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Henry's law constant of N,N-dichloromethylamine.
Application to the contamination of the atmosphere of
indoor swimming pools
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Abstract
The volatility of N,N-dichloromethylamine (DCMA), a disinfection by-product formed during
chlorination of swimming pool water, has been investigated in the present work. The Henry's law
constants for DCMA were experimentally determined at five temperatures (5, 15, 25, 35 and 45 °C)
using the single equilibrium technique. The volumic ratio between the gas and the water phases in
the headspace vessels ranged from 1 to 60 and the initial concentration of DCMA in the aqueous
solutions was \approx 2 mM. The values obtained for the dimensionless Henry's law constant varied from
0.047 at 5 °C to 0.312 at 45°C. The temperature dependence of the Henry's law constants followed
the van't Hoff equation. Trace levels of DCMA (16-70 μ g.m ⁻³) were detected in the atmosphere of
an indoor swimming pool whereas the concentrations of DCMA in water pools were in the range
8.8-15 μg.L ⁻¹ .
KEYWORDS: Henry's law constant, N,N-dichloromethylamine; Temperature dependence;
Headspace technique; Swimming pool.

1. Introduction

25 Chlorination with chlorine gas or sodium hypochlorite is the most common process used for the 26 disinfection of public swimming pool water. Along with its disinfectant action, chlorine also leads 27 to the formation of numerous disinfection by-products such as chloramines, trihalomethanes, 28 haloacetic acids, chloral hydrate and haloacetonitriles by the reaction with pollutants introduced by bathers. It has been estimated that a bather introduces in the water 25-80 mL.h⁻¹ of urine and 200-29 1000 mL.h⁻¹ of sweat (Gunkel and Jessen, 1988; Erdinger et al., 1997). These human materials 30 contain nitrogenous compounds (urea, ammonia, creatinine, amino-acids, amines, ...) and the 31 amount of nitrogenous pollution released by a bather is about equal to 0.85 g N.h⁻¹ (Seux, 1988). 32

33 It is well-known that reactions of aqueous chlorine with ammonia at neutral pH lead to the 34 formation of monochloramine and dichloramine as intermediates and to nitrogen gas as end-product 35 (Jafvert and Valentine, 1992). In addition to N₂, NO₃⁻ is a stable end-product of reactions between 36 chlorine and ammonia. The reactions are largely controlled by the chlorine: ammonia nitrogen ratio 37 (Cl:N), pH, buffer and ionic strength. At Cl:N less than 7.6:1 by weight, chlorine is present as 38 combined chlorine (inorganic chloramines) and at Cl:N higher than 7.6:1, free chlorine is present in 39 water. Small amounts of nitrogen trichloride (or trichloramine) and nitrate ions are also produced. 40 The reactions of chlorine with nitrogenous components present in urine and sweat can also lead to 41 the formation of organic and inorganic chloramines, nitrate as well as many potentially harmful 42 DBPs (Judd and Bullock, 2003). By using a new analytical method (membrane introduction mass 43 spectrometry), Li and Blatchley (2007) have shown that nitrogen trichloride can also be formed by 44 chlorination of four model compounds present in urine (urea, creatinine, histidine and arginine). The 45 reaction of chlorine with creatinine leads to the formation of N₂N-dichloromethylamine (CH₃NCl₂, 46 DCMA) (Tachikawa et al., 2005; Li and Blatchley, 2007) and Li and Blatchley, (2007) first reported 47 the occurrence of CH₃NCl₂ in a swimming pool water.

48 Chloramines and in particular inorganic chloramines have been suggested to largely contribute to 49 the typical smell and to the irritant properties of the atmosphere of indoor swimming pools (Héry *et* 50 *al.*, 1995). Nitrogen trichloride is the most volatile inorganic chloramine (Holzwarth *et al.*, 1984) 51 and is easily released into the air atmosphere of indoor pools. It has been shown to be a powerful 52 irritant for upper respiratory tracts (Gagnaire et al., 1994). Therefore, most of the studies related to 53 the health risks associated with an exposure to DBPs present in the atmosphere of swimming pools 54 have focused on nitrogen trichloride. Published values for nitrogen trichloride concentrations in the atmosphere of swimming pools range from below the detection limit ($\approx 0.05 \text{ mg.m}^{-3}$) to about 2 55 mg.m⁻³ (Jacobs et al., 2007). For swimming pool workers, a dose/response relationship was 56 57 obtained for eye irritation, nasal and throat symptoms (Massin et al., 1998a; Massin et al., 1998b). 58 In France, NCl₃ concentrations in the atmosphere of indoor pools are not regulated. However, the 59 French institute of Occupational Health and Safety (INRS) has proposed a comfort limit value of 0.5 mg.m⁻³ (Héry et al., 1995) even 0.3 mg.m⁻³ (Gérardin et al. 2005) for the concentration of NCl₃ in 60 61 the atmosphere of indoor pools. Carbonelle et al. (2002) have shown that trichloramine in air increases the lung epithelium permeability at concentration levels as low as 0.3 mg.m⁻³. In Belgium, 62 63 the Government of the Brussels-Capital Region has set a recommended maximum value of 0.3 mg.m⁻³ and a maximum admissible value of 0.5 mg.m⁻³ for the concentration of NCl₃ in the 64 65 atmosphere of indoor swimming pool (Gouvernement de la Région de Bruxelles-Capitale, 2002).

