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# Degradation of enoxacin antibiotic by the electro-Fenton process: Optimization, biodegradability improvement and degradation mechanism

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## Highlights

• The efficiency of the electro-Fenton process on the removal of enoxacin was investigated. • A total degradation of the target molecule was observed. • The biodegradability increased from 0 to 0.5 for the BOD<sub>5</sub> on COD ratio. • By-products were identified by UPLC-MS/MS and a degradation pathway was proposed.

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## Abstract

This study aims to investigate the effectiveness of the electro-Fenton process on the removal of a second generation of fluoroquinolone, enoxacin. The electrochemical reactor involved a carbon-felt cathode and a platinum anode. The influence of some experimental parameters, namely the initial enoxacin concentration, the applied current intensity and the Fe(II) amount, was examined. The degradation of the target molecule was accompanied by an increase of the biodegradability, assessed from the BOD<sub>5</sub> on COD ratio, which increased from 0 before treatment until 0.5 after 180 min of electrolysis at 50 mg L<sup>-1</sup> initial enoxacin concentration, 0.2 mmol L<sup>-1</sup> Fe(II) concentration and 300 mA applied current intensity. TOC and COD time-courses were also evaluated during electrolysis and reached maximum residual yields of 54% and 43% after 120 min of treatment, respectively. Moreover, a simultaneous generation of inorganic ions (fluorides, ammonium and nitrates) were observed and 3 short chain carboxylic acids (formic, acetic and oxalic acids) were identified and monitored during 180 min of electrolysis. By-products were identified according to UPLC-MS/MS results and a degradation pathway was proposed.

## Keywords

- Electro-Fenton process;
  - Enoxacin removal;
  - Hydroxyl radicals;
  - Biodegradability improvement;
  - By-products
- 

## 1. Introduction

In recent years, treating environmental hazards due to the significant detection of pharmaceutical compounds in wastewater became an area of interest for the scientific community worldwide (Ellis, 2006, Sui et al., 2011 and Yoon et al., 2010). Excretion after administration and wastes in the production sites are the main causes of the presence of pharmaceutical compounds in the aquatic environment (Bound and Voulvoulis, 2005). This type of compounds has been recently classified as a priority risk pollutants (Hernando et al., 2006 and Watkinson et al., 2009). Most of them are antibiotics. Their presence in wastewater is responsible for the development of antimicrobial resistance among microorganisms (Levy, 1998). Owing to their recalcitrance, the commonly employed treatment processes (biological processes) appear not effective enough for a complete removal of such species. The contamination by these micropollutants is observed not only in the received water but reached also surface waters. Their toxic and adverse effects were proven in previous studies (Daughton and Ternes, 1999 and Giger et al., 2003).

Fluoroquinolones (FQs) are the third largest group of antibiotics with 17% of the global market share (Van Doorslaer et al., 2014). They are frequently detected with relatively high concentrations in hospital wastewaters ( $3 \text{ ng L}^{-1}$ – $240 \text{ } \mu\text{g L}^{-1}$ ), urban wastewaters ( $2 \text{ ng L}^{-1}$ – $14 \text{ } \mu\text{g L}^{-1}$ ) and in bulk drug producer wastewaters ( $6 \text{ ng L}^{-1}$ – $31 \text{ mg L}^{-1}$ ) (Golet et al., 2002, Lin et al., 2008 and Van Doorslaer et al., 2014). FQs are also observed in livestock and aquaculture wastewaters (Van Doorslaer et al., 2014). Their broad antimicrobial spectrum may lead to the proliferation of bacterial drug resistance (Watkinson et al., 2007). FQs are characterized by their chemical stability and their high resistance to hydrolysis and to high temperatures, due to their quinolone ring. Conventional water treatment plants show a high variation in FQ removal efficiency (between 47 and 77% on average) (Van Doorslaer et al., 2014). However, it is suggested by several authors that the predominant removal mechanism for FQ compounds is sorption to activated sludge rather than biodegradation (Hendricks and Pool, 2012, Lindberg et al., 2006 and Van Doorslaer et al., 2014). A previous study reported that more than 80% of the total amount of ciprofloxacin and norfloxacin (FQ molecules) treated in a sewage treatment plant in Sweden was ultimately found in the digested sludge (Lindberg et al., 2006). This behavior is due to the high affinity of FQs for sludge, soils and sediments, through electrostatic and hydrophobic interactions (Lindberg et al., 2006).

