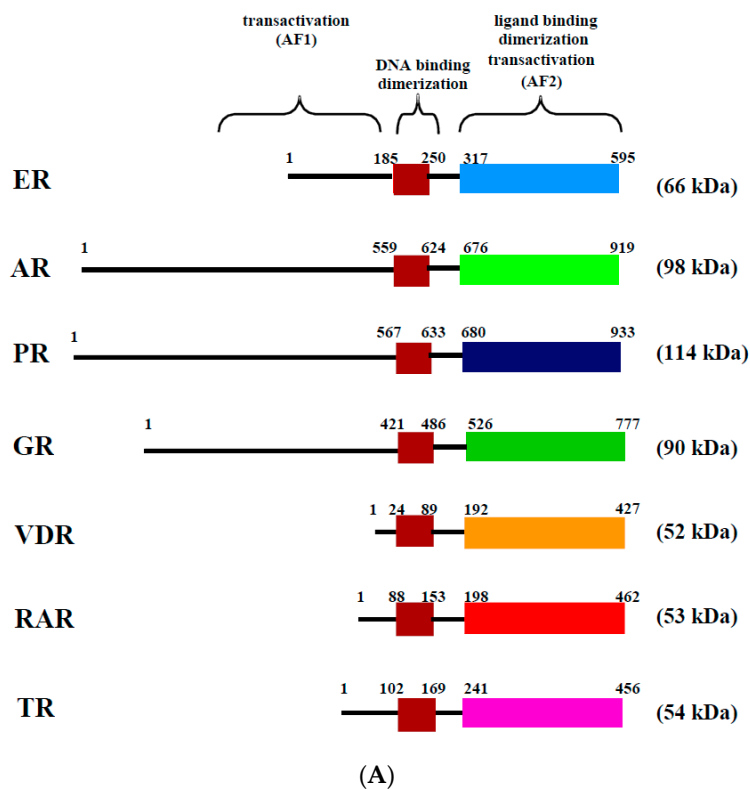


Figure 1. Cont.

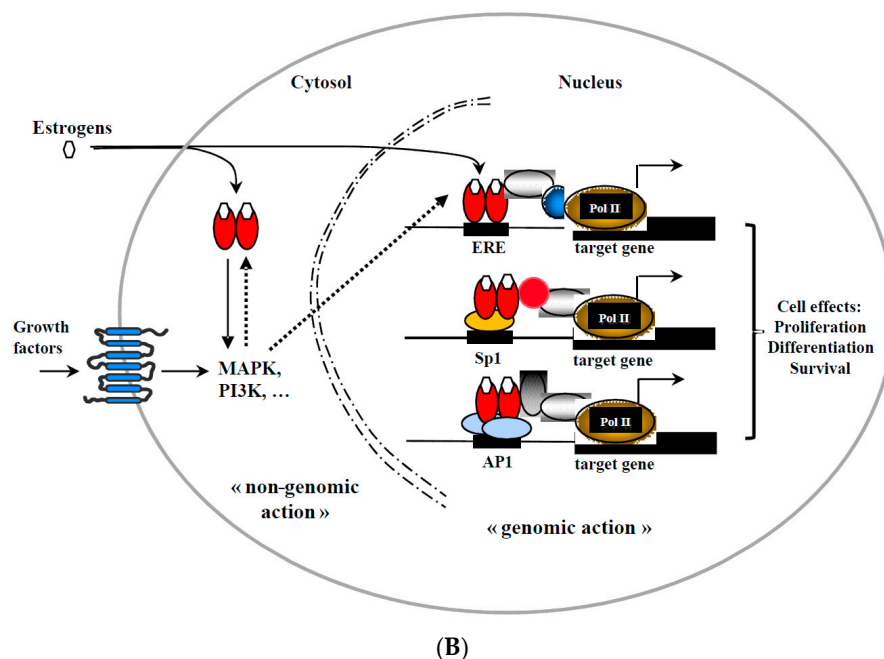


Figure 1. Structure and mechanisms of action of the estrogen receptor (ER). (A) The evolutionarily conserved domains of several nuclear receptors, including ER, AR (androgen receptor), PR (progesterone receptor), GR (glucocorticoid receptor), VDR (vitamin D receptor), RAR (retinoid acid receptor) and TR (thyroid receptor). Domains involved in DNA and ligand binding, as well as in dimerization, ligand-independent transactivation function (AF1) and ligand-dependent transactivation function (AF2) are shown. The number of amino acids for each domain is presented. The approximate molecular weight of each nuclear receptor is also indicated on the right side; (B) estradiol (E2) mediates multiple phenotypic changes in cells by binding to its receptor. E2 enter the cell through the lipid membranes and binds ER in the cytoplasm or the nucleus. ER mediates E2 effects through diverse transcriptional mechanisms. In the nucleus, the activated ER forms a dimer to tightly fix DNA directly at the ERE sites or indirectly at Sp1 or Ap1 sites. The activated ER is then able to recruit cofactors and RNA polymerase II (pol. II), which allows the transcription of target genes (ER genomic action). Furthermore, ERs can use rapid non-genomic action through the activation of intracellular kinases related or not to the growth factor signaling.

Many tissues express both ER subtypes, but with variable expression profiles. For instance, ER α is highly expressed in female reproductive tissues (ovary, womb, mammary gland). ER β is greatly expressed in ovaries but poorly expressed in the mammary gland. In men, ER α is strongly expressed in the testicle (Leydig cells and gubernaculum), whereas ER β is found in the prostate, germinal cells and epididymis. On the other hand, both receptors with variable expression levels are found in male and female, lung, hepatic, fat, osseous, and nervous tissues and endothelial cells [1,17–19]. Knockout in mice demonstrated crucial roles for both ER α and ER β during the development of reproductive tissues, gametogenesis, and neuronal growth and differentiation [1,20,21]. The appearance of ER seems to be under spatio-temporal control during development [1,19,20]. For instance, ER α expression has been found in the developing uterus as soon as the 15th day of fetal in mesenchymal cells, while it appears later in the epithelial cells and it rises during the neonatal period. In the cerebral cortex of rodents, ER α expression is greater in postnatal life and decreases substantially during puberty [20]. In the testis under development, ER α is expressed in the gubernaculum, a ligament which differentiates into the cremaster muscle involved in the final positioning of the testis within the scrotum. Its expression is strong between 17 to 20 dpc and barely detectable between 4 and 12 dpp, indicating a role of estrogens and ER α in the right positioning of the testis [19]. However, during mouse brain development, ER β distribution varies in different areas. ER β is found mostly in the midbrain and hypothalamus at E12.5,

and its expression increases at E15.5 and E16.5. Interestingly, ER β expression appears intensely and extensively throughout the brain, including in the cerebellum and striatum, at E18.5, whereas very few positive cells may be distinguished in the ventricular region [21].

Many natural and synthetic chemicals in the environment and in food have been reported with hormonal activity, particularly showing estrogenic potency [22]. These compounds are called endocrine disrupting chemicals (EDCs). A lot of EDCs are generated from human activities. For example, polycyclic aromatic hydrocarbons (PAH), such as polychlorinated dibenzo-*p*-dioxins and dibenzofurans, or polychlorinated biphenyls (PCBs), which are the most persistent and widespread in the environment. Bisphenol A, nonylphenol and ethinyl estradiol were also reported to be among the major environmental estrogens. A series of experimental and epidemiological studies over the past decades have suggested that these environmental contaminants can interfere with normal hormonal processes and induce deterioration of the reproduction function in males and females [22–27].

Furthermore, numerous natural molecules present in vegetables and plants possess estrogen- and antiestrogen-mimetic activities. These natural molecules are mainly phytoestrogen isoflavones, the most widely consumed. The most abundant isoflavones are genistein and daidzein, which are present in soybean, in particular, and also found in certain fruits, legumes and nuts [28]. Flavones, such as coumestans and lignans are other classes of phytoestrogens, which are also found in certain fruits and legumes [28]. Moreover, some mushrooms, mosses and fungi produce compounds with estrogenic activity. These compounds are called mycoestrogens, such as zearalenone [28]. In this review, we focus on these phytochemicals interacting with ERs and discuss their molecular actions and their potential effects on human health.

2. Structure and Sources of the Major Dietary Phytoestrogens

Against environmental stresses and aggressions, plants produce secondary metabolites belonging to the large family of polyphenols, which have many biological activities, such as antioxidant, antifungal and antibiotic properties. All of these compounds contain one or several aromatic rings with at least one hydroxyl group. Hydroxyl groups can be free, but most of the time they are engaged in another function with an ester, ether or a glycoside. Among these compounds, phytoestrogens have a structural similarity with 17 β -estradiol and could bind both ERs. Phytoestrogens are classified into six groups based on their chemical structures (Figure 2). In this review, we have chosen to present only the aglycone structure of a few phytoestrogens.

2.1. Flavonoids

Flavonoids, from *flavus* (yellow in Latin), are pigments of flowers and fruit, and represent the major group. They are formed by 2 aromatic rings bearing at least one hydroxyl group. The aromatic rings, called A and B, are connected by a carbon bridge consisting of three carbons combined with an oxygen to carbons of the A ring. Together, they formed a new 6-ring structure, called C [29] (Figure 2). Flavonoids could be divided into sub-classes depending on the position of the B ring at position 2 for flavones and derivatives and at position 3 for isoflavones and derivatives. Moreover, depending on hydroxylation degree and/or the position of the hydroxyl group, one can distinguish the flavan-3-ols, the flavanones and the flavonols [29].

Here, we have focused on flavones and isoflavones. Flavones are represented by compounds, such as apigenin, found in parsley or chamomile. Apigenin has a beneficial effect on human health [30]. The daily intake of flavones is very low and estimated between 0.3 and 1.6 mg/day [31]. Isoflavones such as genistein or daidzein are found in large quantities in soybean. The daily intake of isoflavones is low in Western countries (0.1–1.2 mg/day) and higher in Asian countries, where they consume more soy product (up to 47 mg/day) [29,32]. Approximately 30% of the population in Western countries and 60% of the population in Asian countries possess gut microbiota able to metabolize daidzein into the isoflavan equol, which shows a greater affinity for ERs than daidzein. Equol exists through two

enantiomers, the *R*-(+) equol and the *S*-(-) equol. This latter enantiomer is the natural compound produced by microbiota in human and rat [33].

