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Design and Synthesis of a 4-Nitrobromobenzene Derivative Bearing an Ethylene Glycol Tetraacetic Acid Unit for a New Generation of Caged Calcium Compounds with Two-Photon Absorption Properties in the Near-IR Region and Their Application in Vivo

Satish Jakkampudi,^{†,‡} Manabu Abe,^{*,†,‡} Naomitsu Komori,[†] Ryukichi Takagi,[†] Ko Furukawa,[§] Claudine Katan,^{||} Wakako Sawada,[⊥] Noriko Takahashi,[⊥] and Haruo Kasai^{*,†,‡,⊥}

[†]Department of Chemistry & Research Center for Future Science, Graduate School of Science, Hiroshima University, 1-3-1 Kagamiyama, Higashi-Hiroshima, Hiroshima 739-8526, Japan

[‡]JST-CREST, K's Gobancho 7, Gobancho, Chiyodaku, Tokyo 102-0075, Japan

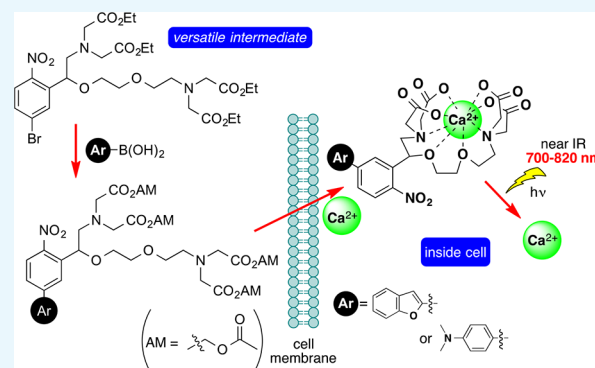
[§]Center for Instrumental Analysis, Institute for Research Promotion, Niigata University, 8050 Ikarashi 2-no-cho, Nishi-ku, Niigata 950-2181, Japan

^{||}Institut des Sciences Chimiques de Rennes, UMR 6226 CNRS-Université Rennes 1, 35042 Rennes, France

[⊥]Laboratory of Structural Physiology, CDBIM, Graduate School of Medicine, The University of Tokyo, Bunkyo-ku, Tokyo 113-0033, Japan

S Supporting Information

ABSTRACT: Among biologically active compounds, calcium ions (Ca^{2+}) are one of the most important species in cell physiological functions. Development of new calcium chelators with two-photon absorption (TPA) properties is a state-of-the-art challenge for chemists. In this study, we report the first and efficient synthesis of 5-bromo-2-nitrobenzyl-substituted ethylene glycol tetraacetic acid (EGTA) as a platform for a new generation of calcium chelators with TPA properties in the near-infrared region. New calcium chelators with high TPA properties, that is, a two-photon (TP) fragmentation efficiency of $\delta_u = 20.7$ GM at 740 nm for 2-(4-nitrophenyl)benzofuran (NPBF)-substituted EGTA (NPBF-EGTA, $K_d = 272$ nM) and $\delta_u = 7.8$ GM at 800 nm for 4-amino-4'-nitro-1,1'-biphenyl (BP)-substituted EGTA (BP-EGTA, $K_d = 440$ nM) derivatives, were synthesized using Suzuki–Miyaura coupling reactions of the bromide with benzofuran-2-boronic acid and 4-(dimethylamino)phenyl boronic acid, respectively. The corresponding acetoxymethyl (AM) esters were prepared and successfully applied to the Ca^{2+} -uncaging reaction triggered by TP photolysis in vivo.



INTRODUCTION

Calcium ions (Ca^{2+}) play pivotal roles in various biological phenomena,^{1–3} including sending and receiving nerve motions, muscle contractions, delivery of hormones and other chemicals such as neurotransmitters, maintaining a normal heartbeat, and blood clotting. Artificial control of Ca^{2+} levels, $[\text{Ca}^{2+}]$, in cells would be key to elucidating the role of Ca^{2+} in biological phenomena. Ca^{2+} levels may be controlled by the “encapsulation (=caging) and release (=uncaging)” of Ca^{2+} . Biologically active cations (Ca^{2+}) can be caged by organic chelators^{4–9} and uncaged by photo-triggered structural changes in the chelator.

Since 1980, the structures of ethylene glycol tetraacetic acid (EGTA)⁵ and 1,2-bis(*o*-aminophenoxy)ethane-*N,N,N',N'*-tetraacetic acid (BAPTA)^{10,11} have been known as excellent chelators that bind selectively to Ca^{2+} (Figure 1). Indeed, Ellis-Davies and co-workers used an EGTA chelator for the

photochemical uncaging of biologically active Ca^{2+} .^{4–9} In 2012, BAPTA was used by Campo and co-workers for pH-independent caging of Ca^{2+} as well as its uncaging by photolysis.¹¹

For state-of-the-art physiological experiments in living cells and organisms, two-photon (TP) uncaging process using near-infrared (IR) light is becoming essential.^{4–9,12–25} Alternatively, lanthanide-doped upconversion nanoparticles have also proved to be efficient in recent years.^{26–32} Meanwhile, the three-dimensional (3D) resolution achieved using TP excitation is higher than that of one-photon (OP) excitation reactions. Furthermore, the near-IR TP excitation process allows deeper

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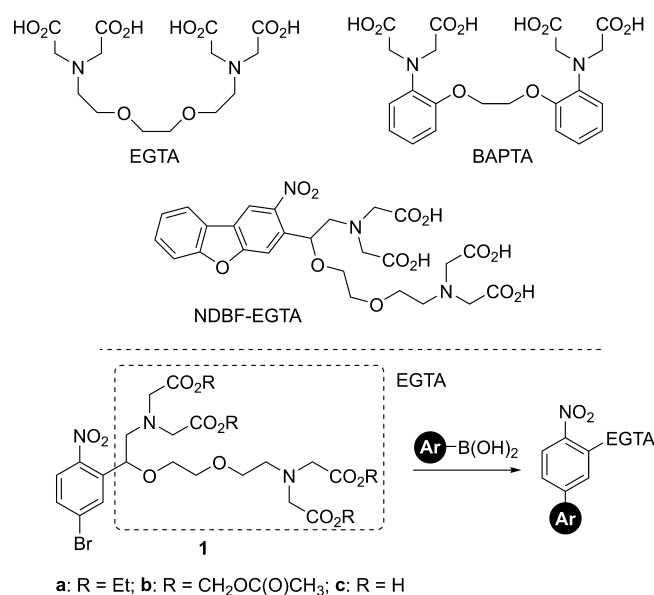


Figure 1. Design and synthesis of new TPA chromophores bearing EGTA unit from platform 1a.

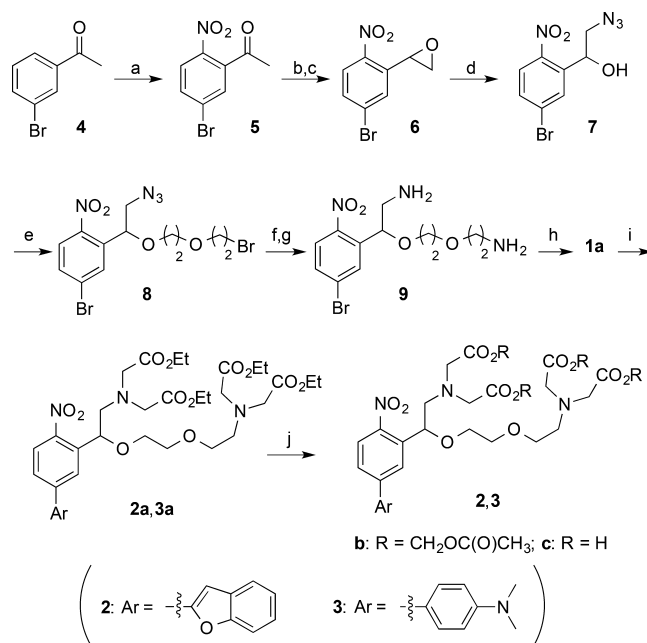
penetration of the light beam in living tissue and reduced light scattering.^{4–9,12–25} The optical absorption by endogenous chromophores is also reduced, so less photo-damage to cells is expected. To date, nitrodibenzofuran (NDBF)-substituted EGTA (NDBF-EGTA)⁵ is the only Ca²⁺ chelator reported to have TP uncaging reactivity when excited at 720 nm (Figure 1). The TP uncaging cross section (δ_u), $\delta_u = \sigma_2 \times \varphi$, was reported to be ~ 0.6 Göppert-Mayer (GM)³³ (1 GM = 10^{-50} cm⁴ s/photon/molecule). The value of σ_2 is the two-photon absorption (TPA) cross section in GM, and φ is the quantum yield of the uncaging reaction. High cross-section GM values in the TP uncaging reaction, for example, 3 GM,³⁴ have been suggested as a requirement for physiological studies.