Therefore, an effective strategy must be found to degrade these compounds from aquatic matrices and to enhance their biodegradability as a pretreatment before their discharge into sewage treatment plants.

Advanced Oxidation Processes (AOP) have received great interest in recent years as an alternative method to treat organic and recalcitrant molecules (Nasuhoglu et al., 2012). Some recent review papers reported the relevance of AOP on the degradation and the mineralization of residual molecules (Feng et al., 2013, Klavarioti et al., 2009, Pera-Titus et al., 2004 and Sirés et al., 2014). These oxidative treatment processes can be classified into four categories: chemical oxidation processes in homogeneous phase ( $\text{H}_2\text{O}_2/\text{Fe}(\text{II})$  and  $\text{H}_2\text{O}_2/\text{O}_3$ ); photocatalytic processes in homogeneous phase and/or heterogeneous phase ( $\text{H}_2\text{O}_2/\text{UV}$ ,  $\text{O}_3/\text{UV}$ ,  $\text{Fe}(\text{II})/\text{H}_2\text{O}_2/\text{UV}$  and  $\text{TiO}_2/\text{UV}$ ); oxidation by sonochemical processes and electrochemical oxidation processes. AOP are characterized by the in-situ production of hydroxyl radicals ( $\text{OH}$ ) that are very strong oxidants ( $E^\circ (\text{OH}/\text{H}_2\text{O}) = 2.81 \text{ V/SHE}$  at  $25^\circ\text{C}$ ) able to oxidize complex organic molecules such as pesticides (Guivarch et al., 2003), industrial dyes (Hammami et al., 2008) and pharmaceuticals (An et al., 2010, Guinea et al., 2010, Ikehata et al., 2006, Mansour et al., 2012 and Rodrigues-Silva et al., 2013).

The electro-Fenton Process (EF) is one of the most efficient electrochemical AOP, which consists in the continuous production of hydroxyl radicals using electrogenerated Fenton reagent ( $\text{H}_2\text{O}_2/\text{Fe}(\text{II})$ ) according to the reactions below (El-Desoky et al., 2010 and Xie and Li, 2006):



EF achieves high reaction yields and may be considered as an environmentally clean process (El-Desoky et al., 2010).

Studies dealing with FQ degradation using AOP are numerous. Flumequine degradation was investigated by EF and by photo-electro-Fenton (PEF) in a non divided electrochemical cell using a Carbon-PTFE-air diffusion cathode and a Boron Doped Diamond (BDD) anode. Results showed an almost total mineralization in 360 min at 300 mA current intensity for both EF and PEF (Garcia-Segura et al., 2012). Ciprofloxacin was completely degraded by EF within 8 min and reached 94% of mineralization after 360 min of treatment using a carbon felt cathode and Platinum (Pt) anode (Yahya et al., 2014). Its degradation was also studied by sonochemistry (Xiao et al., 2014). Anodic oxidation and Solar-photo-electro-Fenton have been tested for Enrofloxacin elimination in an undivided filter press coupled to a photoreactor and using a batch recirculation flow plant. Investigations reported 28% and 86% of mineralization yields respectively after 300 min of treatment with a BDD anode and a Carbon PTFE gas diffusion cathode (Guinea et al., 2010). Nasuhoglu et al. demonstrated the efficiency of ozonation and  $\text{TiO}_2$  photocatalysis to degrade Levofloxacin leading to maximal oxidation yields of 59% and 70% respectively after 300 min of treatment (Nasuhoglu et al., 2012). The efficiency of photocatalysis was also verified for moxifloxacin (Van Doorslaer et al., 2015).