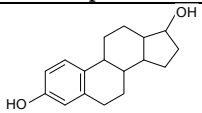
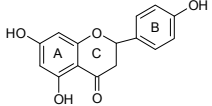
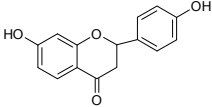
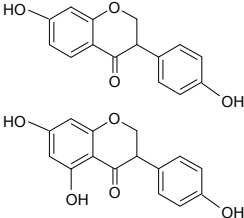
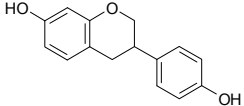
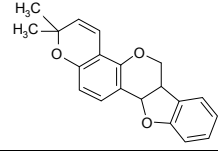
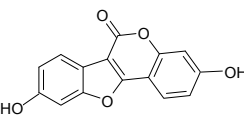
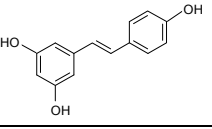
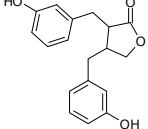
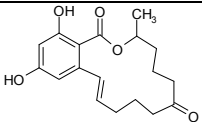
Class	Subclass	Compounds
Hormone	Natural ligand of estrogen receptor: 17 β -estradiol	
Flavonoids	Flavone Ex: apigenin	
	Flavanone Ex: liquiritigenin	
	Isoflavones Ex: daidzein Ex: genistein	
	Isoflavanes Ex: equol	
Pterocarpan	Ex: glyceollin I	
Coumestan	Ex: coumestrol	
Stilbene	Ex: resveratrol	
Enterolignan	Ex: enterolactone	
Mycotoxin	Ex: zearalenone	

Figure 2. Illustration of the chemical structures of different groups of phytoestrogens. Ex: Example.

2.2. Pterocarpan

Pterocarpan derive from isoflavones. Their structure is described as a benzo-pyrano-furano-benzene, where the B-rings are coupled to position 4-one [34]. Glyceollins, which correspond to prenylated 6a-hydroxy pterocarpan, are the main delegates of this family [35]. They belong to phytoalexin and are produced from daidzein via an enzymatic pathway, mainly in soybean, by the

action of a diversity of elicitors such as UV stress, bacterial or fungi infection [36]. These compounds have been known since the 1970s for their involvement in plant defense [37], but the ability of these compounds to act as phytoestrogens was only established in 2000 [38].

2.3. Coumestans

Coumestans are produced by oxidation of pterocarpan [39]. The structure of coumestans consists of a benzoxazole fused to a chromen-2-one [40]. The first, discovered in 1957 by Bickoff et al., and the best documented is coumestrol, which is abundant in alfalfa, soybean and clover [41]. This compound was shown to have a high affinity for both ERs and to induce a response of the same magnitude as that observed with E2 [42].

2.4. Stilbenes

Like glyceollins, stilbene belongs to the phytoalexins and participates in plant defense against injury, stress or infection [29]. Resveratrol, the main representative of the stilbene family, is abundant in grape and red wine, with a concentration up to 12 mg/L [43]. Although resveratrol was reported to interact with ERs, its agonist or antagonistic effects remain controversial [44,45].

2.5. Lignans

The two best-documented lignans are secoisolariciresinol and matairesinol [46]. Lignans are particularly abundant in flaxseed and sesame seed, and at minor concentrations in cereals, vegetables and fruits. The two major metabolites of lignans in human are also produced by gut microbiota. They present a weak estrogenic action and are called enterolactone and enterodiol [47].

2.6. Mycoestrogens

Another family of dietary estrogen, called mycoestrogens, is produced by fungi. In this family, the most documented is zearalenone and its derivatives. Zearalenone is produced by *Fusarium* and is found in poorly stored cereals. Zearalenone structure consists of resorcinol moiety fused with a 14-member macrocyclic lactone [48]. According to the European Safety Authority (EFSA), zearalenone is found in 15% of cereals consumed in Europe [49]. Zearalenone has adverse effects on human health, including reprotoxicity [50,51], genotoxicity, and oxidative stress [49]. This chemical and its metabolites, particularly α -zearalenol, which is used as growth promoter in cattle, are able to bind ERs with high affinity and act as strong ER α agonists [51].

3. In Vitro Effects of Phytoestrogens

The proliferation of ER α -positive breast cancer cells is enhanced by estrogens, which induce multiple growth factors, cyclins and cytokines involved in cell survival and cell cycle progression. Although ER α has a proliferative effect, ER β acts as a negative regulator of ER α in breast cancer cells, counteracting the mitogenic effect of estrogens [15,52–54]. Interestingly, in many reported ER-selective bioassays, such as the proliferation of breast cancer cell lines, gene reporter assays in mammalian or non-mammalian cells, and ER binding assays, it was found that most phytoestrogens preferentially interact with ER β and display high specificity toward ER β transactivation [55–57]. Recently, using a fluorescence resonance energy transfer (FRET) assay, Jiang et al. [57] showed that some phytoestrogens, such as genistein, daidzein, equol and liquiritigenin, recruit the coactivator SRC3 much more efficiently to ER β than to ER α . These data strengthen the ER β -selectivity of many phytoestrogens. Hence, a relationship between the ER α /ER β ratio and phytoestrogen effects exists [58,59]. It is suggested that the presence of ER β is associated with the “good” effect of phytoestrogen whereas a high concentration of phytoestrogen in cells expressing ER α was associated to the “bad” effect of phytoestrogen [60].

Several in vitro studies showed that genistein, the most abundant isoflavone present in soybean, has antiproliferative effects on various cancer cells, including prostate, ovarian, and breast

cancer [61–63]. While genistein effects can be mediated at least in part by ER β , other molecular mechanisms, for example caspase-3 activation, have been reported to explain growth inhibition or proapoptotic effects of genistein. Additionally, by direct inhibition of tyrosine kinase activities, genistein is also able to prevent cancer cell growth. For example, genistein pretreatment could significantly reduce the activation of Akt kinase by epidermal growth factor (EGF). The inhibition of nuclear factor κ B (NF- κ B) activity by genistein was also reported in prostate, breast, lung, and pancreatic cancer cells [64–68]. An explanation of this effect is that genistein significantly inhibits Akt kinase activity by decreasing its phosphorylation at serine 473, which can inhibit NF- κ B activity [65,69]. Another study reported that the inhibition of prostate cancer cell growth exerted by genistein is linked to a reduction of telomerase activity that is pivotal for cellular proliferation capacity and immortality. [70]. Together, these actions of the isoflavone genistein could contribute to its apoptotic effects in different human cancer cells. It is also interesting to note that in ER-positive MCF-7 cells, the biphasic actions of genistein can be observed with growth stimulation at low concentrations and inhibition at high concentrations. These observations indicate the complexity of the actions of genistein and phytoestrogens globally for their anti-cancer properties.

One of the key mechanisms underlying the maintenance of genome stability and gene expression is DNA methylation. This process occurs on the cytosine of cytosine-guanine dinucleotides (CpG regions). In the human genome, the majority of CpG regions are methylated, except for those located within CpG-rich regions, called CpG islands, which are usually found within gene promoters. Methylation of CpG islands could lead to the inactivation of gene expression by inhibiting the recruitment of transcription factors necessary to induce transcription. Indeed, DNA methylation/demethylation is a dynamic process that allows certain genes to switch ON and OFF at different periods of time. This process appears to be particularly crucial during embryonic development, tumorigenesis, cell division and cell differentiation. For instance, the OCT4 gene, which is essential to maintain pluripotency in embryonic stem cells, becomes methylated in differentiated tissues to avoid unsuitable pluripotency [71]. On the other hand, the loss of expression of tumor suppressor genes by DNA methylation is often observed in cancerous cells. Re-expression of these genes by the inhibition of DNA methyltransferases has provided many successes in the treatment of cancers.

Interestingly, a recent study showed that genistein can reduce DNA methylation in the promoter regions of the Wingless-int (Wnt) genes, which induces the expression of Wnt proteins in colon cancer cells [72]. The Wnt signaling pathway includes a large number of proteins involved in organogenesis and cell-cell adhesion, cell proliferation and differentiation. In addition to its importance in normal cellular physiology, Wnt signaling is also closely involved with carcinogenesis. Notably, the loss of the expression of Wnt proteins by promoter hypermethylation or abnormal activation of Wnt signaling have been detected in the majority of colon tumors and colon cell lines [73–75]. It is, therefore, possible that genistein, acting as an inhibitor of the DNA methyltransferase, could be able to induce significant Wnt signaling pathways to protect the development of colon cancer. Another study conducted on the human colon cancer cell lines SW480 and HCT15 showed that genistein blocks cell proliferation in the G2 phase of the cell cycle [76]. The authors showed that the action of genistein-inhibition on cell growth is mediated by overexpression of Dickkopf 1 (DKK1) in SW480 and HCT15 cells treated with genistein. DKK1 is a key regulator of the Wnt signaling pathway that promotes cell differentiation and apoptosis. The repression of the tumor suppressor DKK1 by the hypermethylation of its promoter is reported in various diseases, including colorectal cancer [77–80]. However, DNA methylation of the DKK1 promoter is not affected by genistein treatment in either cell line. However, genistein induces acetylation of histone H3 within the promoter region of the DKK1 gene in SW480 and HCT15 cells. This indicates that genistein induction of the expression of the DKK1 gene is linked to the increase in histone acetylation [76]. Another recent study showed that genistein is able to epigenetically reactivate ER α in ER α -negative breast cancer models, both in vitro and in vivo [66]. Similarly, in the prostate cancer cell lines LNCaP and LAPC-4, genistein was able to increase the expression of ER β through decreasing the methylation of the ER β promoter at physiological ranges (0.5–10 μ mol/L) [81]. Hence,

genistein could increase the sensitivity of these cancers to endocrine therapies, such as the antiestrogen tamoxifen. In this study, the authors showed that genistein significantly increased histone acetylation patterns in the ER α promoter by inhibiting the enzymatic activities of histone deacetylase (HDAC). It is of interest to note that this effect was enhanced in a synergistic manner when ER α -negative MDA-MB-231 breast cancer cells were co-treated with genistein and TSA, an inhibitor of HDACs. Importantly, the anticancer properties of tamoxifen to inhibit cell growth become much more efficient both *in vitro* and *in vivo*, in xenograft nude mice as well as in spontaneous breast tumor mouse models, in the presence of genistein [66]. Together, these studies suggest that in addition to DNA methylation, genistein may also modify histone marks of critical genes to prevent cancer development and progression. These epigenetic actions of genistein mediate the activation of tumor suppressor genes in cancer cell lines but also in animal models. The consequences of endocrine disruptors to different cell types have been widely studied. However, in industrial countries, detectable levels of EDCs were found in human, indicating that people are constantly exposed. Hence, studies on acute effects do not reflect the consequences of constant exposure. Chronic exposure of MCF7 cells with genistein induces a down-regulation of the PI3K/Akt signaling pathway, inhibits the growth-promoting activity of E2 or EGF, and reduces histone H3 acetylation without affecting ER expression. This indicates that chronic treatment leads to epigenetic changes in the cells [82,83].