In the present study, we report the first and efficient synthesis of 5-bromo-2-nitrobenzyl-substituted EGTA derivative 1a (R = Et), which is a versatile platform for a new generation of caged calcium compounds (Figure 1). Bromide 1a can be utilized for the synthesis of a wide variety of TP-responsive Ca²⁺ chelators, using Suzuki–Miyaura coupling reactions.³⁵ Thus, the TP absorption maximum and TP reactivity can be tuned by selecting a different aryl group (Ar) in boronic acids ArB(OH)₂. Compounds 2 and 3 (Scheme 1) were prepared as examples in this study. Efficiencies of TP-induced bond cleavage reactions were found to be as high as $\delta_u = 20.7$ GM at 740 nm for 2a and $\delta_u = 7.8$ GM at 800 nm for 3a. The corresponding acetoxy methyl (AM) esters 2b and 3b were synthesized and were shown to be effective for in vivo Ca²⁺ uncaging reaction.

RESULTS AND DISCUSSION

Synthesis of Compounds 1a, 2, and 3. The synthesis of 1a is summarized in Scheme 1. Commercially available starting material 3-bromo-acetophenone (4) was nitrated³⁶ with nitric acid and sulfuric acid to afford nitro-substituted compound 5 in 64% yield. Bromination of 5 with Br₂ in the presence of acetic acid in dichloromethane at room temperature (RT) provided the brominated compound in 99% yield. Without further purification, the keto function of the brominated compound obtained was reduced using sodium borohydride, followed by

Scheme 1. Synthesis of 1a, 2, and 3^a



^aReagents and conditions: (a) KNO₃, H₂SO₄, CH₂Cl₂, 0 °C–RT, 4 h, 64%; (b) Br₂, AcOH/CH₂Cl₂, RT, 55 h, 99%; (c) NaBH₄, aq NaOH (2.5 M), 1,4-dioxane/MeOH, 0 °C, 1 h, 71%; (d) NaN₃, DMF, RT–40 °C, 4 h, 40%; (e) bis(2-bromoethyl) ether, Cs₂CO₃, ACN, RT–50 °C, 5 h, 53%; (f) NaN₃, DMF, RT, 16 h, 80%; (g) PPh₃, RT, 24 h, 80%; (h) NaI, DIEPA, ethyl bromoacetate, ACN, 0–80 °C, 3 h, 70%; (i) ArB(OH)₂, Pd(PPh₃)₄, K₂CO₃, THF, H₂O, RT–60 °C, 3 h, 95% for 2a and 90% for 3a; (j) (I) KOH, MeOH, 0 °C–RT, 16 h; (II) bromomethyl acetate, DIPEA, DMF, 0 °C–RT, 16 h, 60%. DIPEA = *N,N*-diisopropylethylamine; THF = tetrahydrofuran; ACN = acetonitrile.

in situ cyclization under alkaline treatment for 30 min at 0 °C, giving oxirane 6 in 71% yield. Epoxide 6 was subsequently treated with sodium azide³⁷ in dimethylformamide (DMF) at 40 °C to afford hydroxyl azide 7 in 40% yield. This step was the key in our construction of the EGTA unit. The reaction yield was largely dependent on the equivalents of sodium azide and the reaction temperature. After several attempts, 1 equiv NaN₃ and ~ 40 °C were found to be the best reaction conditions. Hydroxyl azide was alkylated³⁸ with bis(2-bromomethyl)ether in the presence of cesium carbonate in DMF at 50 °C to give compound 8 in 53% yield. The isolated alkylated compound 8 was treated with sodium azide at RT to give the diazide³⁹ in 80% yield. The diazide was then converted to diamine 9 in 80% yield by treatment with PPh₃,⁴⁰ followed by hydrolysis. Diamine 9 was alkylated⁴¹ with ethyl bromoacetate in the presence of sodium iodide in CH₃CN to give tetraacetic acid derivative 1a in 70% yield.

Bromide 1a is a versatile intermediate in Suzuki–Miyaura coupling reactions for producing a new generation of Ca²⁺ chelators with TPA properties. Numerous TPA chromophores can be prepared from the 1a platform. For example, in this study, bromide 1a was treated with benzofuran-2-boronic acid or 4-(dimethylamino)phenylboronic acid in the presence of Pd(PPh₃)₄ and K₂CO₃ to afford EGTA derivatives 2a or 3a with benzofuran or biphenyl units as TP chromophores in 95% and 90% yield, respectively. Ethyl esters 2a and 3a were hydrolyzed with potassium hydroxide (KOH), followed by esterification with acetoxy methyl (AM) ester to give 2b and 3b

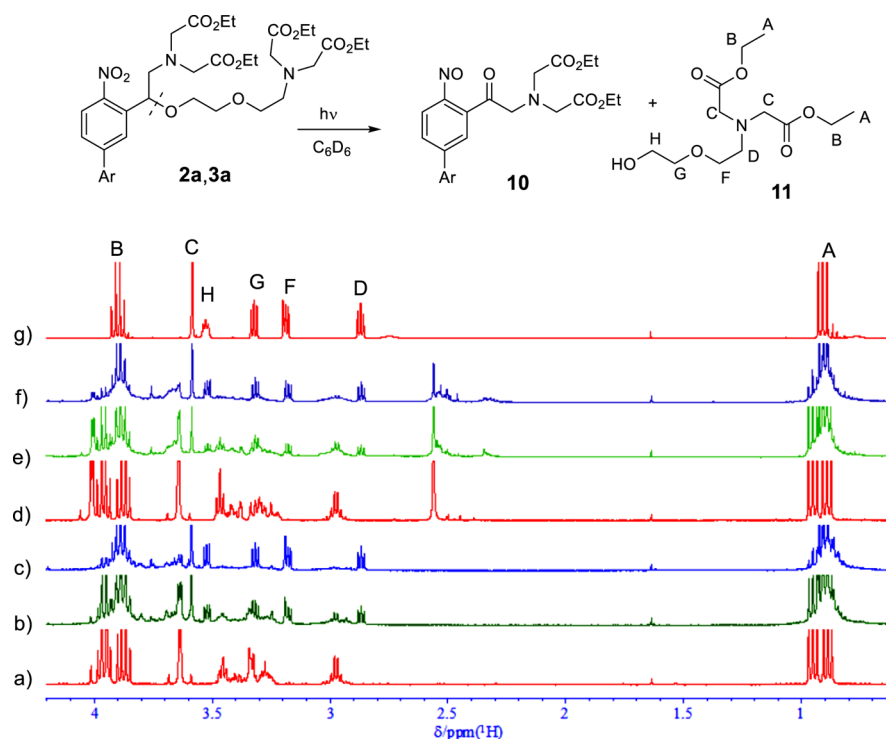


Figure 2. ^1H NMR (400 MHz), (a) δ 0.5–4.2 ppm of **2a** in C_6D_6 , (b) after 4 h, and (c) 8 h irradiation at 360 nm; (d) ^1H NMR (400 MHz), δ 0.5–4.2 ppm of **3a** in C_6D_6 , (e) after 5 h, and (f) 10 h irradiation at 400 nm; and (g) ^1H NMR (400 MHz), δ 0.5–4.2 ppm of **11** in C_6D_6 .

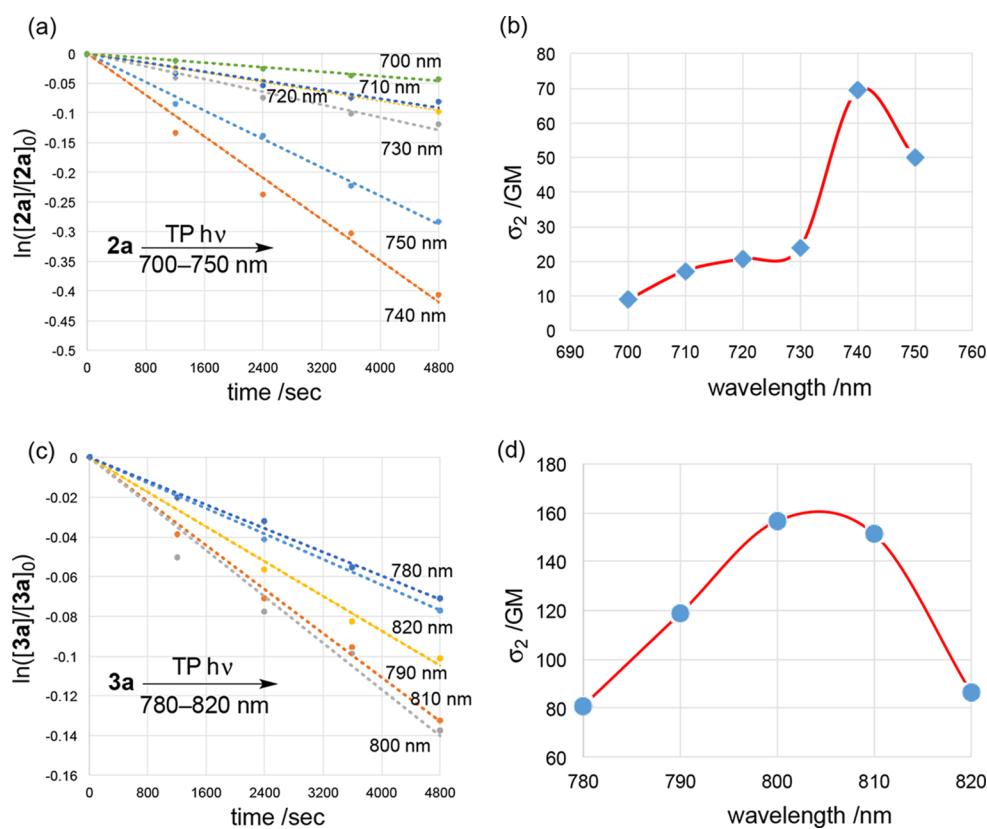


Figure 3. (a) Time profile of TP uncaging of **2a** $\ln([2a]/[2a]_0)$ vs irradiation time at 700–750 nm, (b) an action TP absorption spectrum of **2a**, (c) time profile of TP uncaging of **3a** $\ln([3a]/[3a]_0)$ vs irradiation time at 780–820 nm, and (d) an action TP absorption spectrum of **3a**.

in 60% yield. AM ester **2b** was loaded into cells to generate intracellular EGTA calcium chelators, *vide infra*.