However, there is a lack of studies dealing with the effectiveness of coupling AOP with a biological treatment for FQs. The effectiveness of this type of coupling has been proven with other classes of antibiotics such as sulfonamides (sulfamethazine) (Mansour et al., 2012), macrolides (tylosin) (Yahiat et al., 2011) and tetracycline (Ferrag-Siagh et al., 2013).

Therefore, the focus of this study is to evaluate the applicability of EF for the pretreatment of enoxacin (ENO), an oral broad-spectrum FQ antibacterial agent, used in the treatment of respiratory and urinary tract infections (Patel and Spencer, 1996) and to assess the possibility of coupling EF and a biological process for enoxacin mineralization. In this sense, the biodegradability of the degraded solution was examined. By-products and inorganic species were analyzed as well.

## 2. Materials and methods

### 2.1. Chemicals

Enoxacin (1-ethyl-6-fluoro-4-oxo-7-(piperazin-1-yl)-1,4-dihydro-1,8-naphthyridine-3-carboxylic acid) was generously provided by the LNCM (Laboratoire National de Contrôle des Médicaments – Tunis, Tunisia) (structure displayed in Fig. 1).

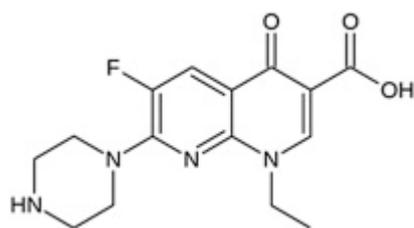


Fig. 1.

Molecular structure of enoxacin.

$\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$  (purity 99%) and  $\text{Na}_2\text{SO}_4$  (purity 99%) were used as catalyst and a supporting electrolyte respectively and were purchased from Acros Organics (Thermo Fisher Scientific, Illkirch, France). Acetonitrile (purity 99.9%) (HPLC grade) was obtained from Sigma–Aldrich (Saint Quentin Fallavier, France). The initial pH of the solutions was adjusted to 3, the optimal pH value for the electro-Fenton process (Diagne et al., 2007), by adding analytical grade sulfuric acid  $\text{H}_2\text{SO}_4$  purchased from Acros Organics. All solutions were prepared with ultrapure water and all the other chemicals used for analysis were supplied from Acros Organics and Sigma Aldrich.

### 2.2. Electrochemical apparatus and procedures

Experiments were performed at room temperature (18 °C) with an undivided cylindrical glass cell containing 250 mL of solution. The cell was equipped with two electrodes, a tri-dimensional carbon felt piece (Carbone Lorraine RVG 4000 Mersen, Paris La Défense, France) as cathode (90 mm × 50 mm × 12 mm) and a cylindrical platinum (Pt) electrode (34.6 cm<sup>2</sup>) as anode, located in the center of the electrochemical reactor to have a good

potential distribution. Compressed air was bubbled for 10 min through the cell to saturate the solution prior to electrolysis. The treated solutions contained a catalytic quantity of  $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$  and their pH was adjusted to 3. Electrolyses were carried out under current controlled conditions. The current was kept at the desired level with a DC power supply (Metrix, model AX 322–Chauvin Arnoux Group, Paris, France) operating in galvanostatic mode to control the current intensity. The ionic strength was maintained constant with the addition of  $50 \text{ mmol L}^{-1} \text{ Na}_2\text{SO}_4$ .