4. In Vivo Effects of Phytoestrogens

The concentration of phytoestrogens in the plasma is considerably different in the human population. For example, in Finnish men the average plasma concentration of genistein is about 0.5 nM whereas it is about 276 nM in Japanese males. However, after absorption of dietary phytoestrogen, a plasmatic peak was detected between 0.2 and 6.5 μ M with bioavailability between 5% and 66% [84]. Moreover a pharmacokinetic study performed on postmenopausal women found that the concentration of free genistein could reach 40 nM [85]. Epidemiological studies reveal that a lower risk of breast and prostate cancers is observed in Asians, who consume 20–50 times more soy products than Americans [86]. *In vivo* experimental studies have also reported that some dietary components, including isoflavones and enterolignans, could inhibit the development of cancers [69,87,88]. This suggests that active molecules in the soybean, such as genistein, daidzein, equol and glycitein, may act as natural chemopreventive agents and could be used against tumor progression in humans. Moreover, clinical studies carried out to assess the effectiveness of isoflavones in patients with prostate cancer found that isoflavone supplementation significantly reduced the expression of the poor prognostic tumoral marker, prostate specific antigen (PSA), and the expression of androgen receptor (AR), but without effecting the expression of ER β or circulating hormones. These studies have suggested that isoflavones, including genistein and daidzein, may be beneficial in the prevention of prostate cancer by inhibiting the expression of AR and PSA [89,90]. Furthermore, *in vivo* xenografts in mice model and *in vitro* studies conducted on androgen-dependent (LNCaP) and androgen-independent (DU145 or PC3) prostate cancer cell lines showed that some phytoestrogens, such as coumestrol, are able to elicit caspase-dependent apoptosis, supporting the hypothesis that phytoestrogens may have anticancer effects in prostate cancer. Conversely, some clinical trials seeking to establish that consumption of phytoestrogens is beneficial in prostate cancer have been inconclusive [91]. For instance, a double-blind trial conducted by Adams and collaborators [92] showed no significant difference in PSA among men who did or did not consume a diet rich in isoflavones for 12 months. Although this study is limited by the relatively small number of patients, it could indicate that the period and duration of treatment may be essential for the anticancer effects of isoflavones. While the exact mechanisms by which isoflavones can prevent the development or progression of prostate cancer remain unclear, many mechanisms have been proposed, including the regulation of genes involved in the cell cycle, such as an upregulation of p21 resulting in cell cycle arrest at the G1/S phase, apoptosis, antioxidant effects, DNA repair, inhibition of angiogenesis and metastasis, and also the antagonism of estrogen and androgen signaling pathways (for review see [93]). It should be noted, therefore, that changes in some signaling pathways

or in the expression of key enzymes involved in steroid metabolism during different stages of prostate cancer could play an essential role in the effects of phytoestrogens.

In vivo studies carried out in a true physiological context in humans and animals have indicated that the food content of isoflavones poses no safety issue, as generally consumed in diets based on soy products [94]. Moreover, the concentration of isoflavones varies considerably depending on the place of soybean cultivation (from 85 mg/100 g in Taiwan to 178 mg/100 g in Korea) or according to the culinary process (6 mg/100 g 195 mg/100 g in Foojook soup) [95]. A similar observation was made in lyophilized cabbage compared to fermented cabbage [96]. As the structure of the major isoflavone compounds is close to that of E2 and because these compounds are known to have weak estrogenic activities, the possible effects of some isoflavones on estrogen-target tissues and on reproductive function have been extensively explored [97–100]. However, there are conflicting results regarding the effects of isoflavones on reproductive function because the long-term studies on the impact of these compounds on the development and function of reproductive tissues are not sufficient. In addition, comparisons between different studies are complicated because there are differences in the experimental design, such as the physiological state of the animal, the presence of circulating hormone, and the duration, doses and methods of exposure (injection or gavage). More importantly, differences in the metabolism of isoflavones between animal models and humans can also give inconclusive results. Thus, all these parameters must be considered when assessing the impact of isoflavones on reproductive function. All major soybean isoflavones, genistein, daidzein, equol and glycitein, were reported to be estrogenic in the mouse or rat uterine growth assay [97,98]. For instance, 100 mg/kg body weight of genistein or equol, administered by gavage for 4 successive days (post-natal at day 17–20), was found to significantly increase uterine weights and the expression of ER α in the uterus [97]. Another study compared the estrogenic potential of several phytoestrogens, including genistein, daidzein and coumestrol in immature mice using different morphological and biochemical tests on the uterus. Interestingly, while certain compounds, such as genistein and coumestrol, showed estrogenic activity in all tests, others showed estrogenicity in only a single test or did not show estrogenicity in any test [101].

The estrogenic potency of isoflavones was also assessed, in vivo, in several non-mammalian model organisms. For example, goldfish or medaka fed for several weeks with a diet containing coumestrol or genistein showed an increased production of the vitellogenin, an egg yolk protein precursor, which is normally produced in the liver under estrogenic stimulation. However, no adverse effects on reproduction function (fecundity and fertility) have been reported [102,103], indicating that the production of vitellogenin may serve as an indicator of estrogen exposure but not as an indicator of reproductive dysfunction by estrogen exposure. In contrast, a recent study from Bennetau-Pellisero and collaborators reported [104] that goldfish fed soybean meal for 20 weeks after hatching show a reduction in fertility success and larvae production. Particularly, both male and female fish groups displayed changes in the plasma testosterone and E2 levels, as well as in their spermatogenesis process and oocyte maturation.

Concerning male reproductive function, a study has been conducted on marmosets fed with soy-based milk during the first six weeks of life and compared to animals fed with a standard cow's milk-based diet [105]. This study reported that soy-fed marmosets had body weights, organ weights (prostate, seminal vesicles, pituitary, thymus and spleen) and penis lengths comparable to the other animals. Although lower blood testosterone and higher Sertoli and Leydig cell numbers per testis were observed in soy-fed marmosets, no adverse reproductive consequences were detected in adulthood, including the timing of puberty and overall fertility [105]. On the other hand, Adachi et al. have carried out a toxicogenomic analysis in mice to investigate long-term effects of neonatal exposure to genistein on testicular gene expression. In addition, the authors used diethylstilbestrol (DES), known as a potent estrogen, as a positive control because exposure to DES has been reported to induce morphological changes and alteration of gene expression in reproductive organs. Male mice fed with genistein (1000 μ g/mouse/day) from days 1–6 after birth did not show any morphological changes in testes at 12 weeks of age, despite decreased ER and AR gene expression. As expected, DES (50 μ g/mouse/day)

did show gene expression and morphological changes in testes at 12 weeks of age [106]. This suggests that neonatal exposure to genistein has no long-term effects, according to this analysis.

Following menopause in women, there is more brain-related pathology, incidence of stroke and loss of bone mass observed than in men [107]. Because estrogens are neuroprotective agents that are involved in bone remodeling, one possible explanation may be the decline in estrogen levels. Isoflavones are generally considered to have beneficial effects on bone and brain, although controversial results have been published [108]. In ovariectomized rats, genistein showed a weak osteoprotective effect by promoting bone mineral density [109,110]. Similarly, coumestrol showed a neuroprotection effect in ovariectomized rats subjected to global ischemia [111]. Interestingly, using an ER antagonist, the authors showed that the neuroprotective actions of coumestrol are only partially abolished, suggesting that in addition to classical ER signaling, coumestrol may act via other cellular pathways. Thus, the beneficial effects of isoflavones may depend on the quantity and ratio of the expression of ER subtypes, the endogenous steroid hormones and period of life [111]. Moreover, it would be interesting to find out the cellular targets of coumestrol mediating its neuroprotective action. These cellular pathways could be used in the therapeutic potential of coumestrol in the treatment of pathologies related to the central nervous system.

There is growing evidence suggesting that during critical windows of prenatal and postnatal development, environmental chemicals can induce epigenetic modifications, affecting gene expression and consequently impacting developmental pathways. Importantly, it has been suggested that the effects of some environmental chemicals could act across generations, leading to phenotypic and physiological variation in the development and behavior of offspring. The transmission can be a consequence of changes in the transcriptome and epigenome programming within germ cells. While these effects have been recently reported for a number of environmental compounds, such as vinclozolin, atrazine, bisphenol A (BPA), DES, and dioxine [112–118], studies on the potential transgenerational effects of phytoestrogens are very rare, if any, and need additional work [69,115].

Although the situation is different for phytoestrogens produced by plants, it is worth noting that some fungi also produce compounds, called mycotoxins, with estrogenic properties. For example, zearalenone and fusarin C act as estrogen agonists and are classified as mycoestrogens. These compounds, which could contaminate improperly stored grains, have been linked to increased cancer rates. *In vitro*, fusarin C, as well as zearalenone and its metabolites, can stimulate the growth and proliferation of human breast tumor cells [119–121]. Moreover, *in vivo* exposure of rats to environmental doses of zearalenone in the last two to three weeks of fetal development and the first days after birth resulted in long-term changes in the development of the mammary gland associated with increased risk for the development of mammary tumors [122].