Organic chloramines formed by the reaction of chlorine with organic nitrogen contaminants may 66 i) contaminate the atmosphere of swimming pools, ii) give a positive interference in the 67 68 determination of NCl₃ concentration and iii) contribute to the irritant properties of the swimming 69 pool air. Among the organic chloramines, low molecular weight compounds such as DCMA might 70 be volatile. To our knowledge, the Henry's law constant of this chlorination by-product of creatinine 71 is unknown. Therefore, the main objective of the present work was to determine the Henry's law 72 constant of DCMA in the water/air system at various temperatures ranging between 5 and 55 °C. A 73 second objective of this work was to determine the concentration of DCMA in a pool water and in 74 the atmosphere of an indoor swimming pool of Poitiers.

75 **2. Material and methods**

76 2.1. Reagents and preparation of standard solutions of DCMA

All solutions were prepared from reagent-grade chemicals and purified water delivered by a
Millipore system (Milli RX75/Synergy 185). Stock solutions of methylamine (methylamine

hydrochloride, Acros Organics, 99%) and of free chlorine (prepared from sodium hypochlorite,
Acros Organics, 13 %) were stored in the dark at 4 °C.

Stock solutions of N,N-dichloromethylamine (2 mmol.L⁻¹) were prepared a few minutes before use by slowly adding free chlorine (pH \approx 8) to a solution of methylamine chloride in a well-stirred reactor and using a chlorine:methylamine chloride molar ratio of 2:1. pH of the final solution was adjusted to pH 3.5 ± 0.5 with hydrochloric acid (37 % by weight). The change of the UV/Visible absorbance with reaction time indicates that the reaction was achieved within a few seconds at pH < 4, in agreement with the previous works of Poncin *et al.* (1984).

87

2.2. Henry's law constant determination

88 Air/water partition coefficients or Henry's law constants were experimentally determined by using a static method with variable headspace. This method can only be applicable for sufficiently volatile 89 90 compounds to be analytically admissible (McAullife, 1971; Gossett, 1987). In this study, a volume V_L of a freshly prepared aqueous solution of DCMA ($C_{0,L} \approx 2 \text{ mmol.L}^{-1}$) was put into a vial (Total 91 volume : V_T : 44 ± 1 mL) or into a bottle (V_T : 260 ± 2 mL). The total volume of each vial or bottle 92 93 was exactly determined. The stock solution, the vessel and the air in the vessel were thermostated at 94 the desired temperature for at least one hour prior to the introduction of the DCMA solution. The 95 vessel was hermetically sealed with a PTFE-faced-silicone septum, immediately after introduction of the DCMA solution. For each run, various ratios of inner gaseous headspace volume to the liquid 96 volume ($1 < V_G/V_L < 60$) were used in order to show the dependency of $C_{eq,G}$ with $C_{eq,L}$. The vessels 97 98 were set on a rotary tumbler for agitation in a thermostated room (Lovibond) for at least one hour. 99 Preliminary experiments demonstrated that the gas/liquid equilibrium was achieved within 10 100 minutes (Fig. S4 in Supplementary Content). After an agitation time of 1 hour, the vial or the bottle 101 was opened, the liquid phase was immediately transferred in a quartz cell (without using a pipette) 102 for a UV-spectroscopic determination of equilibrium DCMA concentration in the liquid phase (Ceq.L) at 302 nm. For each run, some vials were filled without headspace in order to confirm that 103 DCMA was not decomposed under our experimental conditions. 104

105 **2.3. Determination of DCMA level in the atmosphere of an indoor swimming pool**