## **2.3. Analytical procedure**

### **2.3.1. Electrochemical analysis**

Electrochemical analysis of ENO was performed using a conventional three-electrodes cell with a vitreous carbon disk electrode ( $3.14 \text{ mm}^2$ ) as the working electrode and a Pt wire as the counter electrode. All the electrode potentials were measured with respect to an Ag/AgCl electrode located near to the working electrode. Experiments were performed at ambient temperature under nitrogen atmosphere to avoid dissolved oxygen. Voltammograms were obtained with a scan rate of  $20 \text{ mV s}^{-1}$  using a BioLogic SP-150 potentiostat recorded to EC-Lab V10.36 software.

### **2.3.2. High performance liquid chromatography (HPLC)**

The evolution of ENO concentrations was monitored by HPLC using a Waters 996 system equipped with Waters 996 PDA (Photodiode Array Detector) and Waters 600LCD Pump. The separation was achieved on a Waters C-18, ( $5 \mu\text{m}$ ;  $4.6 \times 250 \text{ mm}$ ) reversed-phase. Analyses were carried out isocratically using a mixture of acetonitrile/MilliQ water (10/90, v/v) with 0.1% formic acid as eluent, at a flow rate of  $1 \text{ mL min}^{-1}$ . Detection of ENO was performed at 270 nm and the retention time was approximately 12 min.

### **2.3.3. Total organic carbon (TOC) and total nitrogen (TN) measurements**

TOC and TN were measured by means of a TOC– $\text{V}_{\text{CPH/CPG}}$  Total Organic Analyzer Shimadzu. Organic Carbon compounds were combusted and converted to  $\text{CO}_2$  which was detected by a non-dispersive Infra-Red Detector (NDIR). Dissolved Nitrogen compounds were combusted and converted to NO which was then mixed with Ozone chemiluminescence for detection by a photomultiplier.

### **2.3.4. Chemical oxygen demand (COD) measurements**

COD was measured by means of a Test Nanocolor<sup>®</sup> CSB 40 and 160 from Macherey–Nagel (Düren, Germany) (Mansour et al., 2012).

### **2.3.5. Biochemical oxygen demand (BOD<sub>5</sub>) measurements**

BOD<sub>5</sub> measurements were carried out in Oxitop IS6 (from WTW) in order to evaluate the biodegradability for ENO molecule and its by-products. Inoculation of samples, blank solution and control solution was performed according to the following procedure: 100 g of activated sludge (collected from a local wastewater treatment plant) was mixed in distilled water; after agitation the solution was left stand for 10 min; and 20 mL of supernatant was then harvested and mixed to distilled water to obtain a total volume of 2 L. The following

mineral basis was used for all experiments ( $\text{g L}^{-1}$ ):  $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ , 22.5;  $\text{CaCl}_2$ , 27.5;  $\text{FeCl}_3$ , 0.15;  $\text{NH}_4\text{Cl}$ , 2.0;  $\text{Na}_2\text{HPO}_4$ , 6.80;  $\text{KH}_2\text{PO}_4$ , 2.80. The  $\text{BOD}_5$  value was initially estimated based on the COD value experimentally measured or calculated,  $\text{BOD}_5 = \text{COD}/1.46$ .

Neutral pH ( $7.0 \pm 0.2$ ) was adjusted for all the samples by adding NaOH (Chebli et al., 2010).

### **2.3.6. Inorganic ions quantification**

The monitoring of ammonium species ( $\text{NH}_4^+$ ) was realized via the spectrophotometric method of Nessler at 420 nm (Rodier et al., 1996). Fluorides were quantified according to the standard additions method, using a specific fluoride electrode (Yam et al., 1995). Nitrate and Nitrite ions monitoring was carried out by a spectrophotometric method at 415 nm and 543 nm, respectively (Rodier et al., 1996).

