

AminoMethylPhosphonic acid (AMPA) in natural waters: Its sources, behavior and environmental fate

Alexis Grandcoin, Stéphanie Piel, Estelle Baurès

▶ To cite this version:

Alexis Grandcoin, Stéphanie Piel, Estelle Baurès. AminoMethylPhosphonic acid (AMPA) in natural waters: Its sources, behavior and environmental fate. Water Research, 2017, 117, pp.187-197. 10.1016/j.watres.2017.03.055 . hal-01516026

HAL Id: hal-01516026 https://univ-rennes.hal.science/hal-01516026

Submitted on 5 May 2017

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Accepted Manuscript

AminoMethylPhosphonic acid (AMPA) in natural waters: Its sources, behavior and environmental fate

Alexis Grandcoin, Stéphanie Piel, Estelle Baures

PII: S0043-1354(17)30245-2

DOI: 10.1016/j.watres.2017.03.055

Reference: WR 12792

To appear in: Water Research

Received Date: 22 December 2016

Revised Date: 24 March 2017

Accepted Date: 26 March 2017

Please cite this article as: Grandcoin, A., Piel, S., Baures, E., AminoMethylPhosphonic acid (AMPA) in natural waters: Its sources, behavior and environmental fate, *Water Research* (2017), doi: 10.1016/j.watres.2017.03.055.

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.





1 AminoMethylPhosphonic Acid (AMPA) in natural waters: its

2 sources, behavior and environmental fate

- 3 Alexis Grandcoin^{1, 2, 3}, Stéphanie Piel³ and Estelle Baures^{1, 2}
- 4 ¹ Environment and Health Research laboratory (LERES), EHESP School of Public Health,
- 5 Avenue du Professeur Léon Bernard-CS 74312. Rennes Cedex 35043, France
- 6 ² Inserm, U 1085 Institute of Research in Environmental and Occupational Health (IRSET),
- 7 Avenue du Professeur Léon Bernard-CS 74312. Rennes Cedex 35043, France
- ³ SAUR Research and Development, 2 rue de la Bresle, Maurepas 78310, France
- 9
- 10 E-Mails: <u>Alexis.grandcoin@ehesp.fr;</u> Estelle.baures@ehesp.fr
- 11

12 Abstract:

The widely occurring degradation product aminomethylphosphonic acid (AMPA) is a result of glyphosate and amino-polyphosphonate degradation. Massive use of the parent compounds leads to the ubiquity of AMPA in the environment, and particularly in water.

16 The purpose of this review is to summarize and discuss current insights into AMPA 17 formation, transport, persistence and toxicity.

In agricultural soils, AMPA is concentrated in the topsoil, and degrades slowly in most soils. It 18 19 can reach shallow groundwater, but rarely managed to enter deep groundwater. AMPA is 20 strongly adsorbed to soil particles and moves with the particles towards the stream in rainfall runoff. In urban areas, AMPA comes from phosphonates and glyphosate in wastewater. It is 21 22 commonly found at the outlets of Wastewater Treatment Plants (WWTP). Sediments tend to accumulate AMPA, where it may be biodegraded. Airborne AMPA is not negligible, but does 23 wash-out with heavy rainfall. AMPA is reported to be persistent and can be biologically 24 degraded in soils and sediments. Limited photodegradation in waters exists. AMPA mainly 25 26 has its sources in agricultural leachates, and urban wastewater effluents. The domestic contribution to urban loads is negligible. 27

There is a critical lack of epidemiological data - especially on water exposure - to understand the toxicological effects, if any, of AMPA on humans. Fortunately, well operated water treatment plants remove a significant proportion of the AMPA from water, even though there are not sufficient regulatory limits for metabolites.

Keywords: AminoMethylPhosphonic Acid (AMPA); Glyphosate; Metabolite;
 Sources; Watershed

35 1. Introduction

36

Aminomethylphosphonic acid (AMPA) is a degradation product resulting from phosphonate degradation (Nowack, 2003; Wang et al., 2016). It can be a metabolite of glyphosate microbial degradation in soils (Borggaard and Gimsing, 2008), and a photodegradation product of amino-polyphosphonates in water (Lesueur et al., 2005).

AMPA has been reported to occur widely in the air of agricultural areas (Chang et al., 2011; Battaglin et al., 2014), surface waters (Coupe et al., 2012; Scribner et al., 2007), sediments (Ronco et al., 2016) and shallow groundwater at depths within 2 m of the edge of the streams (Van Stempvoort et al., 2014), as well as less frequently in deep groundwater (Battaglin et al., 2014). Struger et al. (2015) pointed out the uncertainty concerning the respective proportions of AMPA sources, which can be agricultural, industrial, and urban.

Evidence has been found that the sources of AMPA, in a wastewater-influenced stream,
differ depending on hydroclimatic conditions (Botta et al., 2009). Industrial and domestic
phosphonates are an alternative source of AMPA.

50 This paper begins with a review of the current state of knowledge on AMPA sources. Next, it 51 presents results on AMPA co-transport through the various watershed compartments. This is 52 followed by a summary of existing data on degradation and persistence, and we conclude 53 with an update on the known toxicity of AMPA, protection regulations in water, and the state 54 of the art concerning AMPA in water treatment.

55

56 2. Sources of AMPA

58 2.1. Phosphonates

59

2.1.1. Origins and spatial dispersion

61

60

Phosphonates are anions of phosphonic acids occurring in environmental pH conditions and
presenting high polarity and high water solubility (Schmidt et al., 2013).

Phosphonates (and especially amino-methylene-polyphosphonates) are commonly used in both industrial and household applications in industrialized countries (Nowack, 2003). Amino polyphosphonates contain several phosphonic acid groups and at least one amine group. They are detergents, fire retardants, anticorrosives and anti-scaling agents, and in the textile industry they are used as complexing agents (Studnik et al., 2015; Nowack, 2003). According to Studnik et al. (2015) water treatment is the main application of amino-methylenephosphonates, which are used as membrane anti-fouling agents.

This flexibility of purpose means that they are both useful, and produced. Gledhill and Feijtel 71 72 (1992) addressed annual consumption levels of phosphonates in Europe and the USA: the 73 most popular are nitrilotris-methylenephosphonic acid (NTMP) with 3820 tons in the USA and 3360 tons in Europe; diethylenetriaminepenta-methylenephosphonic acid (DTPMP) with 275 74 tons in the USA and 5270 tons in Europe; ethylene diamine-tetra-methylenephosphonic acid 75 76 (EDTMP) with 0 tons in the USA and 1090 tons in Europe; hexaethylenediamine-tetramethylenephosphonic acid (HDTMP) with 180 tons in the USA and 90 tons in Europe; and 1-77 78 Hydroxyethane 1,1-diphosphonic acid (HEDP) with 5270 tons in the USA and 2010 tons in Europe. Glyphosate (N-phosphono-methylglycine) is also an economically significant 79 80 phosphonate, - this subject is tackled at § 2.1.1. Nowack (2004) later reported that worldwide consumption of phosphonates in 1998 was 56,000 tons, and European consumption was 81 82 16,000 tons in 1999. These data may seem outdated, but to the best of our knowledge, these 83 are the most recent global studies on phosphonate consumption in industrialized countries.

Phosphonate chelating agents are used more and more each year in a wide range of applications (Nowack and Stone, 2002), and some countries, such as France (2007) and the USA (2010) have banned or limited phosphates in domestic detergents (European Parliament, 2012), offering yet another application to phosphonates (Studnik et al., 2015).

In the UK, phosphonates are currently present in 81% of laundry products and 4% of dishwasher products, at a rate of 2.5% phosphonates by weight (Comber et al., 2013). Their fate is not well- documented, mainly for want of a sensitive and specific analytical method. This lack of adapted analytical method is caused by the ionic and polar nature of phosphonates (Schmidt et al., 2013).

With his previous ion-pair liquid chromatography method development (Nowack 1997b), 93 Nowack (1997a) spelled out the behavior of major phosphonates in the WWTP, showing that 94 phosphonates were present in influents at concentrations of between 21-254 µg/L for NTMP, 95 31-65 µg/L for EDTMP, and 75-974 µg/L for DTPMP. The effluent concentrations were below 96 detection limits, e.g. 15 µg/L for NTMP, 22 µg/L for EDTMP, and 29 µg/L for DTPMP. 97 Estimated elimination rates were at least 70%. Phosphonates are concentrated in sludge 98 rather than degraded (Figure 1); this raises an issue of pollution transport if sludge is used as 99 100 fertilizer.

101 Phosphonates can be released into the environment through sewer outfall or WWTP.

102

103

Figure 1: please insert here

104

105 2.1.2. Phosphonate degradation pathways

106

107 Biodegradation

108 Because of their strong C-P bond, phosphonates are difficult to degrade. This results in a prolonged lifetime and accumulation in water environments (Forlani et al., 2011). Phosphates 109 110 are preferentially used by microbial fauna as a phosphorus source. Consequently phosphonates are poorly consumed and degraded under natural conditions (Hsieh and 111 Wanner, 2010; Nowack, 2003). Several microorganisms have however been identified as 112 being able to use phosphonates as their only source of phosphorus: Bacillus megaterium, 113 Pseudomonas aeruginosa, Agrobacterium radiobacter, Escherichia coli, Pseudomonas 114 115 stutzeri and Streptomyces morookaensis (Obojska and Lejczak, 2003; Kononova and Nesmeyanova, 2002; Matys et al., 2004). 116

Furthermore, Forlani et al. (2011) successfully used *Spirulina* strains to remove 50% of initial
polyphosphonate pollution from wastewater; although degradation was incomplete, and
potential metabolites are unknown.