OP Photolysis of **2a and **3a**.** The photoreactions of compounds **2a**⁴² and **3a** were investigated first using an OP excitation process (Figure 2). The OP photochemical reaction

of **2a** (10 mM in C_6D_6 , $\lambda_{max} = 362$ nm with ϵ 18 771 $M^{-1} cm^{-1}$) was conducted at 360 ± 10 nm using a monochromator with a xenon lamp (500 W). The photo-induced bond cleavage reaction was monitored using 1H NMR (400 MHz) spectroscopic analysis (Figure 2a–c). As shown in our preliminary results on the photolysis of **2a**,⁴² clear decomposition of **2a** was confirmed after irradiation for 8 h. Using 1H NMR analysis, fragmentation product **11** was observed in the photolysate. Compound **11** was prepared in this study to confirm its formation during the photolysis. The quantum yield (ϕ) of the photochemical decomposition of EGTA ethyl ester **2a** was determined to be 0.3 ± 0.05 at 360 nm irradiation using a xenon lamp. Ferric oxalate was used as a chemical actinometer to determine the quantum yield.⁴³

The OP bond cleavage reaction of **3a** (10 mM in C_6D_6 , $\lambda_{max} = 403$ nm with ϵ 8790 $M^{-1} cm^{-1}$) was also conducted at 400 ± 10 nm using a xenon lamp (Figure 2d–f). The photo-induced bond cleavage was monitored using 1H NMR (400 MHz) analysis. The complete structural decomposition of compound **3a** was observed after irradiation for 10 h. The quantum yield of the decomposition of EGTA ethyl ester **3a** was determined to be 0.05 at 400 nm irradiation.

TP Photolysis of 2a and 3a. Photochemical reactions using TP excitation by near-IR light were conducted for **2a** and **3a** in dimethyl sulfoxide (DMSO). The TP photolysis of **2a** was performed at 700, 710, 720, 730, 740, and 750 nm with a Ti:sapphire laser using ~ 700 mW at ~ 25 °C (Figure 3a,b). The TP photolysis of **3a** was conducted at 780, 790, 800, 810, and 820 nm at ~ 25 °C (Figure 3c,d). Using high-performance liquid chromatography (HPLC) analysis, TP-induced bond cleavage reactions were monitored by the consumption of **2a** or **3a**. Wavelength-dependent changes in the rate constants were determined by the TP decomposition of **2a** or **3a**. As reported previously, the absolute value of the TPA cross section for 2-(4-nitrophenyl) benzofuran was 18 GM at 720 nm.⁴² The TP action spectra for **2a** and **3a** were extrapolated in comparison with the standard value. Thus, the TP absorption characters of **2a** and **3a** were determined to be ~ 70 GM at 740 nm and ~ 160 GM at 800 nm, respectively (Figure 3). The TP efficiencies obtained for photochemical decomposition were 20.7 GM at 740 nm for **2a** and 7.8 GM at 800 nm for **3a**.

Measurement of Ca^{2+} Binding Affinities. The dissociation constants, K_d , of Ca^{2+} ions with **2c** and **3c** ($R = H$) were determined by a titration method using fluo-3 fluorescent dye^{44,45} (Figures 5 and 6). Thus, the concentration of free Ca^{2+} ions, $[Ca^{2+}]_F$, after treating 1.0 mM solutions of **2c** and **3c** was determined by fluorescence intensity at 530 nm from fluo-3 (for a detailed procedure, see Experimental Section). The Scatchard analyses shown in Figures 5 and 6 provided dissociation constants of $K_d = 272$ nM for **2c** and $K_d = 440$ nM for **3c**.

Physiologic Studies of 2b In Vivo. Because compound **2a** had a TP efficiency larger than that of **3a**, we loaded cultured neurons with AM ester **2b** (100 μM , 35 °C for 40 min) to stimulate presynaptic terminals with TP uncaging. We recorded inhibitory postsynaptic currents (IPSCs) from whole-cell-clamped neurons by irradiating with a 720 nm-wavelength femtosecond laser (140 fs, 7.6 mW, 1 ms) at 100 points, encompassing a small dendritic region (Figure 4a). The mapping experiments were repeated 81 times in four cells. We recorded IPSCs selectively at dendrite edges, where synapses are most frequently formed. Laser irradiation seldom induced currents over numerous uncaging events on the

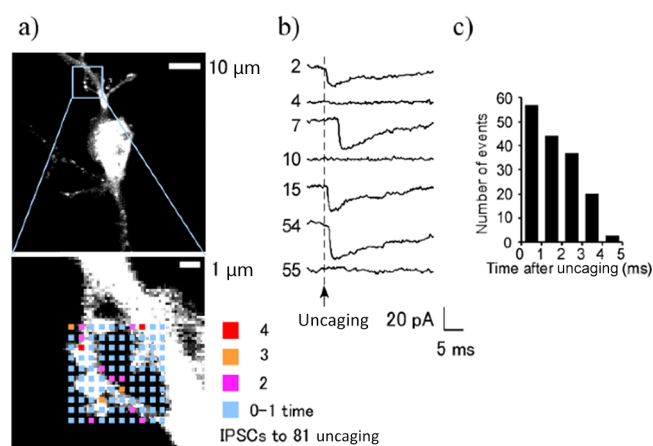


Figure 4. (a) Fluorescence image of a dissociated cultured neuron from cerebral cortices of mice loaded with compound **2b** and a whole cell clamped with Alexa 594 solution. Color squares in the small dendritic region shown in the lower panel indicate the 100 uncaging points separated by $0.55 \mu m$ for TP uncaging and mapping of the IPSCs. The mapping was repeated 81 times. The IPSCs from each point were recorded 0–4 times out of 81 sequences, as denoted by color. (b) IPSCs recorded from the single uncaging point of the cell where four IPSCs were recorded. The sequence numbers are shown next to the current traces. (c) Latency histogram for the onset of all uncaging-induced IPSCs recorded from four cells.

dendrites (Figure 4b), supporting that the IPSCs are not caused by damages of the plasma membrane in the presynaptic terminals. The time course and amplitude of the uncaging-induced IPSCs were similar to those of the spontaneous IPSCs, except that they were elicited shortly after uncaging (Figure 4b,c). The mean latency of 2.5 ms (± 0.4 ms, $n = 150$) was similar to those of IPSCs evoked by electrical stimulation. Uncaging-induced IPSCs were recorded in only up to four out of 81 sequences applied to the dendrite, indicating that the increase in $[Ca^{2+}]_i$ was smaller than that induced by action potentials. Thus, we have succeeded in stimulating presynaptic terminals directly by TP uncaging to elicit neurotransmitter release, whereas further improvements to the TP excitation cross section are necessary to make the most of presynaptic terminal stimulation without the action potential.

CONCLUSIONS

In this study, we designed and synthesized 5-bromo-2-nitrobenzyl-substituted EGTA as a platform for a new generation of calcium chelators with TPA properties. The efficient synthesis of this new bromide was accomplished in nine steps from a commercially available starting material. New calcium chelators with TPA properties, that is, TP fragmentation efficiencies of $\delta_u = 20.7$ GM at 740 nm for NPBF-EGTA ($K_d = 272$ nM) and $\delta_u = 7.8$ GM at 800 nm for BP-EGTA ($K_d = 440$ nM) derivatives, were synthesized using a Suzuki–Miyaura reaction to couple the bromide with benzofuran-2-boronic acid and 4-(dimethylamino)phenyl boronic acid, respectively. The corresponding AM esters were synthesized and successfully applied to a Ca^{2+} -uncaging reaction triggered by photolysis in vivo.

EXPERIMENTAL SECTION

All of the reagents and solvents were reagent grade and used without further purification. Thin-layer chromatography (TLC)

analysis was performed on silica gel plates and imaged under ultraviolet light. ^1H NMR and ^{13}C NMR data were recorded with a 400 MHz NMR spectrometer. CD_3OD and CDCl_3 [0.03% trimethylsilane (TMS)] were used as deuterated solvents. Chemical shifts are described in parts per million (ppm) relative to TMS ($\delta = 0.00$ ppm), and the coupling constants (J) are stated in hertz (Hz). High-resolution mass spectroscopy (HRMS) analyses were conducted using an Orbitrap XL instrument in the positive ion mode.