### **2.3.7. Liquid chromatography-mass spectrometry (UPLC–MS/MS)**

The devices used are detailed in a previous work (Assassi et al., 2011). The analytes were separated by a Waters Acquity UPLC system (Waters Corporation, Milford, MA, USA) consisting of an Acquity UPLC binary solvent manager, an Acquity UPLC sample manager and an Acquity UPLC column heater equipped with a Waters Acquity UPLC BEH Shield RP18 column ( $2.1 \text{ mm} \times 100 \text{ mm}$ ,  $1.7 \mu\text{m}$  particle size) (Milford, MA, USA) maintained at  $45 \text{ }^\circ\text{C}$ . Analyses were performed using 0.1% formic acid in acetonitrile as eluent A and 0.1% formic acid in a mixture of MilliQ Water/Acetonitrile (90/10, v/v) as eluent B, delivered at a flow rate of  $0.4 \text{ mL min}^{-1}$ .

The elution gradient started with 0% of eluent A and increased linearly to 100% of A within 5 min.

The separated compounds were detected with a Waters Micromass Quattro Premier (Waters Corporation, Manchester, UK) triple quadrupole mass spectrometer. It was operating with an electrospray source in positive ionization mode with a cone potential of 30 V. The ionization source conditions were: capillary voltage of 3.0 kV and source temperature of  $350 \text{ }^\circ\text{C}$ . The cone and desolvation gas flows were  $50 \text{ L h}^{-1}$  and  $750 \text{ L h}^{-1}$ , respectively. They were obtained from an in-house nitrogen source, the Peak Scientific NM30LA (Peak Scientific, Inchinnan, United Kingdom). High-purity argon (99.99%, Air Liquid, Paris, France) was used as collision gas and was regulated at  $0.1 \text{ mL min}^{-1}$ . Analyses were performed in full scan and daughter scan modes, with a collision energy of 25 eV. Spectra were acquired between 80 and  $500 \text{ m/z}^{-1}$  and the data were treated with Micromass Mass-Lynx 4.1 software.

### **2.3.8. Ionic chromatography (CI)**

Generated carboxylic acids were identified by DIONEX DX120 ion chromatography equipped with a conductivity detector, using an anion exchange column AS19 ( $4 \times 250 \text{ mm}$ ) as the stationary phase, and a solution of KOH in water as the mobile phase. The analyses were carried out with a gradient elution mode. It started with  $10 \text{ mmol L}^{-1}$  of KOH during 10 min then increased linearly to  $45 \text{ mmol L}^{-1}$  from 10 to 25 min and finished with  $45 \text{ mmol L}^{-1}$  of KOH from 25 to 35 min. The flow rate was set at  $1 \text{ mL min}^{-1}$  (Yahiat et al., 2011).

## **3. Results and discussion**

### **3.1. Electrochemical behavior of enoxacin**

The electrochemical behavior of enoxacin was studied in order to conclude on the contribution of a direct oxidation and/or reduction of the target molecule at the electrode surface to the EF process. For this purpose, current potential curves were plotted by cyclic voltammetry in  $\text{Na}_2\text{SO}_4$   $50 \text{ mmol L}^{-1}$  supporting electrolyte at pH 3 with  $500 \text{ mg L}^{-1}$  of the target compound. In the presence of enoxacin, oxidation and reduction of water were shifted toward anodic and cathodic potentials, respectively. This potential shift was probably due to the adsorption of the target compound onto the electrode surface. A non reproducible signal in reduction was observed around  $-1.3 \text{ V/Ag/AgCl}$  (Supplementary material 1) which indicated an irreversible electrochemical reaction and the influence of the adsorption of the antibiotic on the peak current. The irreversible reduction of ENO has already been observed and has been attributed to a two electrons reduction of the ketone group of the molecule (Zhang et al., 1996 and Zhou et al., 1995).

The three amines groups were protonated at pH 3, thus no signal on their oxidation was observed on the voltammograms.

During the EF process, even if the ketone group was reduced by a direct two electrons transfer at the electrode interface, the formation of alcohol should not significantly enhance the biodegradability of the molecule.

### **3.2. Effect of the operating parameters on enoxacin degradation by EF**

The decrease of the ENO concentration during the EF was monitored and different operating conditions were examined, including the initial concentration of ENO, the initial Fe(II) concentration and the applied current intensity.