120

121 Physical and chemical degradation

Thanks to Iron-phosphonate complexes, the main phosphonate degradation pathway under 122 123 natural conditions is photodegradation, which enables metal-catalyzed photodegradation (Nowack, 2003). According to the review by Jaworska et al. (2002), AMPA is the main 124 125 degradation product formed by degradation of NTMP, EDTMP, DTPMP. Lesueur et al. (2005) showed that phosphonates are converted mainly into orthophosphate (93%) and to a 126 lesser extent into AMPA (7%) under UV light in an aquatic environment. This conversion is 127 enhanced in the presence of Iron and an acidic pH (Figure 2). AMPA is eventually converted 128 129 to orthophosphate.

130

131

Figure 2: Please insert here

The fate and behavior of phosphonates in the environment are poorly described due to the lack of a sensitive, specific and reliable method for their determination in natural systems (Lesueur et al., 2005).

136

137 2.2. Glyphosate

- 138
- 139 2.2.1. Origins and spatial dispersion

140

141 Agricultural

Glyphosate is the most widely used herbicide in the world, with production of 620,000 tons in 2008 (Bøhn et al., 2014), increasing to 825,000 tons in 2014 (Benbrook, 2016). Introduced in 1974, glyphosate is now extensively used with genetically modified herbicide-tolerant crops; it is also commonly used for non-agricultural areas (Scribner et al., 2007).

146 Figure 1 shows the fate of glyphosate in a watershed. Applied glyphosate is initially concentrated in topsoil layers: Yang et al. (2015b) conducted an experiment in an artificial 147 rain-simulation facility on bare soil. They showed that 72% of the applied herbicide is found 148 at a depth of between 0 and 2 cm in clay loam soil; Okada et al. (2016) and Rampazzo et al. 149 (2013) made the same observations for silty clay loam soils and silty loam soils. Glyphosate 150 sorption in soils is broadly described as strong. Okada et al. (2016) proved that adsorption is 151 related to soil clay content, CEC (Cation Exchange Capacity), and negatively related to pH 152 153 and phosphorus.

154 Conversely, Sidoli et al. (2015), Ololade et al. (2014), and Lupi et al. (2015) found that soil 155 pH_{CaCl2} and organic carbon (OC) regulate glyphosate behavior in soils. Furthermore, 156 glyphosate is sorbed onto iron and aluminum oxides (Borggaard and Gimsing, 2008, 157 Vereecken, 2005, Sidoli et al., 2015, Rampazzo et al., 2013), while sorption onto silicates is

limited and soil organic matter (OM), while not itself sorbing glyphosate, does affect glyphosate sorption by stabilizing oxides with high sorption capacity, according to Borggaard and Gimsing (2008). For Candela et al. (2010) glyphosate adsorption is mainly a kinetic process, influenced by pore water velocity and residence time of soil solutions. Surface complexation and precipitation can happen given slow water speed and lengthy residence time. If water velocities are slow and enough time is allowed to interact with the soil matrix, surface complexation and precipitation takes place.

Phosphate and glyphosate are chemically similar, and although there could be competition for sorption sites, phosphates overcome glyphosate on sorption sites in only a few soils and minerals. Kanissery et al. (2015) have shown that phosphate addition in clay loam soil decreases glyphosate sorption. However this phenomenon is actually limited or absent for most soils (Borggaard, 2011).

Where strong sorption is demonstrated, glyphosate accumulation in soils can be expected. La Jeunesse et al. (2015) confirmed that a large amount of glyphosate is stored in a onceper-year application vineyard catchment and released throughout the year with heavy rainfall, despite rapid glyphosate degradation in soils, with a half-life of 7-130 days on site.

174

175 Non-agricultural & WWTP

Glyphosate use is not confined to agricultural sources. Botta et al. (2009) showed that contamination of the Orge basin (France) is of urban origin - roads and railways being the main sources of glyphosate in their study. In addition, Torstensson et al. (2005) have shown railway maintenance to be a major source of glyphosate contamination in Sweden, because glyphosate residues were found in railway drainage and nearby groundwater.

Lastly, the urban contribution to glyphosate load in surface waters has been studied in recent years. In France, Blanchoud et al. (2007) reported that urban runoff flows directly towards

rivers via separate sewer systems (initially constructed to limit storm water overflow), and this contributes to the transfer of pollution to surface waters. Connor et al. (2007) showed that runoff from small-urbanized tributaries may contribute as much or more to the pesticide loads than runoff from the agricultural areas, in San Francisco Bay. Glyphosate is ubiquitous at French storm sewer outlets: Zgheib et al. (2012) found glyphosate in water (dissolved plus particulate phase) with 93% detection frequency.

189 In addition to these general statements, few authors have gained more specific glyphosate behavior knowledge. A small catchment in Switzerland (25 km²), studied by Hanke et al. 190 (2010) found glyphosate to leach with fast runoff from hard surfaces; glyphosate inputs to 191 192 WWTP, sewer system overflow and separate sewer systems account for 60% of glyphosate load in surface water. Kolpin et al. (2006) studied 10 effluent of WWTP across the USA. They 193 194 showed that these WWTP export glyphosate, increasing glyphosate content in the receiving stream by 100%. Following these initial observations, the origins of glyphosate in urban 195 196 effluents called for exploration. Ramwell et al. (2014) and Tang et al. (2015) thus shed light on the contribution of solely domestic usages to surface water drains, and less than 1% of 197 applied glyphosate was recovered in drains. They concluded that losses from similar and 198 European representative residential catchments (5.16 ha with 148 houses, England 199 200 (Ramwell et al., 2014) and 9.5 ha with 112 houses, Belgium (Tang et al., 2015)).were unlikely to contribute significantly to surface water load, in comparison with other urban 201 202 areas.

203

204

205

2.2.2. Glyphosate degradation pathways

206

207 Biodegradation

Glyphosate presents two major degradation pathways (Figure 3), one leading to the formation of AMPA and the other leading to the formation of sarcosine and glycine (Borggaard and Gimsing, 2008; Al-Rajab and Schiavon, 2010; Wang et al., 2016). Duke (2011) showed that glyphosate is already degraded to AMPA in crops and weed. The plant residues can also export AMPA in soils (Mamy et al., 2016).

- 213
- 214

Figure 3: please insert here

215

Glyphosate sorption in soils facilitates microbial degradation, considered the only degradation process in soils (Borggaard and Gimsing, 2008). This statement is confirmed by Kanissery et al. (2015) who did not observe any degradation in sterilized soils, regardless of oxygenation conditions.

220 Soil redox conditions seems to be significant factors in glyphosate degradation in soils, according to Kanissery et al. (2015), who established higher microbial degradation rates 221 under oxic soils than in anoxic soils; 53-63% of the ¹⁴C-glyphosate is mineralized as ¹⁴CO₂ in 222 56 days (oxic), whereas 38-41% of the ¹⁴C-glyphosate is mineralized in 56 days (anoxic). 223 Unexpectedly, soil phosphate addition stimulates degradation in anoxic soils despite 224 glyphosate desorption (see 2.2.1) (Kanissery et al., 2015). Glyphosate degradation rates in 225 soils are not linked to OM and clay content. Degradation rates are positively correlated to soil 226 pH and to a lesser extent to total metal concentrations (Kools et al., 2005). 227

228

Wang et al. (2016) investigated glyphosate fate in the water-sediment system, showing the major role played by sediments in degradation. The sarcosine pathway was the first to occur, associated with microbial growth, whereas the AMPA pathway occurred later under starvation conditions (lack of nutriments). Both pathways contribute to herbicide degradation

in the water-sediment system. Sviridov et al. (2015) listed the strains capable of degrading
glyphosate in different environments, mainly soils and waters. They also specify which
metabolite is mainly created by each microorganism, concluding that among the studied
bacterial strains, AMPA is produced most.

Table 1 summarizes the reported half-lives of glyphosate in soils, leading to the formation of AMPA through biological degradation. Under oxic conditions, glyphosate is degraded quickly in every soil, and more slowly under anoxic conditions (Kanissery et al., 2015). This confirms the importance of soil microflora in AMPA formation.