106 An in situ measurement method was developed to determine DCMA level in the atmosphere of an indoor swimming pool located at Poitiers (France). The sampler consisted of two 250-mL pyrex® 107 glass traps, containing 200 mL sodium sulfite (10 mmol.L⁻¹) and phosphate buffer (10 mmol.L⁻¹, pH 108 = 3.5-4) (Fig. S1 in Supplementary Content). The sampling flow rate was 1.5 L.min^{-1} and the 109 sampling period varied from 2 to 4 h. The two traps were connected in series and no significant 110 breakthrough of DCMA from the first trap to the second one has been observed during any 111 sampling. Under these conditions, DCMA present in the atmosphere of the swimming pool is 112 113 absorbed in the traps and instantaneously reduced into the non volatile methylammonium ion (pKa = 10.62; Hall, 1957) by sodium sulfite: 114

115
$$CH_3NCl_2 + 2 SO_3^{2-} + H_3O^+ + H_2O \rightarrow CH_3NH_3^+ + 2 SO_4^{2-} + 2 HCl_3NCl_2 + 2 HCl_3NCl_3 + 2 HCl_3NCl_3 + 2 HCl_3NCl_2 + 2 HCl_3NCl_3 + 4 HCl$$

116 It should be noted that other volatile chloramines such as trichloramine can also be reduced by 117 sodium sulfite in the traps.

In order to determine the amounts of DCMA which have been trapped, the trapping solution was transferred into a 200 mL volumetric flask and completed to 200 mL with purified water (Fig. S1 in Supplementary Content). A volume of 20 mL of the trapping solution was introduced into a 25 mL volumetric flask (five replicates for each trap). Methylammonium ion present in the trapping solution was converted into DCMA with sodium hypochlorite in a 10 % molar excess (calculated from initial sulfite concentration) and the solution was then adjusted to pH 3.5 ± 0.5 . DCMA concentration was then determined by HPLC and UV detection at 210 nm.

125

126 **2.4. Determination of DCMA in pool water**

A volume of 10.0 L of water was collected from a spa and from a basin located in the indoor swimming pool of Poitiers. DCMA was then stripped with an air flow rate of 1.5 L.min⁻¹ during 10 h at 45°C and the desorbed DCMA was then reduced into methylammonium ion with sodium sulfite by passing the gas through the two same traps than those used for air sampling. It should also be 131 noted that a slow oxidation of sodium sulfite by oxygen occurred in the traps but the concentration 132 of sodium sulfite was always greater than 2 mmol. L^{-1} after 10 h of air sparging.

To assess this method, 10.0 L of a standard aqueous solution of DCMA ([DCMA] = $1.9 \mu \text{mol.L}^{-1}$) has been analyzed under identical conditions. The data showed that DCMA was quantitatively desorbed and trapped in the first absorption vessel. The recovery yield was 104 ± 3 % and DCMA was not detected in the second trap.

137 **2.5. Analytical methods**

Free chlorine and DCMA concentrations in the stock solutions were determined iodometrically by using the US Standard Method 4500B (APHA, 2005) with sodium thiosulfate 0.01 mol.L⁻¹ (Prolabo, > 99.9%). In the case of DCMA, two moles of free iodine is liberated by the reaction with KI :

142
$$CH_3Cl_2 + 4 I^- + 4 H^+ \rightarrow CH_3NH_2 + 2 HCl + 2 I_2$$

pH was measured with a MeterLab PHM 250 pH-meter (Radiometer) equipped with a glass
electrode (Radiometer, CL114 JB).

145 UV spectra of DCMA were recorded from 200 to 350 nm with a Cary 50 (Varian) and treated 146 with WinUV 3.0 software (Varian). DCMA concentrations in dilute solutions (0.35-3.5 mmol.L⁻¹) 147 were determined iodometrically in order to confirm the concentrations of DCMA.

148 DCMA concentration in the traps was determined by HPLC because of the possible presence of 149 other chloramines. HPLC analyses were performed with a Waters system (W600 controller; 150 W717plus Autosampler) equipped with a Zorbax 300Extend-C18 column and a UV detector at 210 nm (Waters W486). DCMA was eluted with a methanol (20%) / phosphate buffer (0.1 mol.L⁻¹; pH 151 = 4) (80%) mobile phase at a flow rate of 0.8 mL.min⁻¹. All HPLC analyses were made on five 152 replicates with a sampling volume of 200 µL. Under these analytical conditions, the absolute 153 detection and quantification limits were evaluated as 3 x 10^{-12} and 9 x 10^{-12} mol, respectively. 154 Relative quantification limit depends on the concentration factor applied during sampling collection 155 156 and on the volume of reactants added before analyses.