The syntheses of **1a**, **2a**, **3a**, **2b**, and **3b** are described below.

1-(5-Bromo-2-nitrophenyl)ethan-1-one (5). A flask was charged with KNO_3 (0.28 g, 2.76 mmol) and cooled to 0°C , to which conc. H_2SO_4 (5 mL) was added slowly, and the mixture was stirred for 10 min. 3-Bromoacetophenone (**4**) (0.5 g, 2.51 mmol) in CH_2Cl_2 (5 mL) was added at 0°C . The temperature was raised to the RT, and the mixture was stirred for 4 h. Subsequently, the reaction mixture (RM) was poured into crushed ice. An aqueous layer was extracted with CH_2Cl_2 . The organic layer was washed with brine, dried over Na_2SO_4 , and purified using silica gel column chromatography to give the required nitro product as a white color solid (400 mg, 64%). mp $96\text{--}98^\circ\text{C}$. IR ν 1710, 1558, 1521, 1345, 1308, 1241, 1087, 900, 858, 830, 756, 694 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): δ 8.02 (d, $J = 8.7$ Hz, 1H), 7.75 (dd, $J = 8.7$ and 2.1 Hz, 1H), 7.56 (d, $J = 2.1$ Hz, 1H), 2.57 (s, 3H). ^{13}C NMR (100 MHz, CDCl_3): δ 198.20, 144.31, 139.43, 133.60, 130.34, 129.52, 125.93, 30.21. HRMS-ESI calcd for $\text{C}_8\text{H}_6\text{BrNO}_3\text{Na}$ [$M + \text{Na}$] $^+$ 265.94233, found 265.94223.

2-Bromo-1-(5-bromo-2-nitrophenyl)ethan-1-one (S1). To a solution of **5** (3.7 g, 15.16 mmol) in $\text{AcOH-CH}_2\text{Cl}_2$ (1:2, 240 mL) was added bromine (2.42 g, 15.2 mmol), and the RM was stirred at RT for 55 h. The solution was evaporated to give **S1** (4.89 g, 99%). mp $100\text{--}103^\circ\text{C}$. IR ν 1720, 1560, 1540, 1520, 1347, 1185, 861, 831, 701 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): δ 8.12 (d, $J = 8.8$ Hz, 1H), 7.81 (dd, $J = 8.8$ and 2.1 Hz, 1H), 7.65 (d, $J = 2.1$ Hz, 1H), 4.3 (s, 2H). ^{13}C NMR (100 MHz, CDCl_3): δ 192.82, 144.07, 136.36, 134.31, 132.11, 130.33, 125.81, 33.46. MS-ESI 321.20.

2-(5-Bromo-2-nitrophenyl) Oxirane (6). To a solution of **S1** (1.3 g, 4.02 mmol) in dioxane-MeOH (3:1, 40 mL) was added NaBH_4 (156 mg, 4.02 mmol) portionwise at 0°C , and the RM was stirred for 30 min at the same temperature. The mixture was treated with aqueous NaOH (25 mL, 2.5 mol/L) and stirred for 30 min at 0°C and then evaporated. The residue was diluted with CH_2Cl_2 (40 mL), washed with water and brine solution, dried over Na_2SO_4 , and evaporated. Flash column chromatography (hexane/EtOAc = 10:1) gave a white color solid **6** (710 mg, 71%). mp $72\text{--}75^\circ\text{C}$. IR ν 1605, 1565, 1524, 1514, 1187, 1097, 990, 898, 864, 835, 756, 699 cm^{-1} . ^1H NMR (400 MHz, CDCl_3): δ 8.06 (d, $J = 8.7$ Hz, 1H), 7.8 (d, $J = 2.2$ Hz, 1H), 7.63 (dd, $J = 8.7$ and 2.2 Hz, 1H), 4.5 (dd, $J = 4.2$ and 2.5 Hz, 1H), 3.33 (dd, $J = 5.3$ and 4.4 Hz, 1H), 2.7 (dd, $J = 5.4$ and 2.5 Hz, 1H). ^{13}C NMR (100 MHz, CDCl_3): δ 146.48, 136.66, 131.83, 130.25, 129.84, 126.27, 50.68, 50.34. HRMS-APCI calcd for $\text{C}_8\text{H}_7\text{BrNO}_3\text{Na}$ [$M + \text{H}$] $^+$ 243.96038, found 243.96060.

2-Azido-1-(5-bromo-2-nitrophenyl)ethan-1-ol (7). To a solution of oxirane **6** (400 mg, 1.63 mmol) in freshly distilled dry DMF (5 mL) was added NaN_3 (106 mg, 1.63 mmol), and the RM was stirred at 40°C for 4 h. The RM was diluted with EtOAc, washed with water and brine solution, dried over Na_2SO_4 , and evaporated. Flash column chromatography (hexane/EtOAc = 10:3) gave a yellow color oil **7** (120 mg,

40%). IR ν 3500, 3100, 2946, 2870, 2130, 1604, 1570, 1533, 1362, 1321, 1176, 1100, 1061, 907, 848, 758 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): δ 8.11 (d, $J = 2.2$ Hz, 1H), 7.94 (d, $J = 8.8$ Hz, 1H), 7.64 (dd, $J = 8.8$ and 2.2 Hz, 1H), 5.55 (m, 1H), 3.77 (dd, $J = 12.5$ and 2.8 Hz, 1H), 3.45 (dd, $J = 12.5$ and 7.5 Hz, 1H), 2.74 (d, $J = 3.9$ Hz, 1H). ^{13}C NMR (100 MHz, CDCl_3): δ 146.02, 138.18, 132.22, 131.97, 129.40, 126.37, 68.80, 57.04. HRMS-ESI calcd for $\text{C}_8\text{H}_7\text{BrN}_4\text{O}_3\text{Na}$ [$M + \text{Na}$] $^+$ 284.96288, found 284.96597.

2-(2-Azido-1-(2-(2-bromoethoxy)ethoxy)ethyl)-4-bromo-1-nitrobenzene (8). To a stirred suspension of **7** (600 mg, 2.09 mmol) and cesium carbonate (3.39 g, 10.45 mmol) in freshly distilled dry acetonitrile (10 mL) was added bis (2-bromomethyl) ether (2.42 g, 10.45 mmol). The mixture was stirred for 5 h at 50°C . Then, the RM was filtered, and the filtrate crude was evaporated using purified flash column chromatography (hexane/EtOAc = 10:2), which gave a yellow color oil **8** (400 mg, 53%). IR ν 2930, 2880, 2114, 1602, 1561, 1530, 1465, 1362, 1303, 1120, 957, 848, 757, 670 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): δ 8.06 (d, $J = 2.2$ Hz, 1H), 7.93 (d, $J = 8.6$ Hz, 1H), 7.63 (dd, $J = 8.6$ and 2.2 Hz, 1H), 5.26 (dd, $J = 7.2$ and 2.8 Hz, 1H), 3.8–3.4 (m, 10H). ^{13}C NMR (100 MHz, CDCl_3): δ 146.90, 137.05, 132.29, 132.19, 129.43, 126.48, 77.49, 71.27, 70.47, 69.52, 55.61, 30.20. HRMS-ESI calcd for $\text{C}_{12}\text{H}_{14}\text{Br}_2\text{N}_4\text{O}_4\text{Na}$ [$M + \text{Na}$] $^+$ 458.92740, found 458.92767.

2-(2-Azido-1-(2-(2-azidoethoxy)ethoxy)ethyl)-4-bromo-1-nitrobenzene (S2). A solution of **8** (400 mg, 0.91 mmol) and sodium azide (118 mg, 1.82 mmol) in dry DMF (4 mL) was stirred at RT for 16 h. Then the RM was diluted with EtOAc, washed with water and brine solution, dried over Na_2SO_4 , and evaporated. Flash column chromatography (hexane/EtOAc = 10:3) gave a yellow color oil **S2** (295 mg, 80%). IR ν 2937, 2882, 2127, 1597, 1560, 1538, 1449, 1370, 1320, 1170, 848, 755 cm^{-1} . ^1H NMR (400 MHz, CDCl_3): δ 8.08 (d, $J = 2.2$ Hz, 1H), 7.95 (d, $J = 8.7$ Hz, 1H), 7.65 (dd, $J = 8.7$ and 2.2 Hz, 1H), 5.28 (dd, $J = 7.2$ and 2.9 Hz, 1H), 3.76–3.58 (m, 6H), 3.56–3.39 (m, 4H). ^{13}C NMR (100 MHz, CDCl_3): δ 146.90, 137.04, 132.28, 132.17, 129.41, 126.48, 77.49, 70.58, 70.17, 69.55, 55.59, 50.68 cm^{-1} . HRMS-ESI calcd for $\text{C}_{12}\text{H}_{14}\text{BrN}_7\text{O}_4\text{Na}$ [$M + \text{Na}$] $^+$ 422.01829, found 422.01773.