- 241
- 242

Table 1: please insert here

243

244 Physical and Chemical degradation

Chemical degradation and photodegradation are minor glyphosate degradation pathways 245 under natural conditions (Mallat and Barceló, 1998). Laboratory experiments proved that 246 247 glyphosate can be abiotically degraded in water, in the presence of manganese oxides. 0.005 g/L birnessite (δ-MnO₂) typically degrades up to 53% of 0.59 mmol/L of glyphosate in 248 50 hours (Barrett and McBride, 2005). Later Li et al. (2016) reported that 5 g/L birnessite (δ-249 250 MnO₂) typically degrades up to 70% of 3 mmol/L of glyphosate in 24 hours. Light increases the degradation rate without affecting the range of degradation. Surface mineral sorption 251 facilitates the formation of reactive complexes. According to Li et al. (2016) glyphosate 252 surface sorption is the key factor in influencing the rate and extent of degradation. Sorption of 253 degradation products onto catalytic surface sites in the mineral decreases both the rate and 254 255 extent of the reaction. Ascolani Yael et al. (2014) recently evidenced that AMPA is produced when glyphosate interacts with metallic ions (Cu²⁺) in an aqueous solution, showing the 256

existence of slight abiotic degradation of glyphosate by metals. Metallic ion concentration
used was 2.7 g/L, e.g. far from natural environment (World Health Organisation, 2004).

259

It has been demonstrated that AMPA may be derived from amino-polyphosphonates and 260 261 glyphosate. In Europe, which doesn't use GMO crops, there is currently no agreement as to the contribution of each source to AMPA contamination of natural waters. According to 262 Struger et al. (2015), occurrences of AMPA in Canadian streams are mainly due to 263 glyphosate applications in urban and rural areas (agricultural - row crop influenced - or 264 265 pristine forest land use) settings. This statement is built on positive correlations between AMPA and glyphosate occurrences, and a lack of positive correlations between AMPA and 266 the wastewater tracer acesulfame. 267

268 3. <u>AMPA and glyphosate transport across the watershed</u>

269

As a degradation product, AMPA usually co-occurs with glyphosate (Aparicio et al., 2013; Lupi et al., 2015). In a spatially wide occurrence study, Battaglin et al., (2014) showed that glyphosate is detected without AMPA in only 2.3% of 3,732 water and sediment samples, and that AMPA is detected without glyphosate in 17.9% of samples. Therefore, AMPA and glyphosate have been described together in the literature regarding their transport; in this paper, we will proceed in the same way.

276

3.1. Transport across soil

278

Glyphosate and AMPA transport can occur with favorable hydrological factors, where they are stable in soils, air or water. Van Stempvoort et al. (2016) showed that glyphosate and AMPA are persistent enough to reach shallow groundwater, be stored in it and then

transferred to surface water. Leaching of glyphosate and AMPA through soil seems to be a 282 slow, minor phenomenon, according to Bergström et al. (2011). This statement is confirmed 283 284 by Al-Rajab and Hakami (2014) who showed that less than 1% of the initially applied glyphosate leached in 2 months in a silty clay loam soil. Moreover Okada et al. (2016) report 285 that small amounts of glyphosate move quickly, but less than 0.24% of the applied 286 glyphosate leach beyond a depth of 15cm in agricultural silty clay loam and silty loam soils. 287 288 Nevertheless, Napoli et al. (2015) have demonstrated that both glyphosate and AMPA can leach through 100 cm of silty clay soil. As a percentage of applied glyphosate, 0.2% and 289 0.58% of glyphosate and AMPA respectively are recovered annually in lysimeters after 290 leaching through 1 m of silty clay soil. 291

292 Candela et al. (2010) found glyphosate traces at 1.9 m depth in a weathered granite soil, with 293 low content in OM and clay, but with Al and Fe oxides and hydroxides. In this granite soil, 294 AMPA is found to be less mobile than glyphosate. This suggests that, in poor soils with low 295 content of organic matter and clays in the soils, the herbicide may reach deep soil layers and 296 then groundwater.

Borggaard and Gimsing (2008) stressed the importance of preferential flows combined with heavy rainfalls shortly after application, for glyphosate and AMPA transport deeper into the soil profile.

Many authors have sought to measure the extent of mobility: Scribner et al. (2007) report glyphosate and AMPA occurrence in USA groundwater for 5.8% and 9.7% respectively of the 485 sites sampled with maximum concentrations of 0.67 μ g/L and 4.7 μ g/L. Later, Battaglin et al. (2014) repeated this type of occurrence study and found glyphosate and AMPA in 5.8% and 14.3% respectively of 1171 groundwater samples, with a maximum of 2.03 μ g/L and 4.88 μ g/L.

This insight supports the idea that glyphosate and AMPA have low mobility through soils, and should not represent a major threat for groundwater quality, with a few exceptions due to

308 geological context. AMPA seems to be a slightly more of a threat to groundwater quality than
309 its parent (Borggaard and Gimsing, 2008; Vereecken, 2005).

310

311 3.2. Transport towards surface water

312

As previously explained in paragraph 2.2.1., glyphosate is mainly stored in topsoil layers; this is true also of AMPA. Both can be exported mainly by runoff and by underground leaching from agricultural soils towards surface water, especially when rainfall occurs shortly after application (Yang et al., 2015b). Coupe et al. (2012) observed high levels of glyphosate and AMPA in surface water correlated with fresh application of herbicide. They concluded that maximal concentration in streams occurs with the first runoff episode after glyphosate application.

According to Yang et al. (2015a), Daouk et al. (2013) and La Jeunesse et al. (2015), rain 320 intensity is an essential factor for the offsite transport of glyphosate and AMPA; the 321 generated soil particle movement will carry adsorbed glyphosate and AMPA, which will end 322 323 up in surface water where it can be desorbed, degraded, or stored in sediments (Degenhardt et al., 2012; Todorovic-Rampazzo et al., 2014). Aparicio et al. (2013) conducted a study in 324 sixteen agricultural sites and forty-four streams in Argentinian agricultural basins, which were 325 sampled three times during 2012. They showed that AMPA is less influenced by this particle-326 affinity and is found in only 20% of suspended particulate matter samples, whereas 327 328 glyphosate is found in 67% of samples. Yang et al. (2015b) conducted their experiment in an artificial rain-simulation facility. They showed that up to 14% of applied glyphosate is 329 transported in runoff and suspended load from a clay loam soil with a rain intensity of 1 330 mm/min for 1 hour (extremes conditions of washing). This corroborates with Kjær et al. 331 (2011) who found that 13-16% of glyphosate leached due to a particle-facilitated transport 332 from a well-drained loamy field in Denmark (1.26 ha). The study were conducted in a 333

research field where the uppermost meter of the soil is heavily fractured and bioturbated, the water table is located 1–3 m b.g.s. (Lindhardt et al., 2001). Conversely, Norgaard et al. (2014) were unable to observe evidence of particle-facilitated transport of glyphosate and AMPA although they present a leaching risk chart based only on rainfall intensity and elapsed time after application.

With regard to non-agricultural soils, Tang et al. (2015) investigated glyphosate and AMPA 339 loss mechanisms in a Belgian residential area, with 1 stormwater outlet and during 13 340 events. The loss mechanisms are mainly governed by rainfall amount and intensity. Less 341 than 0.5% of initially applied glyphosate is recovered in storm drain outflow after 67 days. 342 343 When both glyphosate and AMPA are included in the total load, the loss rate is less than 1%. These low loss rates can be explained by the residential area's low runoff potential e.g. 344 345 runoff from unconnected or indirectly connected hard surfaces which can flow across pervious surfaces and infiltrate. 346

347

348 3.3. Transport in the atmosphere

349

Edge of agricultural field levels of glyphosate and AMPA in air and rain in the USA have been reported by Chang et al. (2011), who found glyphosate in both air and rain in the range of 60-100% of collected samples during two crop-growing seasons. AMPA was found in 40-90% of rain samples and 60-90% of air samples. Wind erosion and spray drift are the main sources for glyphosate and AMPA transport to the atmosphere. Several consecutive studies (condensed in Table 2) support these observations, and Battaglin et al. (2014) found glyphosate and AMPA in more than 70% of 85 rain samples collected at three distant sites.

Concentrations of glyphosate and AMPA reached a maximum of 9.1 ng/m³ and 0.97 ng/m³ respectively in air, and 2.5 μ g/L and 0.48 μ g/L respectively in rain. Weekly rain \geq 30 mm is efficient in removing the majority of glyphosate from the air (Chang et al., 2011).

360 Unfortunately, the proportion of applied glyphosate transmitted to the atmosphere is not 361 known.

362 Despite their low vapor pressure and strong adsorptive trend (Battaglin et al., 2014), 363 glyphosate and AMPA occur in precipitation, and facilitate contamination transport to 364 untreated areas.

365

366

Table2: Please insert here

Transport of AMPA and glyphosate is well documented (Vereecken, 2005), but more realistic experiments and modelling can be useful for a better understanding of exports by leaching and runoff. Scarce AMPA airborne data are available. The proportion of applied glyphosate transmitted to the atmosphere and its fate is unknown (Chang et al., 2011). Long-range transport of glyphosate and AMPA through the atmosphere has not been studied.

372

373 4. AMPA degradation in the watershed

374

AMPA degradation is not well documented because the fate of metabolites has only recently become a concern. Most sources of information are ambitious studies which were designed to gain full understanding of glyphosate fate, including that of AMPA in the total glyphosate in soil and water.