157

3. Background and expressions used

158 **3.1. Data format**

The two-phase equilibrium between air and water phases for a volatile solute in a dilute aqueous solution can be represented by Henry's law coefficients or by air/water partition coefficients. Since these coefficients can be expressed by a multitude of forms and associated units, Table 1 reports the symbols and the expressions that will be used in this study.

163 **3.2. Calculations of Henry law coefficients for DCMA**

164 If DCMA is not decomposed under our experimental conditions, equilibrium concentration in the 165 gas phase ($C_{i,G}$) can be deduced from the mass balance expression :

166
$$C_{i,G} = \frac{V_L}{V_G} (C_{0,L} - C_{i,L})$$
 (1)

167 The molar fractions of DCMA in water and in air were calculated as follows:

168
$$x_{i} = -\frac{C_{i,L}}{C_{i,L} + \rho_{L}/MW_{L}} = \frac{C_{i,L}}{\rho_{L}/MW_{L}} \text{ for dilute aqueous solutions}$$
(2)

169
$$y_i = -\frac{C_{i,G}}{P_T/RT} = \frac{RTV_L}{P_TV_G} (C_{0,L} - C_{i,L})$$
 (3)

170 where ρ_L is approximately equal to 1000 kg.m⁻³ for the temperature range studied in this work, 171 MW_L is the molecular weight of water (0.018 kg.mol⁻¹), R is the universal gas constant (R = 8.314 172 Pa.m³.mol⁻¹ K⁻¹ = 8.206 x 10⁻⁵ atm.m³.mol⁻¹.K⁻¹).

The various Henry's law constants listed in Table 1 were then calculated from $C_{i,L}$, $C_{i,G}$, x_i and y_i values. Assuming that $P_T = 1$ atm, the relationship between the various Henry's law constants (Staudinger and Roberts, 2001) are the following:

176
$$H_{cc} = H_{yx} \frac{0.2194}{T} = H_{px} \frac{2.165 \times 10^{-6}}{T} = H'_{px} \frac{0.2194}{T} = H_{pc} \frac{0.120}{T} = H'_{pc} \frac{12187}{T}$$
(4)

177 **4. Results**

178 4.1. Purity, stability and UV spectrum of aqueous solutions of DCMA at pH 3.5 ± 0.5

The HPLC chromatograms of stock solutions of DCMA prepared at pH 3.5 ± 0.5 with a chlorine:methylamine molar ratio of 2:1 showed only one peak corresponding to DCMA (Fig. S2 in Supplementary Content). Iodometric titrations showed no loss of oxidants because the total amount of chlorine was equal to the amount of free chlorine which has been introduced into the solutions of methylamine. These data indicate that stock solutions of DCMA only contain DCMA and that the concentration of DCMA is equal to the half of the initial concentration of free chlorine. (1 mole of DCMA is equivalent to 2 moles of free chlorine).

UV/Visible spectra of dilute solutions of DCMA (3.4 x 10^{-4} M < [DCMA] < 3.5 x 10^{-3} M) were 186 recorded by using quartz cells (optical length: 1, 5 or 10 cm). As shown the spectrum presented in 187 188 Fig. S3 in Supplementary Content, DCMA absorbs in the UV region. The spectrum presents an absorption band at 302 nm with a molar extinction coefficient equal to $\varepsilon_{302nm} = 309 \pm 18 \text{ mol}^{-1}$ 189 ¹.L.cm⁻¹ (mean value of 39 absorbance values determined from different standard solutions of 190 191 DCMA). Spectrophotometric analyses also demonstrated that the UV spectra of DCMA solutions 192 stored in hermetically closed vials and filled without headspace did not change over a storage time 193 of 15 h at 5 - 55 °C and pH 3.5 ± 0.5 . These results confirm that DCMA did not decompose in water 194 under our experimental conditions.

Fig. S3 in Supplementary Content also shows that the UV/visible spectrum of DCMA is similar to the spectrum of dichloramine (NHCl₂) which presents an absorption band at 293 nm ($\varepsilon_{293nm} = 289 \pm$ 5 mol⁻¹.L.cm⁻¹, mean value of 9 UV-absorbance values).