5. Conclusions

Although more research is needed, it is clear that some natural compounds from plants, such as phytoestrogens, could have beneficial effects on certain diseases, such as cancer or neurodegenerative diseases. However, *in vitro* or *in vivo* studies to analyze the final effects of phytoestrogens may be quite different at low (<1 μM) or high concentrations (>10 μM). For instance, at low doses (from 10 nM to 1 μM), genistein showed mitogenic effects on breast cancer cell growth, whereas at higher concentrations (>10 μM), it showed antiproliferative effects [123,124]. Some of these effects are explained by their interactions with ER subtypes. The ratios and the expressions of ER α and ER β are different in various tissues depending on the period of life. ER α is mostly expressed in tissues such as the mammary gland, uterus, liver and pituitary, while ER β is expressed in tissues such as the brain, bone and bladder. Moreover, the abilities of ER subtypes to recruit cofactors, regulate gene expression and stimulate or inhibit cell growth are slightly different. Therefore, *in vivo*, phytoestrogens may have a complex role, acting as weak estrogens and antiestrogens depending on the tissue. Furthermore, it is believed that the signaling pathways induced by phytoestrogens are not completely identical to those induced by estrogens. As illustrated in Figure 3, phytoestrogens may have different mechanisms of

action; therefore, some of these compounds could be considered therapeutic agents and used alone or in combination with usual hormone therapies. For example, the protective effect of isoflavones on prostate cancer may be related to their effects on metabolic pathways involved in androgen and estrogen synthesis [125] or to their epigenetic modifications of DNA, such as the demethylation of CpG islands within the promoters of tumor suppressor genes [81,126,127]. On the other hand, the phytoestrogen coumestrol, which exhibits an important cancer-preventive effect in estrogen-responsive carcinomas, was recently reported to inhibit epithelial ovarian cancer proliferation and invasion by modifying AKT, p70S6K and ERK1/2 phosphorylation [128]. Moreover, previous studies showed an antagonistic effect of genistein and apigenin against the association of ER α with the ubiquitous calcium-dependent protein, calmodulin (CaM). By interacting with ER α , CaM plays a key role in the stabilization and transcriptional activity of ER α dimers at the ERE. The agonistic effect of genistein and apigenin in this interaction may also account for the anti-tumor origin of these compounds against ER-positive breast cancers [129,130]. It is, therefore, essential to continue advances in the understanding diverse signaling pathways activated by phytoestrogens, to fully exploit their anticancer properties and/or their potential roles in estrogen-related diseases. Accordingly, it should also be remembered that changes in the expression or activity of nuclear and membrane receptors for steroids and growth factors, as well as key steroid synthesis enzymes, during cancer progression could play crucial roles in the effects of phytoestrogens (Figure 3). Indeed, flavonoids, especially flavones (ex: luteolin) and flavanones (ex: naringenin), are described as potent inhibitors of aromatase activity [131]. Aromatase is the main enzyme that participates in the transformation of testosterone into estradiol and is hence involved in breast cancer pathology. Moreover, luteolin was also shown to downregulate aromatase gene expression [131]. Phytoestrogens are also able to inhibit proteasome [132], which appears to be essential for breast cancer cell survival [133]. For example, apigenin is capable of inhibiting the catalytic activity of proteasomes, leading to stabilization of ER β and apoptosis of prostate cancer cells [134].

An important application of phytoestrogens is that they could be used as an alternative to the synthetic selective estrogen receptor modulators (SERMs), which exhibit estrogen agonist or antagonist activity in a tissue-specific manner. Indeed, SERMs are used in the treatment of some estrogen-associated pathologies, such as breast cancer, brain diseases, osteoporosis and menopausal symptoms. In other words, the challenge is to minimize the adverse effects of ER (mitogenic effect) without reducing the beneficial effects (protective effects), such as the control of cell differentiation, neuroprotection, anti-osteoporosis effects, and anti-oxidant activity. Our recent study screening the SERM activity of these compounds revealed a beneficial effect of apigenin and resveratrol, whereas zearalenone has been characterized as having a strong ER-agonist property in breast cancer cell lines and having adverse effects in neuritogenesis [135].

Although recent studies have reported that certain environmental agents caused epigenetic effects that could act across generations, leading to physiological changes of the offspring, there are no examples of perinatal exposure to phytoestrogens at environmentally relevant doses. There is still a need to understand the molecular mechanisms and to investigate how these compounds can influence epigenetic patterns during development.

In this review, we have discussed the effects of phytoestrogens used alone. However, populations are exposed to several compounds at the same time. Thus, it might be important to perform studies of the effect of mixtures of botanical estrogen on human health to improve recommendations for public health.

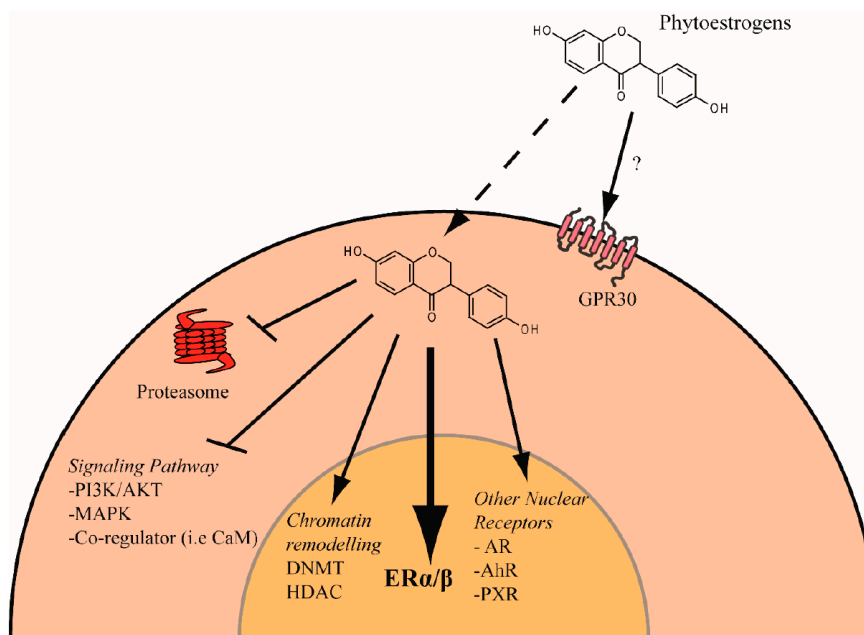


Figure 3. Different targets of phytoestrogens in cells. Cell signaling pathways for estrogens through the nuclear receptors ER α , ER β and the transmembrane receptor G-protein-coupled ER (GPER; formerly known as GPR30) [136] are shown. Phytoestrogens are able to inhibit mitogenic pathways via ER α or PI3K/MAPK, which in turn inhibit cancer cell proliferation and invasion by modifying AKT, p70S6K and ERK1/2 phosphorylation as well as interaction between ER α with various coregulatory proteins such as calmodulin (CaM). Activation of ER β inhibits dedifferentiation pathways and induces apoptosis and cell cycle arrest. GPER activation is anti-tumorigenic, as it upregulates p21 and induces cell cycle arrest in prostate cancer [137]. Epigenetic modifications by phytoestrogens, such as demethylation of CpG islands within the promoters of tumor suppressor genes, could contribute to cell growth arrest. Inhibition of proteasomes by phytoestrogens also appears to be another mechanism of phytoestrogen activity in decreasing cancer cell survival.

Acknowledgments: This work was supported by Fond Unique Interministeriel (FUI, project mVolio), the French Ministry of Ecology, Energy and Sustainable Development (PNRPE); La Ligue Contre le Cancer, the INSERM and CNRS.

Author Contributions: Sylvain Lecomte, François Ferrière and Florence Demay were the major contributors in writing the manuscript. Sylvain Lecomte, Florence Demay, François Ferrière and Farzad Pakdel read, corrected and approved the final manuscript

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Couse, J.F.; Korach, K.S. Estrogen receptor null mice: What have we learned and where will they lead us? *Endocr. Rev.* **1999**, *20*, 358–417. [[CrossRef](#)] [[PubMed](#)]
2. Gustafsson, J.-A. What pharmacologists can learn from recent advances in estrogen signaling. *Trends Pharmacol. Sci.* **2003**, *24*, 479–485. [[CrossRef](#)]
3. Germain, P.; Staels, B.; Dacquet, C.; Spedding, M.; Laudet, V. Overview of nomenclature of nuclear receptors. *Pharmacol. Rev.* **2006**, *58*, 685–704. [[CrossRef](#)] [[PubMed](#)]
4. Safe, S.; Kim, K.; Kim, K. Non-classical genomic estrogen receptor (ER)/specificity protein and ER/activating protein-1 signaling pathways. *J. Mol. Endocrinol.* **2008**, *41*, 263–275. [[CrossRef](#)] [[PubMed](#)]
5. Carroll, J.S.; Meyer, C.A.; Song, J.; Li, W.; Geistlinger, T.R.; Eeckhoute, J.; Brodsky, A.S.; Keeton, E.K.; Fertuck, K.C.; Hall, G.F.; et al. Genome-wide analysis of estrogen receptor binding sites. *Nat. Genet.* **2006**, *38*, 1289–1297. [[CrossRef](#)] [[PubMed](#)]