AMPA is considered persistent by the Pesticide Properties Database (PPDB, 2015), with a typical half-life (DT50) of 121 days. According to Bento et al. (2016), AMPA degradation is mainly a microbial process governed by temperature and soil moisture, in which AMPA degrades faster with heat and humidity. AMPA DT50 at 30°C ranges between 26 and 45 days in loess soil; DT90 (time after which 90% of the initial AMPA is degraded) ranges

between 88 and 148 days. Conversely, Bergström et al. (2011) found correlations between
AMPA degradation rates and amount of OM, observing a half-life of 35 days in clay topsoil
and 98 days in subsoil. Concerning loam top soils, Simonsen et al. (2008) calculated a DT50
of 32 days for AMPA, Zhang et al. (2015) observed AMPA DT50 ranging between 10 and 37
days, and lastly Mamy et al. (2005) observed 25-75 days.

AMPA degradation occurs in wetlands, as shown by Imfeld et al. (2013) and Degenhardt et al. (2012). These authors were not looking for a specific DT50, but did show that AMPA was more persistent than glyphosate in wetlands.

These findings confirm that the rate of AMPA degradation in soils is slower than that of glyphosate, except for high clay content soils (See Table 1 for Glyphosate degradation) (Bergström et al., 2011). Aparicio et al. (2013) explained this enhanced persistence by lower penetrability to cell membranes and stronger adsorption on particles.

Li et al. (2016) investigated AMPA degradation kinetics in laboratory experiments, at pH 7, 22 ℃, under artificial light with 5 g/L of mangane se oxide (birnessite) and 3 mmol/L of AMPA. Birnessite abiotically oxides 70% of initial AMPA in 7 days.

Ronco et al. (2016) evidenced that AMPA tends to accumulate in the bottom sediments of agricultural basin rivers, where it may be degraded by microbial fauna.

401 Presently AMPA is described as persistent and resilient to degradation under natural
402 conditions (Al-Rajab and Schiavon, 2010; Imfeld et al., 2013). AMPA degradation kinetics in
403 the environment should be investigated.

404

405 5. <u>AMPA public health concerns</u>

407 5.1. Toxicity

408

There is no epidemiological data on AMPA exposure from water. Limited data from in vitro studies are available, concerning its toxicity to human and animal cells (Cerdeira and Duke, 2010). Within the public health context, we will refer only to human-cell based studies.

The metabolite nature of AMPA may lead to questions as to its possible formation in the body. According to EFSA (2014), it is unlikely that glyphosate degradation to AMPA occurs in the human body (Niemann et al., 2015). Any AMPA contamination should not, therefore, be the result of glyphosate exposure. There is a lack of knowledge about AMPA formation from phosphonates in the body.

Kwiatkowska et al. (2014) showed that AMPA induces slight toxic effects on human 417 erythrocytes (in vitro). From 0.05 mM AMPA induces hemolysis, and from 0.25 mM AMPA is 418 able to create reactive oxygen species and increase the methemoglobin level in blood. 419 420 Regarding human lymphocytes, Mañas et al. (2009) found AMPA to have clastogenic effects 421 from 1.8 mM. They also noted that from 2.5 mM, Hep-2 DNA suffers significant damage. Finally, Benachour and Séralini (2009) proved that AMPA causes umbilical membrane cell 422 damages and occasionally embryonic or neonatal cells death. No occurrence study has been 423 424 conducted on blood outside of acute poisoning cases, in which AMPA can reach 2.6 µg/mL of blood (Han et al., 2016). 425

Surprisingly, Li et al. (2013) showed that AMPA and glyphosate inhibit growth of cancer cells,
but not of healthy cells. The authors propose the development of anticancer therapy based
on AMPA and glyphosate.

Mesnage et al. (2012) looked for the occurrence of AMPA in the excretions of a farmer and
his family members and were unable to find AMPA in their urine; the detection limit was 1
µg/L (approximatively 9.0 mM), low levels of AMPA in the urine may have been missed.
Hoppe (2013) provided a more accurate method, with a limit of quantification of 0.15 µg/L in

urine. 65 urine samples from 182 subjects were AMPA contaminated, with a mean of 0.18 433 µg/L (approximatively 1.6 mM). However, the author does not specify the type of population 434 435 studied. Exposure and assimilation routes may differ depending on occupation, residence area, and diet. To establish comparisons, future studies should pay attention to the possible 436 exposures of populations. Niemann et al. (2015) reviewed the occurrence of glyphosate in 437 the urine of Europeans and Americans, concluding that AMPA and glyphosate are poorly 438 439 correlated and suggesting that AMPA has other sources than glyphosate biodegradation. Hoppe (2013) analyzed 182 adults urine samples from 18 European countries. The ratios 440 AMPA/Glyphosate are very variable in the sampled urines. It suggests that European 441 populations, exposure to amino-polyphosphonates is added to glyphosate exposure 442

443

444 5.2. Policy framework: water

445

446 In Europe, anthropogenic compounds and potential other pollutants are now monitored in 447 water, in line with the 98/83/CE European Council directive (1998). Raw waters cannot be used to produce drinking water where any individual pesticide (including metabolites) 448 exceeds 2 µg/L, or if total pesticides (including metabolites) exceed 5 µg/L (JORF, 2007). 449 450 This policy also limits each compound to 0.1 µg/L in drinking water, and 0.5 µg/L for total pesticides (including metabolites). Designed to control pesticides, the directive has been 451 frequently updated and extended to cover monitoring of pertinent degradation products and 452 pharmaceuticals. 453

Elsewhere, to the best of our knowledge, regulatory policies concerning pesticides are less stringent in India, Australia, Canada, and USA. These countries have individually established pesticide thresholds, but do not regulate metabolites (Bhushan et al., 2013; NHMRC, 2016; Health Canada, 2014; US EPA, 2016). For example, the maximum admissible level of glyphosate in drinking water in the USA is 700 µg/L. It is 1000 µg/L in Australia, 280 µg/L in

Canada and absent from Indian policy (Bureau of Indian Standards, 2012). The World Health 459 Organization (WHO) in 2011, did not suggest formal guidelines for AMPA and glyphosate 460 (WHO, 2011), but the WHO reviewed its position in 2015 and declared glyphosate "probably 461 carcinogenic to humans" (Guyton et al., 2015). Moreover, Guyton et al. (2015) mentioned 462 that AMPA can induce oxidative stress which leads to chronic inflammation and from there 463 other unfavorable outcomes. Recently, the European Chemicals Agency gave an opposite 464 opinion and did not declared glyphosate as carcinogenic to humans, neither mentioned 465 466 AMPA (European Chemicals Agency, 2017).

Phosphonates are not mentioned by the WHO, probably because of the lack of studiesconcerning their behavior, toxicity and degradation.

469

470 5.3. Drinking water treatment

471

AMPA occurrence has frequently been reported in the various resources. AMPA is found in 472 groundwater at levels ranging from few thousandths µg/L to several tenths µg/L (Van 473 474 Stempvoort et al., 2014; Scribner et al., 2007; Van Stempvoort et al., 2016; (Battaglin et al., 2014). Van Stempvoort et al. (2016) found AMPA in shallow groundwater at depths within 2 475 m of the edge of the streams, with a detection frequency of 5% of analyzed samples. Earlier, 476 Scribner et al., (2007) reported results for two different groundwater surveillance programs. 477 AMPA detection frequency in the first program was 9.7% of 485 samples, with a maximum of 478 479 0.62 µg/L. In the second program concerning an agricultural watershed, AMPA detection frequency was 72.6% of 117 samples, with a maximum of 2.6 µg/L.". 480

In a French agricultural watershed, Piel et al. (2012) showed that AMPA occurs widely in streams all year round. Moreover, maximum AMPA concentrations appear to be higher in urban (5 μ g/L) than rural areas (2 μ g/L). In their broad study, Battaglin et al. (2014) showed that in USA streams, mean AMPA concentration is 0.2 μ g/L, and can reach 28 μ g/L with a

detection frequency of 71.6% of analyzed samples. In large rivers, mean AMPA concentration is 0.22 μ g/L and can reach 4.4 μ g/L, with a detection frequency of 89.3% of the analyzed samples. Many other worldwide studies on water confirm these trends (Poiger et al., 2016; Scribner et al., 2007).

Given that AMPA frequently occurs in water resources, the water treatment plants need to beable to remove AMPA.

AMPA removal is rarely documented; few authors report that AMPA removal or degradation can be achieved through common treatments. Sand filtration and bank filtration are moderately effective in AMPA elimination (25-95%), accommodating degrading microbiome and sorption sites for the pollutant (Jönsson et al., 2013). According to Hall and Camm (2007), coagulation/clarification of AMPA efficiency is uneven, ranging from 10% to 80%, depending on pH, coagulant type and dose.

As has been shown previously, AMPA is glyphosate's main biodegradation and photodegradation product in the environment, and this is also true for chemical oxidation (Brosillon et al., 2006; Mehrsheikh et al., 2006). Simple UV disinfection does not remove AMPA from drinking water (Brosillon et al. 2006; Assalin et al. 2010; Klinger et al. 1998). Chlorination using a HOCI/herbicide molar ratio greater than 2 provides full degradation of 10⁻⁴ M of glyphosate at ph. 7 for 24 hours in the dark (Brosillon et al., 2006). These conditions were used to represent the water residence time in the distribution network.