2-(2-(2-Aminoethoxy)ethoxy)-2-(5-bromo-2-nitrophenyl)ethan-1-amine (9). To a solution of compound **S2** (520 mg, 1.3 mmol) in THF (5 mL) was added PPh_3 (1.02 g, 3.9 mmol), and the RM was stirred for 12 h at RT. Then H_2O (0.5 mL) was added, and the resulting solution was stirred for 12 h. The solvent was evaporated, and the reaction products were purified using flash column chromatography ($\text{CH}_2\text{Cl}_2/\text{MeOH}/\text{Et}_3\text{N} = 7:2:1$), which gave a yellow color liquid **9** (360 mg, 80%). IR ν 3442, 2933, 1531, 1359, 1129, 845, 658 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): δ 7.99 (d, $J = 2.1$ Hz, 1H), 7.92 (d, $J = 8.7$ Hz, 1H), 7.61 (dd, $J = 8.7$ and 2.1 Hz, 1H), 5.1 (dd, $J = 8.1$ and 2.6 Hz, 1H), 3.72–3.51 (m, 6H), 3.33–2.82 (m, 4H), 2.03 (bs, 2H), 1.41 (t, $J = 7.5$ Hz, 2H). ^{13}C NMR (100 MHz, CDCl_3): δ 147.22, 138.14, 131.95, 131.68, 129.20, 126.45, 77.23, 71.1, 70.12, 69.13, 47.33, 40.88. HRMS-ESI calcd for $\text{C}_{12}\text{H}_{18}\text{BrN}_3\text{O}_4\text{Na}$ [$M + \text{Na}$] $^+$ 370.03729, found 370.03726.

Diethyl-5-(5-bromo-2-nitrophenyl)-3,12-bis(2-ethoxy-2-oxoethyl)-6,9-dioxo-3,12-diazatetradecanedioate (1a). To a solution of **9** (800 mg, 2.3 mmol), sodium iodide (3.45 g, 23.05 mmol), and diisopropyl ethylamine (1.48 g, 11.52 mmol) in dry CH_3CN (30 mL), ethyl bromoacetate (3.85 g, 23.05 mmol) was added dropwise at 0°C , and the RM was stirred for

3 h at 80 °C. The RM was filtered, and the filtrate was evaporated. Flash column chromatography (hexane/EtOAc = 2:1) gave a pale yellow color oil **1a** (1.1 g, 70%). IR ν 2985, 1752, 1600, 1530, 1211, 1045, 759, 602 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): δ 7.98 (d, J = 2.1 Hz, 1H), 7.86 (d, J = 8.7 Hz, 1H), 7.54 (dd, J = 8.7 and 2.1 Hz, 1H), 5.2 (dd, J = 7.2 and 3.3 Hz, 1H), 4.19–4.11 (m, 8H), 3.78–3.67 (m, 4H), 3.62–3.38 (m, 10H), 3.09–2.91 (m, 4H), 1.29–1.22 (m, 12H). ^{13}C NMR (100 MHz, CDCl_3): δ 171.70, 171.41, 146.97, 139.13, 132.10, 131.43, 128.98, 126.16, 77.80, 70.32, 70.18, 69.14, 61.17, 60.45, 55.98, 55.73, 53.84, 14.27. HRMS-ESI calcd for $\text{C}_{28}\text{H}_{42}\text{BrN}_3\text{O}_{12}\text{Na}$ [$M + \text{Na}$] $^+$ 714.18441, found 714.18518.

Diethyl-5-(5-benzofuran-2-yl)-2-nitrophenyl)-3,12-bis(2-ethoxy-2-oxoethyl)-6,9-dioxa-3,12-diazatetradecanedioate (2a). A solution of **1a** (200 mg, 0.28 mmol), benzofuran-2-boronic acid (52.4 mg, 0.31 mmol), $\text{Pd}(\text{PPh}_3)_4$ (16.7 mg, 0.14 mmol), and potassium carbonate (52 mg, 0.31 mmol) was taken in 10 mL THF/ H_2O (9:1) in a round-bottom flask and was stirred for 3 h at 60 °C. Then, the reaction mixture was filtered. The filtrate was washed with water, extracted with EtOAc organic layer, dried over Na_2SO_4 , evaporated under reduced pressure, and purified using column chromatography to afford the desire compound as a yellow oil **2a** (95%). IR ν 2974, 2903, 2859, 1730, 1610, 1515, 1445, 1346, 1255, 1178, 1024, 746. ^1H NMR (400 MHz, CDCl_3): δ 8.29 (s, 1H), 8.1 (d, J = 8.6 Hz, 1H), 7.9 (dd, J = 2.0 and 8.6 Hz, 1H), 7.68 (d, J = 7.7 Hz, 1H), 7.6 (d, J = 8.1 Hz, 1H), 7.4–7.29 (m, 3H), 5.34–5.3 (m, 1H), 4.22–4.07 (m, 8H), 3.8 (s, 4H), 3.65–3.5 (m, 10H), 3.17–2.9 (m, 4H), 1.31–1.3 (m, 12H). ^{13}C NMR (100 MHz, CDCl_3): δ 171.82, 171.34, 170.19, 155.42, 147.24, 128.74, 125.68, 124.77, 124.04, 123.45, 121.65, 111.53, 105.18, 78.16, 77.35, 77.24, 77.03, 76.72, 70.29, 69.06, 61.30, 60.42, 55.87, 55.75, 53.71, 21.08, 14.28, 14.22. HRMS-ESI calcd for $\text{C}_{36}\text{H}_{47}\text{N}_3\text{O}_{13}\text{Na}$ [$M + \text{Na}$] $^+$ 752.30011, found 752.30011.

Diethyl-5-(4'-(dimethylamino)-4-nitro-[1,1'-biphenyl]-3-yl)-3,12-bis(2-ethoxy-2-oxoethyl)-6,9-dioxa-3,12-diazatetradecanedioate (3a). A solution of **1a** (200 mg, 0.28 mmol), 4-(*N,N*-dimethyl amino)phenylboronic acid (52.4 mg, 0.31 mmol), $\text{Pd}(\text{PPh}_3)_4$ (16.7 mg, 0.14 mmol), and potassium carbonate (60 mg, 0.43 mmol) was taken in 10 mL THF/ H_2O (9:1) in a round-bottom flask and was stirred for 3 h at 60 °C. Then, the reaction mixture was filtered. The filtrate was washed with water, extracted with EtOAc organic layer, dried over Na_2SO_4 , evaporated under reduced pressure, and purified using column chromatography to afford the desire compound as a yellow oil **3a** (90%): ^1H NMR (400 MHz, CDCl_3): δ 8.27 (d, J = 2.1 Hz, 1H), 7.84 (d, J = 8.6 Hz, 1H), 7.64–7.6 (m, 2H), 7.19 (dd, J = 2.2 and 2.1 Hz, 1H), 6.71–6.67 (m, 2H), 5.32 (dd, J = 8.3 and 2.3 Hz, 1H), 4.03–3.84 (m, 12H), 3.65–3.62 (m, 4H), 3.48–3.2 (m, 8H), 3.16–2.85 (m, 8H), 0.95 (t, J = 7.2 Hz, 6H), 0.89 (t, J = 7.2 Hz, 6H). ^{13}C NMR (100 MHz, CDCl_3): δ 171.80, 171.31, 150.95, 146.67, 145.60, 137.69, 128.11, 125.83, 125.68, 125.25, 124.89, 112.49, 78.28, 77.22, 70.33, 70.24, 68.83, 61.46, 60.39, 60.37, 55.88, 55.68, 53.73, 40.31, 14.28, 14.22. HRMS-ESI calcd for $\text{C}_{36}\text{H}_{52}\text{N}_4\text{O}_{12}\text{Na}$ [$M + \text{Na}$] $^+$ 755.34739, found 755.34631.

Bis(acetoxymethyl) 3,12-Bis(2-(acetoxymethoxy)-2-oxoethyl)-5-(5-(benzofuran-2-yl)-2-nitrophenyl)-6,9-dioxa-3,12-diazatetradecanedioate (2b). To a compound **2a** (50 mg, 0.068 mmol) in 3 mL of MeOH was added potassium hydroxide (KOH) (19.2 mg, 0.34 mmol) at 0 °C. The RM was stirred for 2 h at 60 °C, evaporated under reduced

pressure, and dried over. The yellow color solid obtained was dissolved in freshly distilled dry DMF (3 mL). Diisopropylethyl amine (52 mg, 0.4 mmol) and bromomethylacetate (166 mg, 1.08 mmol) were added at 0 °C. After 16 h of stirring at RT, the RM was diluted with water and extracted with ethyl acetate. The organic layer was washed with water and brine solution, dried over Na_2SO_4 , and evaporated under reduced pressure. The reaction products were purified using silica gel column chromatography and further purified by recycling HPLC with gel permeation chromatography (GPC) column to give pure AM ester as a yellow color oil **2b** (60%). ^1H NMR (400 MHz, CDCl_3): δ 8.27 (d, J = 2.0 Hz, 1H), 8.16 (d, J = 8.7 Hz, 1H), 7.93 (dd, J = 8.6 and 2.0 Hz, 1H), 7.68 (d, J = 8.6 Hz, 1H), 7.61 (d, J = 8.2 Hz, 1H), 7.4–7.37 (m, 1H), 7.31–7.24 (m, 2H), 5.79 (s, 4H), 5.9 (s, 4H), 5.31 (dd, J = 7.9 and 2.45 Hz, 1H), 3.94–3.84 (m, 4H), 3.67–3.47 (m, 10H), 3.23–3.09 (m, 2H), 2.99–2.86 (m, 2H), 2.13 (s, 6H), 2.08 (s, 6H). ^{13}C NMR (100 MHz, CDCl_3): δ 170.51, 170.01, 169.73, 169.51, 155.45, 153.30, 147.12, 137.80, 135.66, 132.12, 132.10, 128.61, 128.58, 125.74, 124.52, 124.11, 123.53, 121.60, 111.57, 105.30, 79.14, 78.17, 77.25, 70.34, 70.29, 69.05, 61.10, 55.43, 55.36, 53.53, 20.77, 20.71. HRMS-ESI calcd for $\text{C}_{40}\text{H}_{47}\text{N}_3\text{O}_{21}\text{Na}$ [$M + \text{Na}$] $^+$ 928.25943, found 928.25897.