6. Le Dily, F.; Beato, M. TADs as modular and dynamic units for gene regulation by hormones. *FEBS Lett.* **2015**, *589*, 2885–2892. [[CrossRef](#)] [[PubMed](#)]
7. Le Dily, F.; Baù, D.; Pohl, A.; Vicent, G.P.; Serra, F.; Soronellas, D.; Castellano, G.; Wright, R.H.G.; Ballare, C.; Filion, G.; et al. Distinct structural transitions of chromatin topological domains correlate with coordinated hormone-induced gene regulation. *Genes Dev.* **2014**, *28*, 2151–2162. [[CrossRef](#)] [[PubMed](#)]
8. Nilsson, S.; Mäkelä, S.; Treuter, E.; Tujague, M.; Thomsen, J.; Andersson, G.; Enmark, E.; Pettersson, K.; Warner, M.; Gustafsson, J.A. Mechanisms of estrogen action. *Physiol. Rev.* **2001**, *81*, 1535–1565. [[PubMed](#)]
9. La Rosa, P.; Pesiri, V.; Leclercq, G.; Marino, M.; Acconcia, F. Palmitoylation regulates 17 β -estradiol-induced estrogen receptor- α degradation and transcriptional activity. *Mol. Endocrinol.* **2012**, *26*, 762–774. [[CrossRef](#)] [[PubMed](#)]
10. Pedram, A.; Razandi, M.; Aitkenhead, M.; Hughes, C.C.W.; Levin, E.R. Integration of the non-genomic and genomic actions of estrogen. Membrane-initiated signaling by steroid to transcription and cell biology. *J. Biol. Chem.* **2002**, *277*, 50768–50775. [[CrossRef](#)] [[PubMed](#)]
11. Vicent, G.P.; Nacht, A.S.; Zaurín, R.; Ballaré, C.; Clausell, J.; Beato, M. Minireview: Role of kinases and chromatin remodeling in progesterone signaling to chromatin. *Mol. Endocrinol.* **2010**, *24*, 2088–2098. [[CrossRef](#)] [[PubMed](#)]
12. Levin, E.R. Extranuclear estrogen receptor's roles in physiology: Lessons from mouse models. *Am. J. Physiol. Endocrinol. Metab.* **2014**, *307*, E133–E140. [[CrossRef](#)] [[PubMed](#)]
13. Gosden, J.R.; Middleton, P.G.; Rout, D. Localization of the human oestrogen receptor gene to chromosome 6q24—q27 by in situ hybridization. *Cytogenet. Cell Genet.* **1986**, *43*, 218–220. [[CrossRef](#)] [[PubMed](#)]
14. Enmark, E.; Pelto-Huikko, M.; Grandien, K.; Lagercrantz, S.; Lagercrantz, J.; Fried, G.; Nordenskjöld, M.; Gustafsson, J.A. Human estrogen receptor β -gene structure, chromosomal localization, and expression pattern. *J. Clin. Endocrinol. Metab.* **1997**, *82*, 4258–4265. [[CrossRef](#)] [[PubMed](#)]
15. Kerdivel, G.; Flouriot, G.; Pakdel, F. Modulation of estrogen receptor α activity and expression during breast cancer progression. *Vitam. Horm.* **2013**, *93*, 135–160. [[PubMed](#)]
16. Wang, Z.; Zhang, X.; Shen, P.; Loggie, B.W.; Chang, Y.; Deuel, T.F. Identification, cloning, and expression of human estrogen receptor- α 36, a novel variant of human estrogen receptor- α 66. *Biochem. Biophys. Res. Commun.* **2005**, *336*, 1023–1027. [[CrossRef](#)] [[PubMed](#)]
17. Saunders, P.T. Oestrogen receptor β (ER β). *Rev. Reprod.* **1998**, *3*, 164–171. [[CrossRef](#)] [[PubMed](#)]
18. Delbès, G.; Levacher, C.; Duquenne, C.; Racine, C.; Pakarinen, P.; Habert, R. Endogenous estrogens inhibit mouse fetal Leydig cell development via estrogen receptor α . *Endocrinology* **2005**, *146*, 2454–2461. [[CrossRef](#)] [[PubMed](#)]
19. Staub, C.; Rauch, M.; Ferrière, F.; Trépos, M.; Dorval-Coiffec, I.; Saunders, P.T.; Cobellis, G.; Flouriot, G.; Saligaut, C.; Jégou, B. Expression of Estrogen Receptor ESR1 and Its 46-kDa Variant in the Gubernaculum Testis. *Biol. Reprod.* **2005**, *73*, 703–712. [[CrossRef](#)] [[PubMed](#)]
20. Wilson, M.E.; Westberry, J.M.; Trout, A.L. Estrogen receptor- α gene expression in the cortex: Sex differences during development and in adulthood. *Horm. Behav.* **2011**, *59*, 353–357. [[CrossRef](#)] [[PubMed](#)]
21. Fan, X.; Kim, H.-J.; Warner, M.; Gustafsson, J.-A. Estrogen receptor β is essential for sprouting of nociceptive primary afferents and for morphogenesis and maintenance of the dorsal horn interneurons. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 13696–13701. [[CrossRef](#)] [[PubMed](#)]
22. Kerdivel, G.; Habauzit, D.; Pakdel, F. Assessment and molecular actions of endocrine-disrupting chemicals that interfere with estrogen receptor pathways. *Int. J. Endocrinol.* **2013**, *2013*, 501851. [[CrossRef](#)] [[PubMed](#)]
23. Sohoni, P.; Sumpter, J.P. Several environmental oestrogens are also anti-androgens. *J. Endocrinol.* **1998**, *158*, 327–339. [[CrossRef](#)] [[PubMed](#)]
24. Sonnenschein, C.; Soto, A.M. An updated review of environmental estrogen and androgen mimics and antagonists. *J. Steroid Biochem. Mol. Biol.* **1998**, *65*, 143–150. [[CrossRef](#)]
25. Rasier, G.; Toppari, J.; Parent, A.-S.; Bourguignon, J.-P. Female sexual maturation and reproduction after prepubertal exposure to estrogens and endocrine disrupting chemicals: A review of rodent and human data. *Mol. Cell. Endocrinol.* **2006**, *254–255*, 187–201. [[CrossRef](#)] [[PubMed](#)]
26. Toppari, J.; Virtanen, H.; Skakkebaek, N.E.; Main, K.M. Environmental effects on hormonal regulation of testicular descent. *J. Steroid Biochem. Mol. Biol.* **2006**, *102*, 184–186. [[CrossRef](#)] [[PubMed](#)]
27. Barouki, R.; Coumoul, X.; Fernandez-Salguero, P.M. The aryl hydrocarbon receptor, more than a xenobiotic-interacting protein. *FEBS Lett.* **2007**, *581*, 3608–3615. [[CrossRef](#)] [[PubMed](#)]

28. Liu, Z.; Kanjo, Y.; Mizutani, S. A review of phytoestrogens: Their occurrence and fate in the environment. *Water Res.* **2010**, *44*, 567–577. [[CrossRef](#)] [[PubMed](#)]
29. Fantini, M.; Benvenuto, M.; Masuelli, L.; Frajese, G.; Tresoldi, I.; Modesti, A.; Bei, R. In vitro and in vivo antitumoral effects of combinations of polyphenols, or polyphenols and anticancer drugs: Perspectives on cancer treatment. *Int. J. Mol. Sci.* **2015**, *16*, 9236–9282. [[CrossRef](#)] [[PubMed](#)]
30. Shukla, S.; Gupta, S. Apigenin: A promising molecule for cancer prevention. *Pharm. Res.* **2010**, *27*, 962–978. [[CrossRef](#)] [[PubMed](#)]
31. Marzocchella, L.; Fantini, M.; Benvenuto, M.; Masuelli, L.; Tresoldi, I.; Modesti, A.; Bei, R. Dietary flavonoids: Molecular mechanisms of action as anti-inflammatory agents. *Recent Pat. Inflamm. Allergy Drug Discov.* **2011**, *5*, 200–220. [[CrossRef](#)] [[PubMed](#)]
32. Beecher, G.R. Overview of dietary flavonoids: Nomenclature, occurrence and intake. *J. Nutr.* **2003**, *133*, 3248S–3254S. [[PubMed](#)]
33. Setchell, K.D.R.; Clerici, C. Equol: History, chemistry, and formation. *J. Nutr.* **2010**, *140*, 1355S–1362S. [[CrossRef](#)] [[PubMed](#)]
34. National Library of Medicine-MeSH National Library of Medicine-MeSH, 2016. Available online: https://www.nlm.nih.gov/cgi/mesh/2016/MB_cgi (accessed on 12 November 2016).
35. Van de Schans, M.G. M.; Vincken, J.-P.; Bovee, T.F. H.; Cervantes, A.D.; Logtenberg, M.J.; Gruppen, H. Structural changes of 6a-hydroxy-pterocarpan upon heating modulate their estrogenicity. *J. Agric. Food Chem.* **2014**, *62*, 10475–10484. [[CrossRef](#)] [[PubMed](#)]
36. Zimmermann, M.C.; Tilghman, S.L.; Boué, S.M.; Salvo, V.A.; Elliott, S.; Williams, K.Y.; Skripnikova, E.V.; Ashe, H.; Payton-Stewart, F.; Vanhoy-Rhodes, L.; et al. Glyceollin I, a novel antiestrogenic phytoalexin isolated from activated soy. *J. Pharmacol. Exp. Ther.* **2010**, *332*, 35–45. [[CrossRef](#)] [[PubMed](#)]
37. Ayers, A.R.; Ebel, J.; Finelli, F.; Berger, N.; Albersheim, P. Host-pathogen interactions: IX. Quantitative assays of elicitor activity and characterization of the elicitor present in the extracellular medium of cultures of *Phytophthora megasperma* var. *sojae*. *Plant Physiol.* **1976**, *57*, 751–759. [[CrossRef](#)] [[PubMed](#)]
38. Nikov, G.N.; Hopkins, N.E.; Boue, S.; Alworth, W.L. Interactions of dietary estrogens with human estrogen receptors and the effect on estrogen receptor-estrogen response element complex formation. *Environ. Health Perspect.* **2000**, *108*, 867–872. [[CrossRef](#)] [[PubMed](#)]
39. Tuskaev, V.A. Synthesis and biological activity of coumestan derivatives. *Pharm. Chem. J.* **2013**, *47*, 1–11. [[CrossRef](#)]
40. Nehybová, T.; Šmarda, J.; Beneš, P. Plant coumestans: Recent advances and future perspectives in cancer therapy. *Anticancer Agents Med. Chem.* **2014**, *14*, 1351–1362. [[CrossRef](#)] [[PubMed](#)]
41. Bickoff, E.M.; Booth, A.N.; Lyman, R.L.; Livingston, A.L.; Thompson, C.R.; Deeds, F. Coumestrol, a new estrogen isolated from forage crops. *Science* **1957**, *126*, 969–970. [[CrossRef](#)] [[PubMed](#)]
42. Kuiper, G.G.; Lemmen, J.G.; Carlsson, B.; Corton, J.C.; Safe, S.H.; van der Saag, P.T.; van der Burg, B.; Gustafsson, J.A. Interaction of estrogenic chemicals and phytoestrogens with estrogen receptor β . *Endocrinology* **1998**, *139*, 4252–4263. [[CrossRef](#)] [[PubMed](#)]
43. Stervbo, U.; Vang, O.; Bonnesen, C. A review of the content of the putative chemopreventive phytoalexin resveratrol in red wine. *Food Chem.* **2007**, *101*, 449–457. [[CrossRef](#)]
44. Gehm, B.D.; McAndrews, J.M.; Chien, P.Y.; Jameson, J.L. Resveratrol, a polyphenolic compound found in grapes and wine, is an agonist for the estrogen receptor. *Proc. Natl. Acad. Sci. USA* **1997**, *94*, 14138–14143. [[CrossRef](#)] [[PubMed](#)]
45. Le Corre, L.; Chalabi, N.; Delort, L.; Bignon, Y.-J.; Bernard-Gallon, D.J. Resveratrol and breast cancer chemoprevention: Molecular mechanisms. *Mol. Nutr. Food Res.* **2005**, *49*, 462–471. [[CrossRef](#)] [[PubMed](#)]
46. Rosmalena, A.; Prasasty, V.D.; Hanafi, M.; Budianto, E.; Elya, B. Lignan derivatives potential as Plasmodium falciparum lactate dehydrogenase inhibitors: Molecular docking approach of antiplasmodial drug design. *Int. J. Pharm. Pharm. Sci.* **2015**, *7*, 394–398.
47. Mueller, S.O.; Simon, S.; Chae, K.; Metzler, M.; Korach, K.S. Phytoestrogens and their human metabolites show distinct agonistic and antagonistic properties on estrogen receptor α (ER α) and ER β in human cells. *Toxicol. Sci.* **2004**, *80*, 14–25. [[CrossRef](#)] [[PubMed](#)]
48. Shier, W.T.; Shier, A.C.; Xie, W.; Mirocha, C.J. Structure-activity relationships for human estrogenic activity in zearalenone mycotoxins. *Toxicon* **2001**, *39*, 1435–1438. [[CrossRef](#)]