Batch laboratory tests showed that Ozonation of 42.28 mg/L glyphosate with 14 mg/L O₃ at pH 10 for 30 minutes removes almost 100% of the residues, including AMPA. Nevertheless, with pH adjustment at 6.5, degradation is incomplete and produces AMPA (Assalin et al., 2010). Advanced processes such as H_2O_2/UVC are effective in fully degrading 50 mg/L of glyphosate and then AMPA, providing basic pH (7-10), 80 W irradiation, and 2.2 to 5.9 mM of H_2O_2 for 5 hours (Manassero et al., 2010). AMPA is poorly eliminated by granular and powdered activated carbon – moreover, because activated carbon tends to remove residual

 O_3 and chlorinated oxidants, it can decrease AMPA oxidation (Besnault et al. 2015; Jönsson et al. 2013). Besnault et al. (2015) showed that a granular activated carbon (GAC) treatment process has good initial efficiency in terms of AMPA removal (>90%). However, from 2.5 m³ of treated water per kg of GAC, the pilot performance falls to 30-70% of elimination, and less than 30% beyond 16 m³/kg GAC.

These statements reveal how sensitive the processes are; they need to be optimized according to pollutant load and context. In many countries drinking water regulations do not include metabolites. Moreover the toxicological and epidemiological knowledges are insufficient or inappropriate to set a guideline (Benachour and Séralini, 2009).

520

521 Conclusions

522

523 Our review has shown that AMPA can have multiple sources; the main one seeming to be 524 glyphosate degradation in agricultural soils. This is explained by the short DT50 of 525 glyphosate and the higher resilience to degradation of amino-polyphosphonates. Moreover, 526 the vast diversity of uses of the herbicide leads to wide contamination, allowing AMPA 527 formation in many sections of the environment.

AMPA presents a low risk of leaching through soils. It occurs in groundwater having specific hydroclimatic and geomorphic conditions. Shallow groundwater is at greater risk of AMPA contamination, particularly where it is close to agricultural surfaces. Surface water bodies are vulnerable to contamination by AMPA. Rainfall intensity and the amount of time since application govern the export of AMPA towards surface waters. High adsorptive qualities induce AMPA-particle bonding, providing transport. Airborne AMPA is significant in agricultural areas.

AMPA is reported to be persistent; it tends to accumulate in soils if not leached. AMPA is substantially biodegraded only in OM-rich soils. In water, AMPA presents limited physical and chemical degradation, related to manganese oxides. It accumulates in sediments, where microbial degradation can occur.

539 In vitro studies have shown evidence of AMPA toxicity to human cells at low concentrations. A manifest lack of water-exposure epidemiological data prevents any conclusion being drawn 540 541 about AMPA safety in water. Current, tolerant water policies across the world demonstrate the uncertainties around AMPA. Its sources are poorly identified by responsible water 542 professionals, and seasonal related flows remain unknown. Fortunately, various water 543 treatment processes can be effective in removing AMPA through adequate adjustments and 544 temporal targeting. This does however raise concerns about rural catchments inherently 545 546 exposed to contamination, and often without water treatment.

547

548 Acknowledgment

549

550 This review has been performed with the support of ANRT (the French National Association

551 of Technical Research),

553 References

554

Al-Rajab, A.J., Hakami, O.M., 2014. Behavior of the non-selective herbicide glyphosate in agricultural soil. Am. J.

556 Environ. Sci. 10, 94–101. doi:10.3844/ajessp.2014.94.101

- 557 Al-Rajab, A.J., Schiavon, M., 2010. Degradation of 14C-glyphosate and aminomethylphosphonic acid (AMPA) in
- 558 three agricultural soils. J. Environ. Sci. 22, 1374–1380. doi:10.1016/S1001-0742(09)60264-3
- 559 Aparicio, V.C., De Gerónimo, E., Marino, D., Primost, J., Carriquiriborde, P., Costa, J.L., 2013. Environmental fate
- 560 of glyphosate and aminomethylphosphonic acid in surface waters and soil of agricultural basins.
- 561 Chemosphere 93, 1866–73. doi:10.1016/j.chemosphere.2013.06.041
- 562 Ascolani Yael, J., Fuhr, J.D., Bocan, G.A., Daza Millone, A., Tognalli, N., Dos Santos Afonso, M., Martiarena,
- 563 M.L., 2014. Abiotic degradation of glyphosate into aminomethylphosphonic acid in the presence of metals.
- 564 J. Agric. Food Chem. 62, 9651–9656. doi:10.1021/jf502979d
- Assalin, M.R., De Moraes, S.G., Queiroz, S.C.N., Ferracini, V.L., Duran, N., 2010. Studies on degradation of
 glyphosate by several oxidative chemical processes: ozonation, photolysis and heterogeneous

567 photocatalysis. J. Environ. Sci. Health. B. 45, 89–94. doi:10.1080/03601230903404598

- Barrett, K. a., McBride, M.B., 2005. Oxidative degradation of glyphosate and aminomethylphosphonate by
 manganese oxide. Environ. Sci. Technol. 39, 9223–9228. doi:10.1021/es051342d
- 570 Battaglin, W.A., Meyer, M.T., Kuivila, K.M., Dietze, J.E., 2014. Glyphosate and its degradation product AMPA
- 571 occur frequently and widely in U.S. soils, surface water, groundwater, and precipitation. J. Am. Water
- 572 Resour. Assoc. 50, 275–290. doi:10.1111/jawr.12159
- 573 Benachour, N., Séralini, G., 2009. Glyphosate Formulations Induce Apoptosis and Necrosis in Human Umbilical ,
 574 Embryonic , and Placental Cells. Chem. res. toxicol. 22, 97–105.
- 575 Benbrook, C.M., 2016. Trends in glyphosate herbicide use in the United States and globally. Environ. Sci. Eur. 28,
 576 3. doi:10.1186/s12302-016-0070-0
- 577 Bento, C.P.M.M., Yang, X., Gort, G., Xue, S., van Dam, R., Zomer, P., Mol, H.G.J.J., Ritsema, C.J., Geissen, V.,
- 578 2016. Persistence of glyphosate and aminomethylphosphonic acid in loess soil under different combinations
- 579 of temperature, soil moisture and light/darkness. Sci. Total Environ. 572, 301–311.
- 580 doi:10.1016/j.scitotenv.2016.07.215

- 581 Bergström, L., Börjesson, E., Stenström, J., 2011. Laboratory and lysimeter studies of glyphosate and
- 582 aminomethylphosphonic acid in a sand and a clay soil. J. Environ. Qual. 40, 98–108.
- 583 doi:10.2134/jeq2010.0179
- 584 Besnault, S., Martin-Ruel, S., Baig, S., Heiniger, B., Esperanza, M., Budzinski, H., Miège, C., Le Menach, K.,
- 585 Dherret, L., Roussel-Galle, A., Coquery, M., 2015. Technical, economic and environmental evaluation of
- 586 advanced tertiary treatments for micropollutants removal. IN FRENCH. Tech. Sci. Méthodes 67–83.
- 587 doi:10.1051/tsm/201503067
- Bhushan, C., Bhardwaj, A., Misra, S.S., 2013. State of Pesticide Regulations in India. Cent. Sci. Environ. New
 Delhi 1–72.
- 590 Blanchoud, H., Moreau-Guigon, E., Farrugia, F., Chevreuil, M., Mouchel, J.M., 2007. Contribution by urban and
- agricultural pesticide uses to water contamination at the scale of the Marne watershed. Sci. Total Environ.
- 592 375, 168–79. doi:10.1016/j.scitotenv.2006.12.009
- 593 Bøhn, T., Cuhra, M., Traavik, T., Sanden, M., Fagan, J., Primicerio, R., 2014. Compositional differences in
- soybeans on the market: Glyphosate accumulates in Roundup Ready GM soybeans. Food Chem. 153,
 207–215. doi:10.1016/j.foodchem.2013.12.054
- Borggaard, O.K., 2011. Does phosphate affect soil sorption and degradation of glyphosate? A review. Trends
 Soil Sci. Plant Nutr. 2, 16–27.
- Borggaard, O.K., Gimsing, A.L., 2008. Fate of glyphosate in soil and the possibility of leaching to ground and
 surface waters: a review. Pest Manag. Sci. 64, 441–456. doi:10.1002/ps.1512
- Botta, F., Lavison, G., Couturier, G., Alliot, F., Moreau-Guigon, E., Fauchon, N., Guery, B., Chevreuil, M.,
- 601 Blanchoud, H., 2009. Transfer of glyphosate and its degradate AMPA to surface waters through urban
- 602 sewerage systems. Chemosphere 77, 133–139. doi:10.1016/j.chemosphere.2009.05.008
- Brosillon, S., Wolbert, D., Lemasle, M., Roche, P., Mehrsheikh, A., 2006. Chlorination kinetics of glyphosate and
 its by-products: Modeling approach. Water Res. 40, 2113–2124. doi:10.1016/j.watres.2006.03.028
- 605 Bureau of indian Standards, 2012. Drinking water specification 1–3.
- 606 Candela, L., Caballero, J., Ronen, D., 2010. Glyphosate transport through weathered granite soils under irrigated
- and non-irrigated conditions Barcelona, Spain. Sci. Total Environ. 408, 2509–2516.
- 608 doi:10.1016/j.scitotenv.2010.03.006
- 609 Cerdeira, a L., Duke, S.O., 2010. Effects of glyphosate-resistant crop cultivation on soil and water quality. GM

- 610 Crops 1, 16–24. doi:10.4161/gmcr.1.1.9404
- 611 Chang, F. chih, Simcik, M.F., Capel, P.D., 2011. Occurrence and fate of the herbicide glyphosate and its
 612 degradate aminomethylphosphonic acid in the atmosphere. Environ. Toxicol. Chem. 30, 548–555.
 613 doi:10.1002/etc.431
- 614 Comber, S., Gardner, M., Georges, K., Blackwood, D., Gilmour, D., 2013. Domestic source of phosphorus to
- 615 sewage treatment works. Environ. Technol. 34, 1349–1358. doi:10.1080/09593330.2012.747003
- 616 Connor, M.S., Davis, J.A., Leatherbarrow, J., Greenfield, B.K., Gunther, A., Hardin, D., Mumley, T., Oram, J.J.,
- 617 Werme, C., 2007. The slow recovery of San Francisco Bay from the legacy of organochlorine pesticides.