Bis(acetoxymethyl) 3,12-Bis(2-(acetoxymethoxy)-2-oxoethyl)-5-(4'-(dimethylamino)-4-nitro-[1,1'-biphenyl]-3-yl)-6,9-dioxa-3,12-diazatetradecanedioate (3b). To the compound **3a** (50 mg, 0.068 mmol) in 3 mL of MeOH was added KOH (19.1 mg, 0.34 mmol) at 0 °C. The RM was stirred for 2 h at 60 °C, evaporated under reduced pressure, and dried over. The yellow color solid obtained was dissolved in freshly distilled dry DMF (3 mL). Diisopropylethyl amine and bromomethylacetate were added at 0 °C. After 16 h of stirring at RT, the RM was diluted with water and extracted with ethyl acetate. The organic layer was washed with water and brine solution, dried over Na_2SO_4 , and evaporated under reduced pressure. The reaction products were purified using silica gel column chromatography and further purified by recycling HPLC with GPC column to give pure AM ester as a yellow color oil **3b** (60%). ^1H NMR (400 MHz, CDCl_3): δ 8.09 (d, J = 8.7 Hz, 1H), 7.96 (d, J = 2.1 Hz, 1H), 7.63–7.57 (m, 3H), 6.84–6.8 (m, J = 8.9, 2H), 5.79 (s, 4H), 5.72 (s, 4H), 5.32 (dd, J = 8.3 and 1.8 Hz, 1H), 3.91 (s, 4H), 3.65–3.44 (m, 10H), 3.23–3.02 (m, 8H), 2.97–2.84 (m, 2H), 2.14 (s, 6H), 2.1 (s, 6H). ^{13}C NMR (100 MHz, CDCl_3): δ 170.63, 170.10, 169.71, 169.58, 150.97, 146.71, 145.45, 137.41, 128.19, 125.84, 125.63, 125.03, 112.46, 79.12, 78.25, 77.23, 70.31, 68.74, 61.19, 55.41, 55.20, 53.63, 40.31, 20.79, 20.72. HRMS-ESI calcd for $\text{C}_{40}\text{H}_{52}\text{N}_4\text{O}_{20}\text{Na}$ [$M + \text{Na}$] $^+$ 931.30671, found 931.30420.

5-(5-(Benzofuran-2-yl)-2-nitrophenyl)-3,12-bis(carboxymethyl)-6,9-dioxa-3,12-diazatetradecanedioic acid (2c). Tetraethyl ester **2a** (100 mg, 0.13 mmol) was dissolved in methanol (4 mL) and then saponified by the addition of KOH (38.4 mg, 0.68 mmol) and stirring for 2 h at 60 °C. After evaporation of methanol and acidification with 2 N aqueous HCl to pH 2, the yellow solid obtained was collected and dried to afford **2c** (92%). ^1H NMR (400 MHz, CD_3OD): δ 8.32–8.30 (bs, 1H), 8.24 (d, J = 8.6 Hz, 1H), 8.13 (dd, J = 1.8 and 8.6 Hz, 1H), 7.71 (d, J = 7.5 Hz, 1H), 7.62–7.56 (m, 2H), 7.4–7.28 (m, 2H), 5.65 (m, 1H), 4.5 (bs, 4H), 4.15 (bs, 4H), 4.0–3.5 (m, 10H). ^{13}C NMR (100 MHz, CD_3OD): 168.86, 168.47, 157.06, 154.37, 148.86, 137.65, 135.27, 130.31, 127.77, 127.47, 127.27, 125.71, 125.05, 123.30, 112.58, 107.5, 74.69, 71.48, 69.75, 66.85, 66.81, 60.87, 57.43, 57.38, 56.66, 54.02.

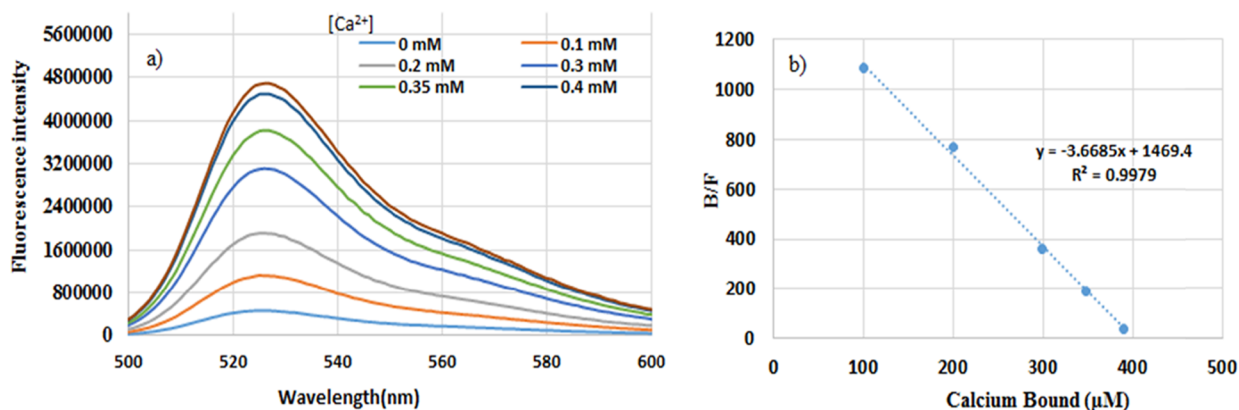


Figure 5. (a) Emission spectra of fluo-3 (10 μM , excitation at 490 nm) resulting from the titration of **2c** (1.0 mM in Hepes/100 mM KCl, pH 7.4 at 298 K) with incremental addition of Ca^{2+} . (b) Scatchard analysis (using K_d value of 500 nM for fluo-3) of these data.

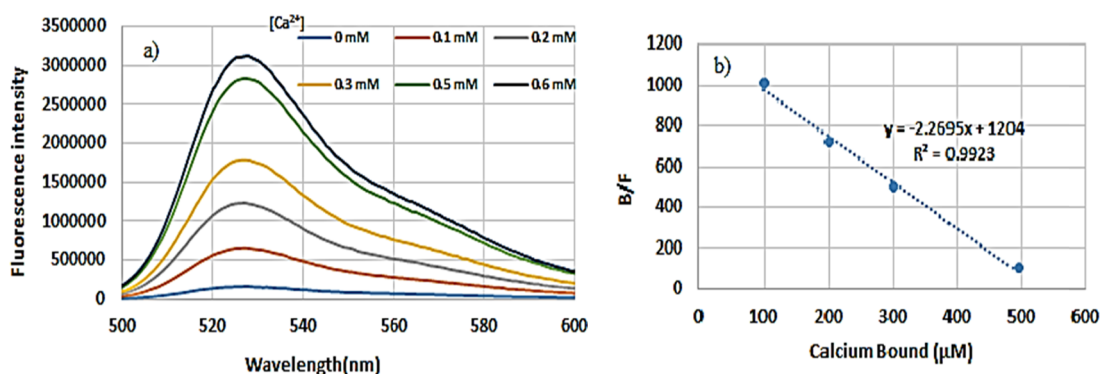


Figure 6. (a) Emission spectra of fluo-3 (10 μM , excitation at 490 nm) resulting from the titration of **3c** (1.0 mM in Hepes/100 mM KCl, pH 7.4 at 298 K) with incremental addition of Ca^{2+} . (b) Scatchard analysis (using K_d value of 500 nM for fluo-3) of these data.

HRMS-ESI calcd for $\text{C}_{28}\text{H}_{31}\text{N}_3\text{O}_{13} [\text{M} - \text{H}]^+$ 616.17841, found 616.17767.