49. EFSA Panel on Contaminants in the Food Chain Scientific. Opinion on the risks for public health related to the presence of zearalenone in food. *EFSA J.* **2011**, *9*, 2197.
50. Lin, P.; Chen, F.; Sun, J.; Zhou, J.; Wang, X.; Wang, N.; Li, X.; Zhang, Z.; Wang, A.; Jin, Y. Mycotoxin zearalenone induces apoptosis in mouse Leydig cells via an endoplasmic reticulum stress-dependent signalling pathway. *Reprod. Toxicol.* **2015**, *52*, 71–77. [[CrossRef](#)] [[PubMed](#)]
51. Le Guevel, R.; Pakdel, F. Assessment of oestrogenic potency of chemicals used as growth promoter by in vitro methods. *Hum. Reprod.* **2001**, *16*, 1030–1036. [[CrossRef](#)] [[PubMed](#)]
52. Frasar, J.; Danes, J.M.; Komm, B.; Chang, K.C.N.; Lyttle, C.R.; Katzenellenbogen, B.S. Profiling of estrogen up- and down-regulated gene expression in human breast cancer cells: Insights into gene networks and pathways underlying estrogenic control of proliferation and cell phenotype. *Endocrinology* **2003**, *144*, 4562–4574. [[CrossRef](#)] [[PubMed](#)]
53. Chang, E.C.; Frasar, J.; Komm, B.; Katzenellenbogen, B.S. Impact of estrogen receptor β on gene networks regulated by estrogen receptor α in breast cancer cells. *Endocrinology* **2006**, *147*, 4831–4842. [[CrossRef](#)] [[PubMed](#)]
54. Chang, E.C.; Cham, T.H.; Park, S.-H.; Helferich, W.G.; Komm, B.; Katzenellenbogen, J.A.; Katzenellenbogen, B.S. Estrogen Receptors α and β as Determinants of Gene Expression: Influence of Ligand, Dose, and Chromatin Binding. *Mol. Endocrinol.* **2008**, *22*, 1032–1043. [[CrossRef](#)] [[PubMed](#)]
55. Zhao, L.; Mao, Z.; Brinton, R.D. A select combination of clinically relevant phytoestrogens enhances estrogen receptor β -binding selectivity and neuroprotective activities in vitro and in vivo. *Endocrinology* **2009**, *150*, 770–783. [[CrossRef](#)] [[PubMed](#)]
56. Shanle, E.K.; Hawse, J.R.; Xu, W. Generation of stable reporter breast cancer cell lines for the identification of ER subtype selective ligands. *Biochem. Pharmacol.* **2011**, *82*, 1940–1949. [[CrossRef](#)] [[PubMed](#)]
57. Jiang, Y.; Gong, P.; Madak-Erdogan, Z.; Martin, T.; Jeyakumar, M.; Carlson, K.; Khan, I.; Smillie, T.J.; Chittiboyina, A.G.; Rotte, S.C.K.; et al. Mechanisms enforcing the estrogen receptor β selectivity of botanical estrogens. *FASEB J.* **2013**, *27*, 4406–4418. [[CrossRef](#)] [[PubMed](#)]
58. Pons, D.G.; Nadal-Serrano, M.; Blanquer-Rossello, M.M.; Sastre-Serra, J.; Oliver, J.; Roca, P. Genistein Modulates Proliferation and Mitochondrial Functionality in Breast Cancer Cells Depending on ER α /ER β Ratio. *J. Cell. Biochem.* **2014**, *115*, 949–958. [[CrossRef](#)] [[PubMed](#)]
59. Sotoca, A.M.; Ratman, D.; van der Saag, P.; Ström, A.; Gustafsson, J.A.; Vervoort, J.; Rietjens, I.M.C.M.; Murk, A.J. Phytoestrogen-mediated inhibition of proliferation of the human T47D breast cancer cells depends on the ER α /ER β ratio. *J. Steroid Biochem. Mol. Biol.* **2008**, *112*, 171–178. [[CrossRef](#)] [[PubMed](#)]
60. Russo, M.; Russo, G.L.; Daglia, M.; Kasi, P.D.; Ravi, S.; Nabavi, S.F.; Nabavi, S.M. Understanding genistein in cancer: The good and the bad effects: A review. *Food Chem.* **2016**, *196*, 589–600. [[CrossRef](#)] [[PubMed](#)]
61. Kuo, S.M. Antiproliferative potency of structurally distinct dietary flavonoids on human colon cancer cells. *Cancer Lett.* **1996**, *110*, 41–48. [[CrossRef](#)]
62. Hwang, K.-A.; Park, M.-A.; Kang, N.-H.; Yi, B.-R.; Hyun, S.-H.; Jeung, E.-B.; Choi, K.-C. Anticancer effect of genistein on BG-1 ovarian cancer growth induced by 17 β -estradiol or bisphenol A via the suppression of the crosstalk between estrogen receptor α and insulin-like growth factor-1 receptor signaling pathways. *Toxicol. Appl. Pharmacol.* **2013**, *272*, 637–646. [[CrossRef](#)] [[PubMed](#)]
63. Prietsch, R.F.; Monte, L.G.; da Silva, F.A.; Beira, F.T.; Del Pino, F.A.B.; Campos, V.F.; Collares, T.; Pinto, L.S.; Spanevello, R.M.; Gamaro, G.D.; et al. Genistein induces apoptosis and autophagy in human breast MCF-7 cells by modulating the expression of proapoptotic factors and oxidative stress enzymes. *Mol. Cell. Biochem.* **2014**, *390*, 235–242. [[CrossRef](#)] [[PubMed](#)]
64. Li, Y.; Sarkar, F.H. Inhibition of nuclear factor κ B activation in PC3 cells by genistein is mediated via Akt signaling pathway. *Clin. Cancer Res.* **2002**, *8*, 2369–2377. [[PubMed](#)]
65. Gong, L.; Li, Y.; Nedeljkovic-Kurepa, A.; Sarkar, F.H. Inactivation of NF- κ B by genistein is mediated via Akt signaling pathway in breast cancer cells. *Oncogene* **2003**, *22*, 4702–4709. [[CrossRef](#)] [[PubMed](#)]
66. Li, Y.; Meeran, S.M.; Patel, S.N.; Chen, H.; Hardy, T.M.; Tollefsbol, T.O. Epigenetic reactivation of estrogen receptor- α (ER α) by genistein enhances hormonal therapy sensitivity in ER α -negative breast cancer. *Mol. Cancer* **2013**, *12*, 9. [[CrossRef](#)] [[PubMed](#)]
67. Suzuki, R.; Kang, Y.; Li, X.; Roife, D.; Zhang, R.; Fleming, J.B. Genistein potentiates the antitumor effect of 5-Fluorouracil by inducing apoptosis and autophagy in human pancreatic cancer cells. *Anticancer Res.* **2014**, *34*, 4685–4692. [[PubMed](#)]