618 Environ. Res. 105, 87–100. doi:10.1016/j.envres.2006.07.001

- 619 Coupe, R.H., Kalkhoff, S.J., Capel, P.D., Gregoire, C., 2012. Fate and transport of glyphosate and
- 620 aminomethylphosphonic acid in surface waters of agricultural basins. Pest Manag. Sci. 68, 16–30.
- 621 doi:10.1002/ps.2212
- Daouk, S., Grandjean, D., Chevre, N., De Alencastro, L.F., Pfeifer, H.-R., 2013. The herbicide glyphosate and its
 metabolite AMPA in the Lavaux vineyard area, Western Switzerland: proof of widespread export to surface
 waters. Part I: method validation in different water matrices. J. Environ. Sci. Health. B. 48, 717–724.

625 doi:10.1080/03601234.2013.780535

- Degenhardt, D., Cessna, A.J., Raina, R., Farenhorst, A., Pennock, D.J., 2012. Dissipation of six acid herbicides in
 water and sediment of two Canadian prairie wetlands. Environ. Toxicol. Chem. 30, 1982–1989.
- 628 doi:10.1002/etc.598
- Duke, S.O., 2011. Glyphosate degradation in Glyphosate-Resistant and-susceptible crops and weeds, Journal of
 Agricultural and Food Chemistry 59, 5835–5841.
- 631 EFSA, E.F.S.A., 2014. Renewal Assessment Report of 18 December 2013. Rapporteur Member State (RMS):
- 632 Germany, Co-RMS: Slovakia [WWW Document]. URL http://dar.efsa.europa.eu/dar-web/provision
- 633 European Chemicals Agency, 2017. Glyphosate not classified as a carcinogen by ECHA [WWW Document]. URL
- 634 https://echa.europa.eu/fr/-/glyphosate-not-classified-as-a-carcinogen-by-echa (accessed 3.20.17).
- 635 European Council, 1998. DIRECTIVE 98/83/CE [WWW Document]. URL http://eur-lex.europa.eu/legal-

636 content/FR/TXT/?uri=celex:31998L0083

- 637 European Parliament, 2012. REGULATION (EU) No 259/2012 OF THE EUROPEAN PARLIAMENT AND OF
- 638 THE COUNCIL of 14 March 2012 amending Regulation (EC) No 648/2004 as regards the use of

- 639 phosphates and other phosphorus compounds in consumer laundry detergents and consumer automatic640 dishwashe 16–21.
- Forlani, G., Prearo, V., Wieczorek, D., Kafarski, P., Lipok, J., 2011. Phosphonate degradation by Spirulina strains:
 Cyanobacterial biofilters for the removal of anticorrosive polyphosphonates from wastewater. Enzyme
- 643 Microb. Technol. 48, 299–305. doi:10.1016/j.enzmictec.2010.12.005
- 644 Gledhill, W.E., Feijtel, T.C., 1992. Environmental properties and safety assessment of organic phosphonates used
 645 for detergent and water treatment application.
- 646 Guyton, K.Z., Loomis, D., Grosse, Y., El Ghissassi, F., Benbrahim-Tallaa, L., Guha, N., Scoccianti, C., Mattock,
- H., Straif, K., Blair, A., Fritschi, L., McLaughlin, J., Sergi, C.M., Calaf, G.M., Le Curieux, F., Baldi, I.,
- 648 Forastiere, F., Kromhout, H., 't Mannetje, A., Rodriguez, T., Egeghy, P., Jahnke, G.D., Jameson, C.W.,
- 649 Martin, M.T., Ross, M.K., Rusyn, I., Zeise, L., 2015. Carcinogenicity of tetrachlorvinphos, parathion,
- 650 malathion, diazinon, and glyphosate. Lancet Oncol. 16, 490–491. doi:10.1016/S1470-2045(15)70134-8
- Hall, T., Camm, R., 2007. WRC report: Removal of glyphosate by water treatment.
- Han, J., Moon, H., Hong, Y., Yang, S., Jeong, W. joon, Lee, K.S., Chung, H., joon Jeong, W., Lee, K.S., Chung,
 H., 2016. Determination of glyphosate and its metabolite in emergency room in Korea. Forensic Sci. Int.
- 654 265, 41–46. doi:10.1016/j.forsciint.2015.12.049
- Hanke, I., Wittmer, I., Bischofberger, S., Stamm, C., Singer, H., 2010. Relevance of urban glyphosate use for
 surface water quality. Chemosphere 81, 422–429. doi:10.1016/j.chemosphere.2010.06.067
- 657 Health Canada, 2014. Guidelines for Canadian Drinking Water Quality Summary Table. Water and Air Quality
- Bureau, Healthy Enviroments and Consumer Safety Branch, Health Canada, Ottawa, Ontario. 1–25.
- Hoppe, H., 2013. Determination of Glyphosate residues in human urine samples from 18 European countries 22–
 23.
- Hsieh, Y.-J., Wanner, B.L., 2010. Global regulation by the seven-component Pi signaling system. Curr. Opin.
 Microbiol. 13, 198–203. doi:10.1016/j.mib.2010.01.014
- Imfeld, G., Lefrancq, M., Maillard, E., Payraudeau, S., 2013. Transport and attenuation of dissolved glyphosate
 and AMPA in a stormwater wetland. Chemosphere 90, 1333–9. doi:10.1016/j.chemosphere.2012.04.054
- Jaworska, J., Van Genderen-Takken, H., Hanstveit, A., Van de Plassche, E., Feijtel, T., 2002. Environmental risk
- assessment of phosphonates, used in domestic laundry and cleaning agents in the Netherlands.
- 667 Chemosphere 47, 655–665. doi:10.1016/S0045-6535(01)00328-9

- Jönsson, J., Camm, R., Hall, T., 2013. Removal and degradation of glyphosate in water treatment: a review. J.
- 669 Water Supply Res. Technol. 62, 395. doi:10.2166/aqua.2013.080
- 570 JORF, 2007. Ministerial decree of the 11/01/2007 concerning quality and references limits of raw waters, and
- drinking waters. IN FRENCH (Arrêté du 11 janvier 2007 relatif aux limites et références de qualité des eaux
- 672 brutes et des eaux destinées à la consommation hum [WWW Document]. URL
- 673 https://www.legifrance.gouv.fr/affichTexte.do?cidTexte=JORFTEXT000000465574
- 674 Kanissery, R., Welsh, A., Sims, G., 2015. Effect of Soil Aeration and Phosphate Addition on the Microbial
- 675 Bioavailability of Carbon-14-Glyphosate. J. Environ. {...} 44, 137–144. doi:10.2134/jeq2014.08.0331
- 676 Kjær, J., Ernstsen, V., Jacobsen, O.H., Hansen, N., de Jonge, L.W., Olsen, P., 2011. Transport modes and
- 677 pathways of the strongly sorbing pesticides glyphosate and pendimethalin through structured drained soils.

678 Chemosphere 84, 471–479. doi:10.1016/j.chemosphere.2011.03.029

- Klinger, J., Lang, M., Sacher, F., Brauch, H.-J., Maier, D., Worch, E., 1998. Formation of Glyphosate and AMPA
- 680 During Ozonation of Waters Containing Ethylenediaminetetra(methylenephosphonic acid). Ozone Sci. {&}
- 681 Eng. 20, 99–110. doi:10.1080/01919519808547279
- Kolpin, D.W., Thurman, E.M., Lee, E.A., Meyer, M.T., Furlong, E.T., Glassmeyer, S.T., 2006. Urban contributions
- of glyphosate and its degradate AMPA to streams in the United States. Sci. Total Environ. 354, 191–197.