198 4.2. Henry's law constant of N,N-dichloromethylamine

The Henry's law constant was determined at five different temperatures by measuring the equilibrium concentration of DCMA in the aqueous phase ($C_{i,L}$). All the experimental conditions ($C_{0,L}$, V_L , V_G , T) and the equilibrium concentrations ($C_{i,L}$ and $C_{i,G}$) have been reported in Supplementary Content (Tables S1-S5 and Fig. S5).

From the experimental data, the various forms of the Henry's law constants reported in Table 1 (H_{cc} , H'_{px} , H'_{pc} and H_{yx}) have been calculated by linear regression analysis. As shown the data in Fig. 1, straight lines were obtained for the dependence of $Ci_{,G}$ with $C_{i,L}$ at 288 and 308 K as well as for the other temperatures studied (Tables S1-S5 in Supplementary Content).

Values of Henry's law constants determined in this work are reported in Table 2. The values obtained for the air-to-water concentration ratio (H_{cc}) vary from 0.048 at 5 °C to 0.335 at 45 °C. The temperature dependence of Henry's law constants is correctly described by the van't Hoff equation (Fig. 2):

$$\ln(H) = A + \frac{B}{T}$$

where B = 4298.11 K (excepted for H_{cc}) and A, a constant which depends on the unit used for the Henry's law constants. Values of A and B have been reported in Table 2.

214 The solvatation enthalpy (ΔH°_{sol}) can be calculated from the slope of the ln(H) = f(1/T) straight 215 line (for H constant other than H_{cc}):

216
$$\Delta H^{\circ}_{sol} = B R \text{ with } R = 8.314 \text{ Pa.m}^3 \text{.mol}^{-1} \text{ K}^{-1}$$

The solvatation enthalpy of $-35.7 \text{ kJ.mol}^{-1}$ obtained for DCMA is very close to the values published for NH₂Cl, NHCl₂ and NCl₃ (Sander, 1999) which are in the range 34.2-39.9 kJ.mol⁻¹ (Table 3).

220 A comparison of the Henry's law constants (Table 3) or of the air-water partition coefficient (Fig. 221 3) of some disinfection by-products formed during chlorination of swimming pool water shows that DCMA is a little less volatile than chloroform, about 3 times less volatile than trichloramine but far 222 223 much volatile than monochloramine and dichloramine. At the average temperature of recreation pools ($\approx 30^{\circ}$ C), the air-water coefficients (H_{cc} values) of trichloramine, chloroform, DCMA, 224 dichloramine and monochloramine are equal to 0.504, 0.195, 0.135, 1.75 x 10^{-3} and 5.58 x 10^{-4} 225 respectively. If DCMA is produced in water pools at concentration levels of about 0.1 μ mol.L⁻¹ (10 226 μ g.L⁻¹), the theoretical concentration of DCMA in air in a closed system should be ≈ 1.9 mg.m⁻³. 227 This concentration might cause irritation to bathers and to pool attendance. 228

4.3. Concentration levels of DCMA in water pools and in the atmosphere of an indoorswimming pool.

The DCMA levels in water and in the atmosphere of an indoor swimming pool located at Poitiers have been determined between February 2009 and May 2009. The swimming pool has a 25 meter pool (436 m³), a leisure pool (190 m³), a children pool (10 m³), a toboggan pool (26 m³) and a spa (5 m³). The total volume of the atmosphere of the indoor pool (15 000 m³) can be contaminated by volatile compounds that escape from the five basins. Table 4 reports the main operating conditions of the swimming pool for the four campaigns (C1 to C4) made in this study.

237 DCMA was determined in water collected from the spa (campaigns C1 and C2) and from the 25 meter pool (campaigns C3 and C4). The spa water is totally renewed every night in order to 238 maintain the concentration of combined chlorine below 0.6 mg Cl₂.L⁻¹. The mean residence time of 239 water in the 25 meter pool is 10 days. Analyses showed that the concentrations of DCMA in water 240 samples ranged from 88 to 150 nmol.L⁻¹ (8.8 and 15 μ g.L⁻¹) (Table 4) and that about 3 % of the 241 242 combined chlorine concentration can be accounted for DCMA (Table 4). Our concentrations of DCMA in water are of the same order of magnitude than those obtained by Weaver et al. (2009) 243 using membrane introduction mass spectrometry (MIMS) for the determination of DCMA in 11 244 water pools (1.6 μ g.L⁻¹ < [DCMA] < 50.5 μ g.L⁻¹). It should be noted that about 80 % of the spa 245 water is renewed every night and there is always about 12-13 bathers in the spa. 246