68. Li, C.; Teng, R.-H.; Tsai, Y.-C.; Ke, H.-S.; Huang, J.-Y.; Chen, C.-C.; Kao, Y.-L.; Kuo, C.-C.; Bell, W.R.; Shieh, B. H-Ras oncogene counteracts the growth-inhibitory effect of genistein in T24 bladder carcinoma cells. *Br. J. Cancer* **2005**, *92*, 80–88. [[CrossRef](#)] [[PubMed](#)]
69. Sarkar, F.H.; Li, Y.; Wang, Z.; Padhye, S. Lesson learned from nature for the development of novel anti-cancer agents: Implication of isoflavone, curcumin, and their synthetic analogs. *Curr. Pharm. Des.* **2010**, *16*, 1801–1812. [[CrossRef](#)] [[PubMed](#)]
70. Jagadeesh, S.; Kyo, S.; Banerjee, P.P. Genistein represses telomerase activity via both transcriptional and posttranslational mechanisms in human prostate cancer cells. *Cancer Res.* **2006**, *66*, 2107–2115. [[CrossRef](#)] [[PubMed](#)]
71. Wong, C.J.; Casper, R.F.; Rogers, I.M. Epigenetic changes to human umbilical cord blood cells cultured with three proteins indicate partial reprogramming to a pluripotent state. *Exp. Cell Res.* **2010**, *316*, 927–939. [[CrossRef](#)] [[PubMed](#)]
72. Zhang, Y.; Chen, H. Genistein attenuates WNT signaling by up-regulating sFRP2 in a human colon cancer cell line. *Exp. Biol. Med.* **2011**, *236*, 714–722. [[CrossRef](#)] [[PubMed](#)]
73. Gregorieff, A.; Clevers, H. Wnt signaling in the intestinal epithelium: From endoderm to cancer. *Genes Dev.* **2005**, *19*, 877–890. [[CrossRef](#)] [[PubMed](#)]
74. MacDonald, B.T.; Tamai, K.; He, X. Wnt/ β -catenin signaling: Components, mechanisms, and diseases. *Dev. Cell* **2009**, *17*, 9–26. [[CrossRef](#)] [[PubMed](#)]
75. Qi, J.; Zhu, Y.-Q.; Luo, J.; Tao, W.-H. Hypermethylation and expression regulation of secreted frizzled-related protein genes in colorectal tumor. *World J. Gastroenterol.* **2006**, *12*, 7113–7117. [[CrossRef](#)] [[PubMed](#)]
76. Wang, H.; Li, Q.; Chen, H. Genistein affects histone modifications on Dickkopf-related protein 1 (DKK1) gene in SW480 human colon cancer cell line. *PLoS ONE* **2012**, *7*, e40955. [[CrossRef](#)] [[PubMed](#)]
77. Rawson, J.B.; Manno, M.; Mrkonjic, M.; Daftary, D.; Dicks, E.; Buchanan, D.D.; Younghusband, H.B.; Parfrey, P.S.; Young, J.P.; Pollett, A.; et al. Promoter methylation of Wnt antagonists DKK1 and SFRP1 is associated with opposing tumor subtypes in two large populations of colorectal cancer patients. *Carcinogenesis* **2011**, *32*, 741–747. [[CrossRef](#)] [[PubMed](#)]
78. Aguilera, O.; Fraga, M.F.; Ballestar, E.; Paz, M.F.; Herranz, M.; Espada, J.; García, J.M.; Muñoz, A.; Esteller, M.; González-Sancho, J.M. Epigenetic inactivation of the Wnt antagonist *DICKKOPF-1* (*DKK-1*) gene in human colorectal cancer. *Oncogene* **2006**, *25*, 4116–4121. [[CrossRef](#)] [[PubMed](#)]
79. Hirata, H.; Hinoda, Y.; Nakajima, K.; Kawamoto, K.; Kikuno, N.; Ueno, K.; Yamamura, S.; Zaman, M.S.; Khatri, G.; Chen, Y.; et al. Wnt antagonist *DKK1* acts as a tumor suppressor gene that induces apoptosis and inhibits proliferation in human renal cell carcinoma. *Int. J. Cancer* **2011**, *128*, 1793–1803. [[CrossRef](#)] [[PubMed](#)]
80. Ravindranath, M.H.; Muthugounder, S.; Presser, N.; Viswanathan, S. Anticancer therapeutic potential of soy isoflavone, genistein. *Adv. Exp. Med. Biol.* **2004**, *546*, 121–165. [[PubMed](#)]
81. Mahmoud, A.M.; Al-Alem, U.; Ali, M.M.; Bosland, M.C. Genistein increases estrogen receptor β expression in prostate cancer via reducing its promoter methylation. *J. Steroid Biochem. Mol. Biol.* **2015**, *152*, 62–75. [[CrossRef](#)] [[PubMed](#)]
82. Anastasiou, N.; Boston, S.; Lacey, M.; Storing, N.; Whitehead, S.A. Evidence that low-dose, long-term genistein treatment inhibits oestradiol-stimulated growth in MCF-7 cells by down-regulation of the PI3-kinase/Akt signalling pathway. *J. Steroid Biochem. Mol. Biol.* **2009**, *116*, 50–55. [[CrossRef](#)] [[PubMed](#)]
83. Jawaid, K.; Crane, S.R.; Nowers, J.L.; Lacey, M.; Whitehead, S.A. Long-term genistein treatment of MCF-7 cells decreases acetylated histone 3 expression and alters growth responses to mitogens and histone deacetylase inhibitors. *J. Steroid Biochem. Mol. Biol.* **2010**, *120*, 164–171. [[CrossRef](#)] [[PubMed](#)]
84. Whitten, P.L.; Patisaul, H.B. Cross-species and interassay comparisons of phytoestrogen action. *Environ. Health Perspect.* **2001**. [[CrossRef](#)]
85. Soukup, S.T.; Al-Maharik, N.; Botting, N.; Kulling, S.E. Quantification of soy isoflavones and their conjugative metabolites in plasma and urine: An automated and validated UHPLC-MS/MS method for use in large-scale studies. *Anal. Bioanal. Chem.* **2014**, *406*, 6007–6020. [[CrossRef](#)] [[PubMed](#)]
86. Lee, S.-A.; Shu, X.-O.; Li, H.; Yang, G.; Cai, H.; Wen, W.; Ji, B.-T.; Gao, J.; Gao, Y.-T.; Zheng, W. Adolescent and adult soy food intake and breast cancer risk: Results from the shanghai women's health study. *Am. J. Clin. Nutr.* **2009**, *89*, 1920–1926. [[CrossRef](#)] [[PubMed](#)]
87. Barnes, S. The chemopreventive properties of soy isoflavonoids in animal models of breast cancer. *Breast Cancer Res. Treat.* **1997**, *46*, 169–179. [[CrossRef](#)] [[PubMed](#)]

88. Adlercreutz, H. Lignans and human health. *Crit. Rev. Clin. Lab. Sci.* **2007**, *44*, 483–525. [[CrossRef](#)] [[PubMed](#)]
89. Hamilton-Reeves, J.M.; Rebello, S.A.; Thomas, W.; Slaton, J.W.; Kurzer, M.S. Isoflavone-rich soy protein isolate suppresses androgen receptor expression without altering estrogen receptor- β expression or serum hormonal profiles in men at high risk of prostate cancer. *J. Nutr.* **2007**, *137*, 1769–1775. [[PubMed](#)]
90. Pendleton, J.M.; Tan, W.W.; Anai, S.; Chang, M.; Hou, W.; Shiverick, K.T.; Rosser, C.J. Phase II trial of isoflavone in prostate-specific antigen recurrent prostate cancer after previous local therapy. *BMC Cancer* **2008**, *8*, 132. [[CrossRef](#)] [[PubMed](#)]
91. Goetzl, M.A.; Van Veldhuizen, P.J.; Thrasher, J.B. Effects of soy phytoestrogens on the prostate. *Prostate Cancer Prostatic Dis.* **2007**, *10*, 216–223. [[CrossRef](#)] [[PubMed](#)]
92. Adams, K.F.; Chen, C.; Newton, K.M.; Potter, J.D.; Lampe, J.W. Soy isoflavones do not modulate prostate-specific antigen concentrations in older men in a randomized controlled trial. *Cancer Epidemiol. Biomark. Prev.* **2004**, *13*, 644–648.
93. Mahmoud, A.M.; Yang, W.; Bosland, M.C. Soy isoflavones and prostate cancer: A review of molecular mechanisms. *J. Steroid Biochem. Mol. Biol.* **2014**, *140*, 116–132. [[CrossRef](#)] [[PubMed](#)]
94. Munro, I.C.; Harwood, M.; Hlywka, J.J.; Stephen, A.M.; Doull, J.; Flamm, W.G.; Adlercreutz, H. Soy isoflavones: A safety review. *Nutr. Rev.* **2003**, *61*, 1–33. [[CrossRef](#)] [[PubMed](#)]
95. He, F.-J.; Chen, J.-Q. Consumption of soybean, soy foods, soy isoflavones and breast cancer incidence: Differences between Chinese women and women in Western countries and possible mechanisms. *Food Sci. Hum. Wellness* **2013**, *2*, 146–161. [[CrossRef](#)]
96. Ju, Y.H.; Carlson, K.E.; Sun, J.; Pathak, D.; Katzenellenbogen, B.S.; Katzenellenbogen, J.A.; Helferich, W.G. Estrogenic effects of extracts from cabbage, fermented cabbage, and acidified brussels sprouts on growth and gene expression of estrogen-dependent human breast cancer (MCF-7) cells. *J. Agric. Food Chem.* **2000**, *48*, 4628–4634. [[CrossRef](#)] [[PubMed](#)]
97. Breinholt, V.; Hossaini, A.; Svendsen, G.W.; Brouwer, C.; Nielsen, E. Estrogenic activity of flavonoids in mice. The importance of estrogen receptor distribution, metabolism and bioavailability. *Food Chem. Toxicol.* **2000**, *38*, 555–564. [[CrossRef](#)]
98. Owens, W.; Ashby, J.; Odum, J.; Onyon, L. The OECD program to validate the rat uterotrophic bioassay. Phase 2: Dietary phytoestrogen analyses. *Environ. Health Perspect.* **2003**, *111*, 1559–1567. [[CrossRef](#)] [[PubMed](#)]
99. Phrakonkham, P.; Chevalier, J.; Desmetz, C.; Pinnert, M.-F.; Bergès, R.; Jover, E.; Davicco, M.-J.; Bennetau-Pelissero, C.; Coxam, V.; Artur, Y.; et al. Isoflavonoid-based bone-sparing treatments exert a low activity on reproductive organs and on hepatic metabolism of estradiol in ovariectomized rats. *Toxicol. Appl. Pharmacol.* **2007**, *224*, 105–115. [[CrossRef](#)] [[PubMed](#)]
100. Cederroth, C.R.; Zimmermann, C.; Nef, S. Soy, phytoestrogens and their impact on reproductive health. *Mol. Cell. Endocrinol.* **2012**, *355*, 192–200. [[CrossRef](#)] [[PubMed](#)]
101. Jefferson, W.N.; Padilla-Banks, E.; Clark, G.; Newbold, R.R. Assessing estrogenic activity of phytochemicals using transcriptional activation and immature mouse uterotrophic responses. *J. Chromatogr. B Anal. Technol. Biomed. Life Sci.* **2002**, *777*, 179–189. [[CrossRef](#)]
102. Inudo, M.; Ishibashi, H.; Matsumura, N.; Matsuoka, M.; Mori, T.; Taniyama, S.; Kadokami, K.; Koga, M.; Shinohara, R.; Hutchinson, T.H.; et al. Effect of estrogenic activity, and phytoestrogen and organochlorine pesticide contents in an experimental fish diet on reproduction and hepatic vitellogenin production in medaka (*Oryzias latipes*). *Comp. Med.* **2004**, *54*, 673–680. [[PubMed](#)]
103. Kobayashi, M.; Ishibashi, H.; Moriwaki, T.; Koshiishi, T.; Ogawa, S.; Matsumoto, T.; Arizono, K.; Watabe, S. Production of low-estrogen goldfish diet for in vivo endocrine disrupter test. *Environ. Sci.* **2006**, *13*, 125–136. [[PubMed](#)]
104. Bagheri, T.; Imanpoor, M.R.; Jafari, V.; Bennetau-Pelissero, C. Reproductive impairment and endocrine disruption in goldfish by feeding diets containing soybean meal. *Anim. Reprod. Sci.* **2013**, *139*, 136–144. [[CrossRef](#)] [[PubMed](#)]
105. Tan, K.A. L.; Walker, M.; Morris, K.; Greig, I.; Mason, J.I.; Sharpe, R.M. Infant feeding with soy formula milk: Effects on puberty progression, reproductive function and testicular cell numbers in marmoset monkeys in adulthood. *Hum. Reprod.* **2006**, *21*, 896–904. [[CrossRef](#)] [[PubMed](#)]