684 doi:10.1016/j.scitotenv.2005.01.028

- 685 Kononova, S. V, Nesmeyanova, M. a, 2002. Phosphonates and their degradation by microorganisms.
- 686 Biochemistry. (Mosc). 67, 184–195. doi:10.1023/A:1014409929875
- Kools, S.A.E., Van Roovert, M., Van Gestel, C.A.M., Van Straalen, N.M., 2005. Glyphosate degradation as a soil
 health indicator for heavy metal polluted soils. Soil Biol. Biochem. 37, 1303–1307.
- 689 doi:10.1016/j.soilbio.2004.11.026
- Kwiatkowska, M., Huras, B., Bukowska, B., 2014. The effect of metabolites and impurities of glyphosate on
 human erythrocytes (in vitro). Pestic. Biochem. Physiol. 109, 34–43. doi:10.1016/j.pestbp.2014.01.003
- /
- La Jeunesse, I. La, Amiot, A., Landry, D., Jadas-hécart, A., Communal, P., Ballouche, A., Vitrai, B., 2015.
- 693 Transferts de pesticides dans un petit bassin versant viticole des coteaux du Layon : importance des pics
 694 lors du ruissellement. Press. Univ. Rennes 235.
- 695 Lesueur, C., Pfeffer, M., Fuerhacker, M., 2005. Photodegradation of phosphonates in water. Chemosphere 59,
- 696 685–91. doi:10.1016/j.chemosphere.2004.10.049

- 697 Li, H., Joshi, S.R., Jaisi, D.P., 2016. Degradation and Isotope Source Tracking of Glyphosate and
- 698 Aminomethylphosphonic Acid. J. Agric. Food Chem. 64, 529–538. doi:10.1021/acs.jafc.5b04838
- Li, Q., Lambrechts, M.J., Zhang, Q., Liu, S., Ge, D., Yin, R., Xi, M., You, Z., 2013. Glyphosate and AMPA inhibit
 cancer cell growth through inhibiting intracellular glycine synthesis. Drug Des. Devel. Ther. 7, 635–643.
- 701 doi:10.2147/DDDT.S49197
- T02 Lindhardt, B., Abildtrup, C., Vosgerau, H., Olsen, P., Torp, S., Iversen, B. V, Jørgensen, J.O., Plauborg, F.,
- 703 Rasmussen, P., Gravesen, P., 2001. The danish pesticide leaching assessment programme, Site
- 704 Characterization and monitoring design, Geological survey of Denmark and Greeland.
- 705 Lupi, L., Miglioranza, K.S.B., Aparicio, V.C., Marino, D., Bedmar, F., Wunderlin, D.A., 2015. Occurrence of
- 706 glyphosate and AMPA in an agricultural watershed from the southeastern region of Argentina. Sci. Total
 707 Environ. 536, 687–94. doi:10.1016/j.scitotenv.2015.07.090
- 708 Mallat, E., Barceló, D., 1998. Analysis and degradation study of glyphosate and of aminomethylphosphonic acid
- in natural waters by means of polymeric and ion-exchange solid-phase extraction columns followed by ion
- 710 chromatography–post-column derivatization with fluorescence detection. J. Chromatogr. A 823, 129–136.
- 711 doi:10.1016/S0021-9673(98)00362-8
- Mamy, L., Barriuso, E., Gabrielle, B., 2016. Glyphosate fate in soils when arriving in plant residues. Chemosphere
 154, 425–433. doi:10.1016/j.chemosphere.2016.03.104
- 714 Mamy, L., Barriuso, E., Gabrielle, B., 2005. Environmental fate of herbicides trifluralin, metazachlor, metamitron
- and sulcotrione compared with that of glyphosate, a substitute broad spectrum herbicide for different
- 716 glyphosate-resistant crops. Pest Manag. Sci. 61, 905–916. doi:10.1002/ps.1108
- 717 Mañas, F., Peralta, L., Raviolo, J., Ovando, H.G., Weyers, A., Ugnia, L., Cid, M.G., Larripa, I., Gorla, N., 2009.
- 718 Genotoxicity of glyphosate assessed by the comet assay and cytogenetic tests. Environ. Toxicol.
- 719 Pharmacol. 28, 37–41. doi:10.1016/j.etap.2009.02.001
- Manassero, A., Passalia, C., Negro, A.C.C., Cassano, A.E.E., Zalazar, C.S.S., 2010. Glyphosate degradation in
 water employing the H2O2/UVC process. Water Res. 44, 3875–3882. doi:10.1016/j.watres.2010.05.004
- 722 Matys, S., V, Kuzmina, N., M., Laurinavichius, K., S., Nesmeyanova, M., A., 2004. Effect of environmental factors
- on degradation of the C P bond of methylphosphonate by Escherichia coli cells. Process Biochem. 39,
- 724 1063–1071. doi:10.1016/S0032-9592(03)00231-0
- 725 Mehrsheikh, A., Bleeke, M., Brosillon, S., Laplanche, A., Roche, P., 2006. Investigation of the mechanism of

- 726 chlorination of glyphosate and glycine in water. Water Res. 40, 3003–3014.
- 727 doi:10.1016/j.watres.2006.06.027
- Mesnage, R., Moesch, C., Le grand, R., Lauthier, G., Spiroux de vendomois, J., Gress, S., Seralini, G., 2012.
 Glyphosate Exposure in a Farmer's Family. Am. J. Hum. Genet. 70, 1077–1088. doi:10.1086/340363
- 730 Napoli, M., Cecchi, S., Zanchi, C. a., Orlandini, S., 2015. Leaching of Glyphosate and Aminomethylphosphonic
- 731 Acid through Silty Clay Soil Columns under Outdoor Conditions. J. Environ. Qual. 0, 0.
- 732 doi:10.2134/jeq2015.02.0104
- 733 NHMRC, 2016. Australian Drinking Water Guidelines 6: Volume 1. doi:1864965118
- 734 Niemann, L., Sieke, C., Pfeil, R., Solecki, R., 2015. A critical review of glyphosate findings in human urine
- samples and comparison with the exposure of operators and consumers. J. fur Verbraucherschutz und Leb.
- 736 10, 3–12. doi:10.1007/s00003-014-0927-3
- Norgaard, T., Moldrup, P., Ferré, T.P. a., Olsen, P., a.E. Rosenbom, de Jonge, L.W.W., Rosenbom, a. E., de
- Jonge, L.W.W., 2014. Leaching of Glyphosate and Aminomethylphosphonic Acid from an Agricultural Field
 over a Twelve-Year Period. Vadose Zo. J. 13. doi:10.2136/vzj2014.05.0054
- 740 Nowack, B., 2004. Environmental chemistry of phosphonic acids. Integr. Environ. Technol. Ser. 147–173.
- 741 Nowack, B., 2003. Environmental chemistry of phosphonates. Water Res. 37, 2533–2546. doi:10.1016/S0043742 1354(03)00079-4
- Nowack, B., 1997. The behavior of phosphonates in wastewater treatment plants of Switzerland. Water Res. 32,
 1271–1279. doi:10.1016/S0043-1354(97)00338-2
- 745 Nowack, B., Stone, A.T., 2002. Homogeneous and heterogeneous oxidation of nitrilotrismethylenephosphonic
- acid (NTMP) in the presence of manganese(II, III) and molecular oxygen. J. Phys. Chem. B 106, 6227–
 6233. doi:10.1021/jp014293+
- Obojska, A., Lejczak, B., 2003. Utilisation of structurally diverse organophosphonates by Streptomycetes. Appl.
 Microbiol. Biotechnol. 62, 557–563. doi:10.1007/s00253-003-1281-z
- Okada, E., Costa, J.L., Bedmar, F., 2016. Adsorption and mobility of glyphosate in different soils under no-till and
 conventional tillage. Geoderma 263, 78–85. doi:10.1016/j.geoderma.2015.09.009
- 752 Ololade, I.A., Oladoja, N.A., Oloye, F.F., Alomaja, F., Akerele, D.D., Iwaye, J., Aikpokpodion, P., 2014. Sorption of
- 753 Glyphosate on Soil Components: The Roles of Metal Oxides and Organic Materials. Soil {&} Sediment

	ACCEPTED MANUSCRIPT										
754	Contam. 23, 571–585. doi:Doi 10.1080/15320383.2014.846900										
755	Piel, S., Baurès, E., Thomas, O., 2012. Contribution to surface water contamination understanding by pesticides										
756	and pharmaceuticals, at a watershed scale. Int. J. Environ. Res. Public Health 9, 4433–4451.										
757	doi:10.3390/ijerph9124433										
758	Poiger, T., Buerge, I.J., Bächli, A., Müller, M.D., Balmer, M.E., 2016. Occurrence of the herbicide glyphosate and										
759	its metabolite AMPA in surface waters in Switzerland determined with on-line solid phase extraction LC-										
760	MS/MS. Environ. Sci. Pollut. Res. 1–9. doi:10.1007/s11356-016-7835-2										
761	PPDB, 2015. Aminomethylphosphonic acid; Verified in September 2016 [WWW Document]. URL										
762	http://sitem.herts.ac.uk/aeru/ppdb/en/Reports/842.htm										
763	Rampazzo, N., Rampazzo Todorovic, G., Mentler, a., Blum, W.E.H.E.H., 2013. Adsorption of glyphosate and										
764	aminomethylphosphonic acid in soils. Int. Agrophysics 27, 203–209. doi:10.2478/v10247-012-0086-7										
765	Ramwell, C.T., Kah, M., Johnson, P.D., 2014. Contribution of household herbicide usage to glyphosate and its										
766	degradate aminomethylphosphonic acid in surface water drains. Pest Manag. Sci. 70, 1823–1830.										
767	doi:10.1002/ps.3724										
768	Ronco, A.E., Marino, D.J.G., Abelando, M., Almada, P., Apartin, C.D., 2016. Water quality of the main tributaries										
769	of the Paran?? Basin: glyphosate and AMPA in surface water and bottom sediments. Environ. Monit.										
770	Assess. 188. doi:10.1007/s10661-016-5467-0										
771	Schmidt, C.K., Raue, B., Brauch, HJ., Sacher, F., 2013. Trace-level analysis of phosphonates in environmental										
772	waters by ion chromatography and inductively coupled plasma mass spectrometry. Int. J. Environ. Anal.										
773	Chem. 94, 385–398. doi:10.1080/03067319.2013.831410										
774	Scribner, E. a., Battaglin, W. a., Gilliom, R.J.J., Meyer, M.T.T., 2007. Concentrations of Glyphosate, Its										
775	Degradation Product, Aminomethylphosphonic Acid, and Glufosinate in Ground- and Surface-Water,										
776	Rainfall, and Soil Samples Collected in the United States, 2001–06. U.S. Geol. Surv. Investig. Rep. 2007-										
777	5122 111.										
778	Sidoli, P., Baran, N., Angulo-Jaramillo, R., 2015. Glyphosate and AMPA adsorption in soils: laboratory										
779	experiments and pedotransfer rules. Environ. Sci. Pollut. Res. Int. doi:10.1007/s11356-015-5796-5										