247 The water samples collected from the 25 meter pool were chlorinated at an arbitrary dose of 14 mg Cl₂.L⁻¹ in a 10 L headspace-free bottle in order to show if the DCMA concentration could 248 249 increase by increasing free chlorine concentration and reaction time. After a reaction time of 72 h, analyses showed that this postchlorination step led to a chlorine consumption of about 8 mg Cl₂.L⁻¹ 250 251 and to an increase of the DCMA concentration by a factor of 2.5-3. The DCMA concentrations were in the range 230-270 nmol.L⁻¹ (23-27 μ g.L⁻¹). This increase of the concentration of DCMA suggests 252 253 that reactions leading to the formation of DCMA are not very fast, in agreement with the kinetic 254 data obtained by Li and Blatchley (2007) for the formation of DCMA from creatinine. Methylamine 255 and creatinine are known to produce DCMA by chlorination (Poncin et al., 1984; Li and Blatchley, 256 2007). As the concentration of methylamine in urine is about 10 times less than the concentration of 257 creatinine (Mitchell and Zhang, 2001) most of the DCMA formed in swimming pool is probably

formed from creatinine. On the basis of a production of 0.66 mole of DCMA per mole of creatinine (Li and Blatchley; 2007), the DCMA formation potentials measured in our study ($250 \pm 20 \text{ nmol.L}^{-1}$) ¹) should correspond to a concentration of creatinine in the water samples of 350-400 nmol.L⁻¹. This is fully consistent with the concentrations of creatinine which can be estimated from the concentrations of urea of 30-50 µmol.L⁻¹ (1.8-3.0 mg.L⁻¹) which have been measured in this swimming pool water (unpublished data) and from a mean molar ratio of urea to creatinine of about 100 in urine.

As far as air analyses are concerned, air was sampled at a distance of about 4 m from the two 265 main basins and at a height of 1.5 m. DCMA concentrations in air were in the range 0.16-0.70 266 umol.m⁻³ (16-70 µg.m⁻³) and far below the theoretical DCMA concentration in air (1.5-2.0 mg 267 DCMA m⁻³) that should be in equilibrium with the aqueous concentrations of DCMA at 30 °C (11-268 15 µg.L⁻¹) at 30 °C (Table 4). The concentration of DCMA was higher for the first campaign (70 \pm 269 30 μ g.m⁻³) than for the second one (16 ± 2 μ g.m⁻³). This was consistent with the observations made 270 by the attendant pools who reported that the atmosphere was found to be more irritant during the 271 first campaign of analyses. It should also be noted that the lowest value of DCMA concentration 272 obtained during the second campaign might be attributed to the fact that the ventilation rate was 273 higher and that all the roof windows were opened. 274

Nowadays, the concentrations of DCMA in water and in the atmosphere of indoor swimming 275 pools are not controlled. As trichloramine exposure is considered to be at the origin of eye and 276 277 airway irritations, only trichloramine concentrations have been determined in the atmosphere of indoor pools (Massin et al., 1998b; Jacobs et al., 2007). In these studies, the analytical method used 278 279 for the determination of trichloramine in the atmosphere of indoor pools is based on the reduction of 280 trichloramine by arsenic trioxide and subsequent determination of the chloride ion formed (Héry et al., 1995). The concentration of trichloramine in water pools can also be determined by this method 281 after a gas/liquid desorption of trichloramine. The data obtained in the present work suggest that the 282 use of this method might lead to an overestimation of the concentration of trichloramine in the 283 284 presence of other volatile organic chloramines such as DCMA. The membrane introduction mass

spectrometry (MIMS) is a more convenient method for the differentiation and the quantification of inorganic and organic chloramines in water and in air (Weaver *et al.*, 2009).

287 **6.** Conclusion

The Henry's law constants determined in the present work show that N,N-dichloromethylamine 288 (DCMA) is guite as volatile than trichloramine. This work confirms the presence of DCMA in pool 289 290 water previously reported by Li and Blatchley (2007) and Weaver et al. (2009) and this work 291 demonstrates for the first time the presence of DCMA in the atmosphere of indoor swimming pools. 292 These data suggest that the health risks associated with the formation of this volatile organic 293 chloramine during chlorination of swimming pool water must be considered in further 294 epidemiological studies. Additional research is also needed in order to develop specific analytical 295 methods for the determination of DCMA and trichloramine and to evaluate the irritant properties for 296 eyes and the respiratory tracts of DCMA and for mixture of DCMA and trichloramine.

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300 Appendix A : Supplementary Content

301 Supplementary data associated with this article can be found in the online version at DOI ...

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