3,12-Bis(carboxymethyl)-5-(4'-(dimethylamino)-4-nitro-[1,1'-biphenyl]-3-yl)-6,9-dioxo-3,12-diazatetradecanedioic acid (3c**).** Tetraethyl ester **3a** (100 mg, 0.13 mmol) was dissolved in methanol (4 mL) and then saponified by the addition of KOH (38.3 mg, 0.68 mmol) and stirring for 2 h at 60 $^\circ\text{C}$. After evaporation of methanol and acidification with 2 N aqueous HCl to pH 2, the yellow solid obtained was collected and dried to afford **3c** (90%). ^1H NMR (400 MHz, CD_3OD): δ 8.15 (d, $J = 8.4$ Hz, 1H), 8.05 (s, 1H), 7.95–7.85 (m, 3H), 7.81–7.65 (m, 2H), 5.62–5.58 (m, 1H), 4.46 (s, 2H), 4.45 (s, 2H), 4.17 (s, 2H), 4.16 (s, 2H), 3.94–3.73 (m, 4H), 3.66–3.45 (m, 6H), 3.27 (s, 6H). ^{13}C NMR (100 MHz, CD_3OD): 168.72, 168.42, 149.21, 146.64, 145.04, 141.1, 134.77, 130.94, 130.05, 128.67, 127.68, 112.80, 74.67, 71.44, 69.73, 66.91, 60.91, 57.34, 56.67, 47.27. HRMS-ESI calcd for $\text{C}_{28}\text{H}_{36}\text{N}_4\text{O}_{12} [\text{M} + \text{H}]^+$ 621.24025, found 621.24084.

Calcium Binding Affinities. The association constant for the two chelators **2c** and **3c** was calculated using the method of Ellis-Davies et al. by the equation

$$[\text{Ca}^{2+}]_{\text{B}}/[\text{Ca}^{2+}]_{\text{F}} = K_a([\text{chelator}]_{\text{T}} - [\text{Ca}^{2+}]_{\text{B}})$$

Here $[\text{Ca}^{2+}]_{\text{B}}$ is the concentration of Ca^{2+} bound to the chelator, $[\text{Ca}^{2+}]_{\text{F}}$ is the concentration of the free Ca^{2+} , and $[\text{chelator}]_{\text{T}}$ is the total concentration of the chelator (NPBF-EGTA acid or BP-EGTA acid) in the solution. The free and bound calcium concentrations were obtained from the emission

spectra of a calcium-sensitive dye (fluo-3, $K_d = 500$ nm) through titration with calcium chloride solution.

In this measurement, chelators (1.0 mM), fluo-3 (0.01 mM), Hepes (10 mM) pH 7.4, and 100 mM KCl solutions were used. A 3.0 mL solution of 1.0 mM chelator was used. The solution was buffered to pH 7.4 with Hepes, and the ionic strength was set with 100 mM KCl. Data from the titration with incremental addition of Ca^{2+} are shown in Figures 5 and 6.

$[\text{Ca}^{2+}]_{\text{B}}/[\text{Ca}^{2+}]_{\text{F}}$ were obtained from the absorption curves shown in Figures 5 and 6. The concentration of chelators was 1.0 mM, and the CaCl_2 concentrations were 0.1, 0.2, 0.3, 0.35, 0.4, and 0.5 mM for NPBF-EGTA acid and 0.1, 0.2, 0.3, 0.5, and 0.6 mM for BP-EGTA acid. After these concentrations, any change was not observed in the absorption curve.

The free calcium concentrations were calculated from the equation

$$[\text{Ca}^{2+}]_{\text{F}} = K_d(A_{\text{int}} - A_{\text{min}})/(A_{\text{max}} - A_{\text{int}})$$

where $K_d = 500$ nM (fluo-3), A_{int} = absorbance of chelator at setted Ca^{2+} concentration, A_{min} = absorbance of chelator without Ca^{2+} , and A_{max} = absorbance of chelator with excess Ca^{2+} .

Dissociated Cultured Neurons, Electrophysiology, and Two-Photon Uncaging. Cerebral cortices from male C57BL/6 mouse brains (E17.5) with 0.15% trypsin in ethylenediaminetetraacetic acid (Invitrogen, Carlsbad, CA, USA) and DNase I (30 mg mL^{-1}) were used. The cells were plated on laminin- and poly-D-lysine-coated cover glasses with a density of 1.7×10^5 cells per square centimeter. The neurons

were maintained in a serum-free medium (Neurobasal Medium; Invitrogen) supplemented with 1% GlutaMAX and 2% B-27 supplement (Invitrogen). For electrophysiological experiments, an extracellular solution was based on SoLA containing 10 mM CaCl₂, 100 mM cyclothiazide, 50 mM picrotoxin, and 320 nM Na-pyruvate, at pH 7.4. The cultured cells were first loaded with the compound **2a** (100 μM) for 40 min in a solution containing CrEL (0.03%, Invitrogen) and bovine serum albumin (5%) in SoLA, followed by washing with SoLA for 20 min at 35 °C. The postsynaptic cells (DIV 10–15) were whole-cell-clamped (holding potential = -70 mV) and perfused with intracellular solution (in mM: CsCl 140, Hepes 10, MgATP 4, Na₂GTP 0.3, Na₂-phosphocreatine 10, Alexa-594 0.025, pH = 7.25 with CsOH). Electrical stimulation of 70 μA amplitude was applied, at 100–150 μm from the dendrites.

Two-Photon Excitation Imaging and Uncaging. The cultured neurons were placed on an inverted laser-scanning microscope (FV1000 and IX81; Olympus Corporation, Tokyo, Japan) equipped with a water-immersion objective lens (UPlanApo60xW/IR; N.A. 1.2; Olympus Corporation) and a femtosecond laser (Mai Tai; Spectra Physics Inc., Mountain View, CA, USA). Tunable wavelengths were set at 950 and 720 nm for imaging and uncaging, respectively.

■ ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acsomega.6b00119.

Experimental procedures and compound characterization (PDF)

■ AUTHOR INFORMATION

Corresponding Authors

*E-mail: mabe@hiroshima-u.ac.jp (M.A.).

*E-mail: hkasai@m.u-tokyo.ac.jp (H.K.).

Notes

The authors declare no competing financial interest.

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■ REFERENCES

- (1) Ellis-Davies, G. C. R. Neurobiology with caged calcium. *Chem. Rev.* **2008**, *108*, 1603–1613.
- (2) Crowe, S. E.; Kantevari, S.; Ellis-Davies, G. C. R. Photochemically initiated intracellular astrocytic calcium waves in living mice using two-photon uncaging of IP₃. *ACS Chem. Neurosci.* **2010**, *1*, 575–585.
- (3) Zucker, R. In *Calcium in Living Cells, Methods in Cell Biology*; Whitaker, M., Ed.; Elsevier, 2010; Vol. 99, Chapter 2.
- (4) Ellis-Davies, G. C.; Kaplan, J. H. Nitrophenyl-EGTA, a photolabile chelator that selectively binds Ca²⁺ with high affinity and releases it rapidly upon photolysis. *Proc. Natl. Acad. Sci. U.S.A.* **1994**, *91*, 187–191.
- (5) Momotake, A.; Lindegger, N.; Niggli, E.; Barsotti, R. J.; Ellis-Davies, G. C. R. The nitrodibenzofuran chromophore: A new caging group for ultra-efficient photolysis in living cells. *Nat. Methods* **2006**, *3*, 35–40.
- (6) Brieke, C.; Rohrbach, F.; Gottschalk, A.; Mayer, G.; Heckel, A. Light-controlled tools. *Angew. Chem., Int. Ed.* **2012**, *51*, 8446–8476.

(7) Bort, G.; Gallavardin, T.; Ogden, D.; Dalko, P. I. From one-photon to two-photon probes: "Caged" compounds, actuators, and photoswitches. *Angew. Chem., Int. Ed.* **2013**, *52*, 4526–4537.

(8) Kim, H. M.; Cho, B. R. Small molecule two-photon probes for bioimaging applications. *Chem. Rev.* **2015**, *115*, 5014–5055.

(9) Klán, P.; Šolomek, T.; Bochet, C. G.; Blanc, A.; Givens, R.; Rubina, M.; Popik, V.; Kostikov, A.; Wirz, J. Photoremovable protecting groups in chemistry and biology: Reaction mechanisms and efficacy. *Chem. Rev.* **2013**, *113*, 119–191.

(10) Tsien, R. Y.; Zucker, R. S. Control of cytoplasmic calcium with photolabile tetracarboxylate 2-nitrobenzhydrol chelators. *Biophys. J.* **1986**, *50*, 843–853.

(11) Cui, J.; Gropeanu, R. A.; Stevens, D. R.; Rettig, J.; del Campo, A. New photolabile BAPTA-based Ca²⁺ cages with improved photo-release. *J. Am. Chem. Soc.* **2012**, *134*, 7733–7740.

(12) Amatrudo, J. M.; Olson, J. P.; Lur, G.; Chiu, C. Q.; Higley, M. J.; Ellis-Davies, G. C. R. Wavelength-selective one- and two-photon uncaging of GABA. *ACS Chem. Neurosci.* **2014**, *5*, 64–70.

(13) Donato, L.; Mourot, A.; Davenport, C. M.; Herbivo, C.; Warther, D.; Léonard, J.; Bolze, F.; Nicoud, J.-F.; Kramer, R. H.; Goeldner, M.; Specht, A. Water-soluble, donor–acceptor biphenyl derivatives in the 2-(*o*-nitrophenyl)propyl series: Highly efficient two-photon uncaging of the neurotransmitter γ -aminobutyric acid at $\lambda = 800$ nm. *Angew. Chem., Int. Ed.* **2012**, *51*, 1840–1843.