- Simonsen, L., Fomsgaard, I.S., Svensmark, B., Spliid, N.H., 2008. Fate and availability of glyphosate and AMPA
 in agricultural soil. J. Environ. Sci. Health. B. 43, 365–375. doi:10.1080/10934520701795517
- 782 Struger, J., Van Stempvoort, D.R.R., Brown, S.J.J., 2015. Sources of aminomethylphosphonic acid (AMPA) in

ACCEPTED MANUSCRIPT 783 urban and rural catchments in Ontario, Canada: Glyphosate or phosphonates in wastewater? Environ. 784 Pollut. 204, 289-297. doi:10.1016/j.envpol.2015.03.038 785 Studnik, H., Liebsch, S., Forlani, G., Wieczorek, D., Kafarski, P., Lipok, J., 2015. Aminopolyphosphonates -786 chemical features and practical uses, environmental durability and biodegradation. N. Biotechnol. 32, 1-6. 787 doi:10.1016/j.nbt.2014.06.007 788 Sviridov, A. V, Shushkova, T. V, Ermakova, I.T., Ivanova, E. V, Epiktetov, D.O., Leontievsky, A.A., 2015. Microbial 789 Degradation of Glyphosate Herbicides (Review). Appl. Biochem. Microbiol. 51, 188–195. 790 doi:10.1134/S0003683815020209 791 Tang, T., Boenne, W., Desmet, N., Seuntjens, P., Bronders, J., van Griensven, A., 2015. Quantification and 792 characterization of glyphosate use and loss in a residential area. Sci. Total Environ. 517, 207-214. 793 doi:10.1016/j.scitotenv.2015.02.040 794 Todorovic-Rampazzo, G., Rampazzo, N., Mentler, A., Blum, W.E.H., Eder, A., Strauss, P., 2014. Influence of soil 795 tillage and erosion on the dispersion of glyphosate and aminomethylphosphonic acid in agricultural soils. 796 Int. Agrophysics 28, 93–100. doi:10.2478/intag-2013-0031 797 Torstensson, L., Börjesson, E., Stenström, J., 2005. Efficacy and fate of glyphosate on Swedish railway 798 embankments. Pest Manag. Sci. 61, 881-886. doi:10.1002/ps.1106 799 US EPA, O., 2016. Table of Regulated Drinking Water Contaminants. 800 Van Stempvoort, D.R., Roy, J.W., Brown, S.J., Bickerton, G., 2014. Residues of the herbicide glyphosate in 801 riparian groundwater in urban catchments. Chemosphere 95, 455–463. 802 doi:10.1016/j.chemosphere.2013.09.095 803 Van Stempvoort, D.R., Spoelstra, J., Senger, N.D., Brown, S.J., Post, R., Struger, J., 2016. Glyphosate residues

- in rural groundwater, Nottawasaga River Watershed, Ontario, Canada. Pest Manag. Sci.
- 805 doi:10.1002/ps.4218
- Vereecken, H., 2005. Mobility and leaching of glyphosate: A review. Pest Manag. Sci. 61, 1139–1151.
 doi:10.1002/ps.1122
- 808 Wang, S., Seiwert, B., Kästner, M., Miltner, A., Schäffer, A., Reemtsma, T., Yang, Q., Nowak, K.M., 2016.
- 809 (Bio)degradation of glyphosate in water-sediment microcosms A stable isotope co-labeling approach.
- 810 Water Res. 99, 91–100. doi:10.1016/j.watres.2016.04.041
- 811 WHO, 2011. Guidelines for Drinking-water Quality. Fourth Ed. 564. doi:10.1016/S1462-0758(00)00006-6

812 World Health Organisation, 2004. Copper in Drinking-water Background document for development of WHO

813 Guidelines for Drinking-water Quality. doi:10.1016/j.kjms.2011.05.002

- Yang, X., Wang, F., Bento, C.P.M., Meng, L., van Dam, R., Mol, H., Liu, G., Ritsema, C.J., Geissen, V., 2015a.
- 815 Decay characteristics and erosion-related transport of glyphosate in Chinese loess soil under field
- 816 conditions. Sci. Total Environ. 530–531, 87–95. doi:10.1016/j.scitotenv.2015.05.082
- Yang, X., Wang, F., Bento, C.P.M., Xue, S., Gai, L., van Dam, R., Mol, H., Ritsema, C.J., Geissen, V., 2015b.
- 818 Short-term transport of glyphosate with erosion in Chinese loess soil--a flume experiment. Sci. Total
- 819 Environ. 512–513, 406–414. doi:10.1016/j.scitotenv.2015.01.071
- Zgheib, S., Moilleron, R., Chebbo, G., 2012. Priority pollutants in urban stormwater: Part 1 Case of separate
- 821 storm sewers. Water Res. 46, 6683–6692. doi:10.1016/j.watres.2011.12.012
- 822 Zhang, C., Hu, X., Luo, J., Wu, Z., Wang, L., Li, B., Wang, Y., Sun, G., 2015. Degradation Dynamics of
- 823 Glyphosate in Different Types of Citrus Orchard Soils in China. Molecules 20, 1161–1175.
- 824 doi:10.3390/molecules20011161

Table 1: Biological degradation of glyphosate in soils, leading to AMPA (Batch laboratory)

	Glyphosate half-life (DT50) (days)			
	Topsoil ((soil (0-30 cm) Subsoil (30-80 cm)		
Soil	Oxic	Anoxic	Oxic	
	18	45		Kanissery et al. (2015)
Silty clay loam	14.5			Al-Rajab et Hakami (2014)
	19			Al-Rajab et Schiavon (2010)
Silt loom	15	51		Kapissony et al. (2015)
Siit ioani	18	42		Kanissery et al. (2015)
Sand	16.9		36.5	Borgström at al. (2011)
Clay	110		151	Bergstrom et al. (2011)
Loam	9			Simonsen et al. (2008)
	3.5			Yang et al. (2015b)
LUESS	1.5 - 53.5			Bento et al. (2016)
	7.1		Y	Mamy et al. (2016)
Clay Loam	10.6		\sim	Druart et al. (2011)
	4			Al-Rajab et Schiavon (2010)
Sandy loam	14.5			Al-Rajab et Schiavon (2010)

		-							
	Location		%	[Glyphosate] (µg/L)		%	[AMPA] (µg/L)		
			detection	max	median	detection	max	median	
Scribner et al. (2007)		14	86.0	1.1	0.274	86.0	0.47	0.113	
Battaglin et al. (2014)	USA	85	70.6	2.5	0.11	71.8	0.48	0.04	
	Mississippi 2007	11	73.0	1.9	0.2	73.0	0.3	0.1	
	2008	19	68.0	1.6	0.15	74.0	0.48	< 0.1	
Chang et al. (2011)	lowa 2007	14	71.0	2.5	0.2	36.0	0.2	< 0.1	
	2008	24	63.0	1.8	0.1	50.0	0.24	< 0.1	
	Indiana 2004	12	92.0	1.1	0.14	92.0	0.47	< 0.1	
Van Stempvoort et al. (2016)	Canada	15	86.7	0.135	-	26.7	0.019	-	
		A)'						
		Y							

Table 2: Glyphosate and AMPA concentrations in rain



Figure 1: AMPA in natural waters: its sources (glyphosate and phosphonates), behavior and environmental fate



Figure 2: Phosphonate degradation pathways to AMPA in the aquatic environment





- > AMPA is ubiquitous in all environmental compartments, particularly water.
- > AMPA is derived from urban phosphonates, and glyphosate.
- > Off-site movement of AMPA is mainly due to rainfall, towards surface waters.
- > AMPA is persistent but can be biologically degraded in soils and sediments.
- > Harmful effects of AMPA are currently unknown due to a lack of studies.