106. Adachi, T.; Ono, Y.; Koh, K.B.; Takashima, K.; Tainaka, H.; Matsuno, Y.; Nakagawa, S.; Todaka, E.; Sakurai, K.; Fukata, H.; et al. Long-term alteration of gene expression without morphological change in testis after neonatal exposure to genistein in mice: Toxicogenomic analysis using cDNA microarray. *Food Chem. Toxicol.* **2004**, *42*, 445–452. [[CrossRef](#)] [[PubMed](#)]
107. Persky, R.W.; Turtzo, L.C.; McCullough, L.D. Stroke in women: Disparities and outcomes. *Curr. Cardiol. Rep.* **2010**, *12*, 6–13. [[CrossRef](#)] [[PubMed](#)]
108. Wuttke, W.; Jarry, H.; Westphalen, S.; Christoffel, V.; Seidlová-Wuttke, D. Phytoestrogens for hormone replacement therapy? *J. Steroid Biochem. Mol. Biol.* **2002**, *83*, 133–147. [[CrossRef](#)]
109. Sehmis, S.; Uffenorde, J.; Maehlmeyer, S.; Tezval, M.; Jarry, H.; Stuermer, K.M.; Stuermer, E.K. Evaluation of bone quality and quantity in osteoporotic mice—the effects of genistein and equol. *Phytomedicine* **2010**, *17*, 424–430. [[CrossRef](#)] [[PubMed](#)]
110. Raghu Nadhanan, R.; Skinner, J.; Chung, R.; Su, Y.-W.; Howe, P.R.; Xian, C.J. Supplementation with fish oil and genistein, individually or in combination, protects bone against the adverse effects of methotrexate chemotherapy in rats. *PLoS ONE* **2013**, *8*, e71592. [[CrossRef](#)] [[PubMed](#)]
111. Canal Castro, C.; Pagnussat, A.S.; Orlandi, L.; Worm, P.; Moura, N.; Etgen, A.M.; Alexandre Netto, C. Coumestrol has neuroprotective effects before and after global cerebral ischemia in female rats. *Brain Res.* **2012**, *1474*, 82–90. [[CrossRef](#)] [[PubMed](#)]
112. Anway, M.D.; Leathers, C.; Skinner, M.K. Endocrine disruptor vinclozolin induced epigenetic transgenerational adult-onset disease. *Endocrinology* **2006**, *147*, 5515–5523. [[CrossRef](#)] [[PubMed](#)]
113. Jirtle, R.L.; Skinner, M.K. Environmental epigenomics and disease susceptibility. *Nat. Rev. Genet.* **2007**, *8*, 253–262. [[CrossRef](#)] [[PubMed](#)]
114. Salian, S.; Doshi, T.; Vanage, G. Impairment in protein expression profile of testicular steroid receptor coregulators in male rat offspring perinatally exposed to Bisphenol A. *Life Sci.* **2009**, *85*, 11–18. [[CrossRef](#)] [[PubMed](#)]
115. Guerrero-Bosagna, C.M.; Skinner, M.K. Environmental epigenetics and phytoestrogen/phytochemical exposures. *J. Steroid Biochem. Mol. Biol.* **2014**, *139*, 270–276. [[CrossRef](#)] [[PubMed](#)]
116. Bruner-Tran, K.L.; Osteen, K.G. Developmental exposure to TCDD reduces fertility and negatively affects pregnancy outcomes across multiple generations. *Reprod. Toxicol.* **2011**, *31*, 344–350. [[CrossRef](#)] [[PubMed](#)]
117. Manikkam, M.; Tracey, R.; Guerrero-Bosagna, C.; Skinner, M.K. Dioxin (TCDD) induces epigenetic transgenerational inheritance of adult onset disease and sperm epimutations. *PLoS ONE* **2012**, *7*, e46249. [[CrossRef](#)] [[PubMed](#)]
118. Hao, C.; Gely-Pernot, A.; Kervarrec, C.; Boudjema, M.; Becker, E.; Khil, P.; Tevosian, S.; Jégou, B.; Smagulova, F. Exposure to the widely used herbicide atrazine results in deregulation of global tissue-specific RNA transcription in the third generation and is associated with a global decrease of histone trimethylation in mice. *Nucleic Acids Res.* **2016**. [[CrossRef](#)] [[PubMed](#)]
119. Sondergaard, T.E.; Hansen, F.T.; Purup, S.; Nielsen, A.K.; Bonfeld-Jørgensen, E.C.; Giese, H.; Sørensen, J.L. Fusarin C acts like an estrogenic agonist and stimulates breast cancer cells in vitro. *Toxicol. Lett.* **2011**, *205*, 116–121. [[CrossRef](#)] [[PubMed](#)]
120. Khosrokhavar, R.; Rahimifard, N.; Shoeibi, S.; Hamedani, M.P.; Hosseini, M.-J. Effects of zearalenone and α -Zearalenol in comparison with Raloxifene on T47D cells. *Toxicol. Mech. Methods* **2009**, *19*, 246–250. [[CrossRef](#)] [[PubMed](#)]
121. Parveen, M.; Zhu, Y.; Kiyama, R. Expression profiling of the genes responding to zearalenone and its analogues using estrogen-responsive genes. *FEBS Lett.* **2009**, *583*, 2377–2384. [[CrossRef](#)] [[PubMed](#)]
122. Belli, P.; Bellaton, C.; Durand, J.; Balleydier, S.; Milhau, N.; Mure, M.; Mornex, J.-F.; Benahmed, M.; Le Jan, C. Fetal and neonatal exposure to the mycotoxin zearalenone induces phenotypic alterations in adult rat mammary gland. *Food Chem. Toxicol.* **2010**, *48*, 2818–2826. [[CrossRef](#)] [[PubMed](#)]
123. Hsieh, C.-Y.; Santell, R.C.; Haslam, S.Z.; Helferich, W.G. Estrogenic effects of genistein on the growth of estrogen receptor-positive human breast cancer (MCF-7) cells in vitro and in vivo. *Cancer Res.* **1998**, *58*, 3833–3838. [[PubMed](#)]
124. Wang, T.T. Y.; Sathyamoorthy, N.; Phang, J.M. Molecular effects of genistein on estrogen receptor mediated pathways. *Carcinogenesis* **1996**, *17*, 271–275. [[CrossRef](#)] [[PubMed](#)]
125. Rahman, H.P.; Hofland, J.; Foster, P.A. In touch with your feminine side: How oestrogen metabolism impacts prostate cancer. *Endocr. Relat. Cancer* **2016**, *23*, R249–R266. [[CrossRef](#)] [[PubMed](#)]

126. Majid, S.; Dar, A.A.; Ahmad, A.E.; Hirata, H.; Kawakami, K.; Shahryari, V.; Saini, S.; Tanaka, Y.; Dahiya, A.V.; Khatri, G.; et al. *BTG3* tumor suppressor gene promoter demethylation, histone modification and cell cycle arrest by genistein in renal cancer. *Carcinogenesis* **2009**, *30*, 662–670. [[CrossRef](#)] [[PubMed](#)]
127. Vardi, A.; Bosviel, R.; Rabiau, N.; Adjakly, M.; Satih, S.; Dechelotte, P.; Boiteux, J.-P.; Fontana, L.; Bignon, Y.-J.; Guy, L.; et al. Soy phytoestrogens modify DNA methylation of *GSTP1*, *RASSF1A*, *EPH2* and *BRCA1* promoter in prostate cancer cells. *In Vivo* **2010**, *24*, 393–400. [[PubMed](#)]
128. Lim, W.; Jeong, W.; Song, G. Coumestrol suppresses proliferation of ES2 human epithelial ovarian cancer cells. *J. Endocrinol.* **2016**, *228*, 149–160. [[CrossRef](#)] [[PubMed](#)]
129. Leclercq, G.; Jacquot, Y. Interactions of isoflavones and other plant derived estrogens with estrogen receptors for prevention and treatment of breast cancer—considerations concerning related efficacy and safety. *J. Steroid Biochem. Mol. Biol.* **2014**, *139*, 237–244. [[CrossRef](#)] [[PubMed](#)]
130. Li, Z.; Zhang, Y.; Hedman, A.C.; Ames, J.B.; Sacks, D.B. Calmodulin lobes facilitate dimerization and activation of estrogen receptor- α . *J. Biol. Chem.* **2017**, *292*, 4614–4622. [[CrossRef](#)] [[PubMed](#)]
131. Li, F.; Ye, L.; Lin, S.; Leung, L.K. Dietary flavones and flavonones display differential effects on aromatase (CYP19) transcription in the breast cancer cells MCF-7. *Mol. Cell. Endocrinol.* **2011**, *344*, 51–58. [[CrossRef](#)] [[PubMed](#)]
132. Nakamura, K.; Yang, J.-H.; Sato, E.; Miura, N.; Wu, Y.-X. Effects of hydroxy groups in the A-ring on the anti-proteasome activity of flavone. *Biol. Pharm. Bull.* **2015**, *38*, 935–940. [[CrossRef](#)] [[PubMed](#)]
133. Marcotte, R.; Sayad, A.; Brown, K.R.; Sanchez-Garcia, F.; Reimand, J.; Haider, M.; Virtanen, C.; Bradner, J.E.; Bader, G.D.; Mills, G.B.; et al. Functional genomic landscape of human breast cancer drivers, vulnerabilities, and resistance. *Cell* **2016**, *164*, 293–309. [[CrossRef](#)] [[PubMed](#)]
134. Singh, V.; Sharma, V.; Verma, V.; Pandey, D.; Yadav, S.K.; Maikhuri, J.P.; Gupta, G. Apigenin manipulates the ubiquitin–proteasome system to rescue estrogen receptor- β from degradation and induce apoptosis in prostate cancer cells. *Eur. J. Nutr.* **2015**, *54*, 1255–1267. [[CrossRef](#)] [[PubMed](#)]
135. Lecomte, S.; Lelong, M.; Bourguin, G.; Efstathiou, T.; Saligaut, C.; Pakdel, F. Assessment of the potential activity of major dietary compounds as selective estrogen receptor modulators in two distinct cell models for proliferation and differentiation. *Toxicol. Appl. Pharmacol.* **2017**, *325*, 61–70. [[CrossRef](#)] [[PubMed](#)]
136. Prossnitz, E.R.; Barton, M. The G-protein-coupled estrogen receptor GPER in health and disease. *Nat. Rev. Endocrinol.* **2011**, *7*, 715–726. [[CrossRef](#)] [[PubMed](#)]
137. Chan, Q.K.Y.; Lam, H.-M.; Ng, C.-F.; Lee, A.Y.Y.; Chan, E.S.Y.; Ng, H.-K.; Ho, S.-M.; Lau, K.-M. Activation of GPR30 inhibits the growth of prostate cancer cells through sustained activation of Erk1/2, c-jun/c-fos-dependent upregulation of p21, and induction of G(2) cell-cycle arrest. *Cell Death Differ.* **2010**, *17*, 1511–1523. [[CrossRef](#)] [[PubMed](#)]