(14) Svoboda, K.; Yasuda, R. Principles of two-photon excitation microscopy and its applications to neuroscience. *Neuron* **2006**, *50*, 823–839.

(15) Ellis-Davies, G. C. R. Caged compounds: Photorelease technology for control of cellular chemistry and physiology. *Nat. Methods* **2007**, *4*, 619–628.

(16) Specht, A.; Bolze, F.; Omran, Z.; Nicoud, J.-F.; Goeldner, M. Photochemical tools to study dynamic biological processes. *HFSP J.* **2009**, *3*, 255–264.

(17) Pawlicki, M.; Collins, H. A.; Denning, R. G.; Anderson, H. L. Two-photon absorption and the design of two-photon dyes. *Angew. Chem., Int. Ed.* **2009**, *48*, 3244–3266.

(18) Matsuzaki, M.; Ellis-Davies, G. C. R.; Nemoto, T.; Miyashita, Y.; Iino, M.; Kasai, H. Dendritic spine geometry is critical for AMPA receptor expression in hippocampal CA1 pyramidal neurons. *Nat. Neurosci.* **2001**, *4*, 1086–1092.

(19) Ellis-Davies, G. C. R. Two-photon microscopy for chemical neuroscience. *ACS Chem. Neurosci.* **2011**, *2*, 185–197.

(20) Olson, J. P.; Kwon, H.-B.; Takasaki, K. T.; Chiu, C. Q.; Higley, M. J.; Sabatini, B. L.; Ellis-Davies, G. C. R. Optically selective two-photon uncaging of glutamate at 900 nm. *J. Am. Chem. Soc.* **2013**, *135*, 5954–5957.

(21) Boinapally, S.; Huang, B.; Abe, M.; Katan, C.; Noguchi, J.; Watanabe, S.; Kasai, H.; Xue, B.; Kobayashi, T. Caged glutamates with π -extended 1,2-dihydronaphthalene chromophore: Design, synthesis, two-photon absorption property, and photochemical reactivity. *J. Org. Chem.* **2014**, *79*, 7822–7830.

(22) Šolomek, T.; Wirz, J.; Klán, P. Searching for improved photoreleasing abilities of organic molecules. *Acc. Chem. Res.* **2015**, *48*, 3064–3072.

(23) Palao, E.; Slanina, T.; Muchová, L.; Šolomek, T.; Vitek, L.; Klán, P. Transition-metal-free CO-releasing BODIPY derivatives activatable by visible to NIR light as promising bioactive molecules. *J. Am. Chem. Soc.* **2016**, *138*, 126–133.

(24) Ciuciu, A. I.; Korzycka, K. A.; Lewis, W. J. M.; Bennett, P. M.; Anderson, H. L.; Flamigni, L. Model dyads for 2PA uncaging of a protecting group via photoinduced electron transfer. *Phys. Chem. Chem. Phys.* **2015**, *17*, 6554–6564.

(25) Mahmoodi, M. M.; Abate-Pella, D.; Pundsack, T. J.; Palsuledesai, C. C.; Goff, P. C.; Blank, D. A.; Distefano, M. D. Nitrodibenzofuran: A one- and two-photon sensitive protecting group that is superior to brominated hydroxycoumarin for thiol caging in peptides. *J. Am. Chem. Soc.* **2016**, *138*, 5848–5859.

(26) Chen, Z.; Zhou, L.; Bing, W.; Zhang, Z.; Li, Z.; Ren, J.; Qu, X. Light controlled reversible inversion of nanophosphor-stabilized

pickering emulsions for biphasic enantioselective biocatalysis. *J. Am. Chem. Soc.* **2014**, *136*, 7498–7504.

(27) Sedlmeier, A.; Gorris, H. H. Surface modification and characterization of photon-upconverting nanoparticles for bioanalytical applications. *Chem. Soc. Rev.* **2015**, *44*, 1526–1560.

(28) Chen, G.; Qiu, H.; Prasad, P. N.; Chen, X. Upconversion nanoparticles: Design, nanochemistry, and applications in theranostics. *Chem. Rev.* **2014**, *114*, 5161–5214.

(29) Guo, S.; Xie, X.; Huang, L.; Huang, W. Sensitive water probing through nonlinear photon upconversion of lanthanide-doped nanoparticles. *ACS Appl. Mater. Interfaces* **2016**, *8*, 847–853.

(30) Huang, P.; Zheng, W.; Zhou, S.; Tu, D.; Chen, Z.; Zhu, H.; Li, R.; Ma, E.; Huang, M.; Chen, X. Lanthanide-doped LiLuF₄ upconversion nanoproboscopes for the detection of disease biomarkers. *Angew. Chem., Int. Ed.* **2014**, *53*, 1252–1257.

(31) Wong, H.-T.; Tsang, M.-K.; Chan, C.-F.; Wong, K.-L.; Fei, B.; Hao, J. In vitro cell imaging using multifunctional small sized KGdF₄: Yb³⁺, Er³⁺ upconverting nanoparticles synthesized by a one-pot solvothermal process. *Nanoscale* **2013**, *5*, 3465–3473.

(32) Yang, Y.; Shao, Q.; Deng, R.; Wang, C.; Teng, X.; Cheng, K.; Cheng, Z.; Huang, L.; Liu, Z.; Liu, X.; Xing, B. In Vitro and In Vivo Uncaging and Bioluminescence Imaging by Using Photocaged Upconversion Nanoparticles. *Angew. Chem., Int. Ed.* **2012**, *51*, 3125–3129.

(33) Göppert-Mayer, M. Elementary processes with two quantum jumps. *Ann. Phys.* **1931**, *9*, 272–294.

(34) Kiskin, N. I.; Chillingworth, R.; McCray, J. A.; Piston, D.; Ogden, D. The efficiency of two-photon photolysis of a “caged” fluorophore, *o*-1-(2-nitrophenyl)ethylpyranine, in relation to photo-damage of synaptic terminals. *Eur. Biophys. J.* **2002**, *30*, 588–604.

(35) Miyaura, N.; Yanagi, T.; Suzuki, A. The palladium-catalyzed cross-coupling reaction of phenylboronic acid with halogens in the presence of bases. *Synth. Commun.* **1981**, *11*, 513–519.

(36) Ullah, N. Synthesis of 6-piperazinyl-3,4-dihydroquinazolin-2(1H)-ones. *Asian J. Chem.* **2014**, *26*, 2031–2036.

(37) Lupattelli, P.; Bonini, C.; Caruso, L.; Gambacorta, A. Single route to chiral syn- and anti-2-amino-1,2-diphenylethanols via a new stereodivergent opening of trans-1,2-diphenyloxirane. *J. Org. Chem.* **2003**, *68*, 3360–3362.

(38) Hou, J.-L.; Jia, M.-X.; Jiang, X.-K.; Li, Z.-T.; Chen, G.-J. Solvophobicity-driven oligo(ethylene glycol) helical foldamers. Synthesis, characterization, and complexation with ethane-1,2-diaminium. *J. Org. Chem.* **2004**, *69*, 6228–6237.

(39) Colombano, G.; Travelli, C.; Galli, U.; Caldarelli, A.; Chini, M. G.; Canonico, P. L.; Sorba, G.; Bifulco, G.; Tron, G. C.; Genazzani, A. A novel potent nicotinamide phosphoribosyltransferase inhibitor synthesized via click chemistry. *J. Med. Chem.* **2010**, *53*, 616–623.

(40) Shan, M.; Carlson, K. E.; Bujotzek, A.; Wellner, A.; Gust, R.; Weber, M.; Katzenellenbogen, J. A.; Haag, R. Nonsteroidal bivalent estrogen ligands: An application of the bivalent concept to the estrogen receptor. *ACS Chem. Biol.* **2013**, *8*, 707–715.

(41) Gould, I. R.; Lenhard, J. R.; Farid, S. A curve-crossing model for oxidative decarboxylation. Kinetics of anilino carboxylate fragmentations. *J. Phys. Chem. A* **2004**, *108*, 10949–10956.

(42) Komori, N.; Jakkampudi, S.; Motoishi, R.; Abe, M.; Kamada, K.; Furukawa, K.; Katan, C.; Sawada, W.; Takahashi, N.; Kasai, H.; Xue, B.; Kobayashi, T. Design and synthesis of a new chromophore, 2-(4-nitrophenyl)benzofuran, for two-photon uncaging using near-IR light. *Chem. Commun.* **2016**, *52*, 331–334.

(43) Montalti, M.; Credi, A.; Prodi, L.; Gandolfi, M. T. *Handbook of Photochemistry*, 3rd ed.; CRC Press: Boca Raton, FL, 2006; pp 601–604.

(44) Ellis-Davies, G. C. R.; Barsotti, R. J. Tuning caged calcium: Photolabile analogues of EGTA with improved optical and chelation properties. *Cell Calcium* **2006**, *39*, 75–83.

(45) Agarwal, H. K.; Janicek, R.; Chi, S.-H.; Perry, J. W.; Niggli, E.; Ellis-Davies, G. C. R. Calcium uncaging with visible light. *J. Am. Chem. Soc.* **2016**, *138*, 3687–3693.