

Overview of cryo-EM image processing procedure.

Maximum likelihood classification scheme and masks used to obtain the complexes of eIF6, SBDS and EFL1

bound to the 60S ribosomal subunit. Insets show the position of the spherical masks (grey) on the 60S subunit

(cyan).



Resolution of the 60S ribosomal subunit complexes and validation of the atomic model.

(a) Gold-standard Fourier shell correlation (FSC) curves for the three 60S ribosome complexes after 3D refinement and statistical movie processing in RELION (Bai, X. C. *et al.*, *Elife* 2, e00461, 2013; Scheres, S. H. *J Struct Biol* 180, 519-530, 2012).

(b) Surface (top panel) and cross-sectional (bottom panel) views of the unsharpened final maps colored according to local resolution calculated with the ResMap software package (Kucukelbir, A. *et al.*, *Nat Methods*, **11**, 63-65, 2014).

(c, d, e, f) Examples of densities with the model fitted showing helix 72 of the 26S rRNA from the 60S-eIF6-SBDS complex with the density filtered at 3.3 Å (c), α -helix of the uL6 protein from the 60S-eIF6-SBDS complex with the density filtered at 3.3 Å (d), SBDS from the 60S-eIF6-SBDS complex with the density is filtered at 4 Å (e), EFL1 from the 60S-eIF6-SBDS-EFL1 complex with the density is filtered at 8 Å (f).

(g, h, i) Cross-validation against over-fitting. FSC curves are shown for the 60S-eIF6-SBDS (g), 60S-eIF6-SBDS-EFL1 (h) and 60S-SBDS-EFL1 (i) complexes between the final refined atomic model and the reconstructions from all particles (black); between the model refined in the reconstruction from only half of the particles and the reconstruction from that same half (FSC_{work}, red); and between that same model and the reconstruction from the other half of the particles (FSC_{test}, green).



Atomic models of the 60S-SBDS-eIF6 complex.

(a) Secondary structure diagram of modeled *Dictyostelium* 26S rRNA. The diagram was modified from *S. cerevisiae* 25S rRNA (Comparative RNA Web Site, www.rna.ccbb.utexas.edu). The PTC is shown in bold. Base pairs involved in tertiary interactions are indicated with lines connecting the circles or boxes around the bases involved in the interaction. (-) corresponds to canonical base pairs, while (•, \circ and •) correspond to non-canonical base pairs.

(b, c) Back (b) and front (c) views of the atomic model of the 60S-eIF6-SBDS complex. SRL, sarcin-ricin loop.



Molecular environment of eIF6, SBDS and EFL1.

(**a**, **b**) Schematic representations of *Homo sapiens* EF-2 (**a**) and EFL1 (**b**).

(c) Ribbon representation of the atomic model of human EFL1 with the subdomain structure highlighted using

the same color scheme as (**b**).

(d-g) Back (d, f) and front (e, g) views of the atomic models of the 60S-eIF6-SBDS-EFL1 and 60S-SBDS-

EFL1 complexes, respectively. SRL, sarcin-ricin loop.



uL16 is required to recruit Sdo1 to the 60S subunit *in vivo*.

(a) The EFL1 domain II insertion is dispensable *in vivo*. Random sporulation assay showing *efl1* Δ cells transformed with empty vector (pRS316) control or plasmids expressing wild type *EFL1* or the *EFL1* Δ 420-580 mutant (left).

(b) Tenfold serial dilutions (left to right) of the indicated yeast cells spotted onto selective –URA medium (left). *efl1* Δ cells transformed with empty vector (pRS316) are non-viable and are therefore not shown.

(c) S174 and S175 are dispensable for Tif6 function *in vivo*. Random sporulation assay showing *tif6* Δ cells transformed with empty vector (pRS316) control or plasmids expressing wild type *TIF6* or the indicated *TIF6* mutants.

(d) Tenfold serial dilutions (left to right) of the indicated yeast strains spotted onto selective –URA medium (left). *tif6* Δ cells transformed with empty vector (pRS316) are non-viable and are therefore not shown.

(e) Severe fitness defect of the *uL16-H123P* allele. Indicated yeast cells with (galactose) or without (glucose) endogenous uL16 spotted in tenfold serial dilutions (left to right) onto solid media.

(f) The *uL16-H123P* allele markedly reduces uL16 expression. uL16 and uL14 (control) were visualized by immunoblotting of extracts from the indicated strains with (galactose) or without (glucose) endogenous uL16. As controls, uL16 and uL14 were visualized by immunoblotting of extracts from strain C375 (wild type).

(g) The T-ALL associated alleles *uL16-H123P*, *uL16-R98S* and *uL16-R98C* impair Sdo1 binding to the 60S subunit *in vivo*. Sdo1-FLAG or uL14 were visualized by immunoblotting of supernatant (S) or pellet (P) from cell extracts with (galactose) or without (glucose) endogenous uL16 across the indicated range of NaCl concentrations.