#### **3.2.1. Effect of the initial enoxacin concentration**

The electrolysis was carried out at  $200 \text{ mA}$ , with  $0.2 \text{ mmol L}^{-1}$  Fe(II) and with initial amounts of ENO in the range of  $50\text{--}200 \text{ mg L}^{-1}$  (Fig. 2).

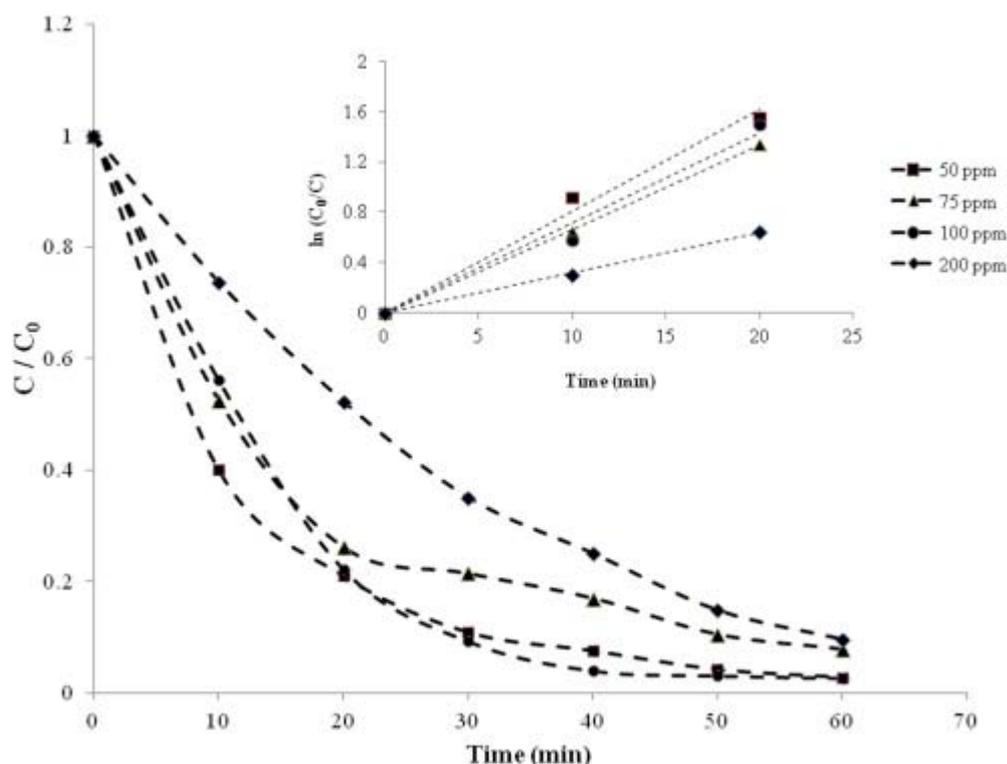


Fig. 2.

Effect of the initial ENO concentration on its degradation and kinetic analysis for the pseudo first-order reaction of ENO. Experimental conditions:  $[\text{Fe(II)}] = 0.2 \text{ mmol L}^{-1}$ ,  $I = 200 \text{ mA}$ ,  $\text{pH} = 3$ ,  $[\text{Na}_2\text{SO}_4] = 50 \text{ mmol L}^{-1}$ ,  $V = 250 \text{ mL}$ .

The results show a clear impact of the initial ENO amount on the degradation efficiency, especially between 100 and 200  $\text{mg L}^{-1}$  (Fig. 2); while almost similar final degradation yields can be observed after 60 min (Fig. 2). This trend can be explained by an increase of organic molecules in the solution which requires a greater number of hydroxyl radicals to their degradation (Sirés et al., 2007a). Experimental kinetics can be satisfactorily described by a pseudo-first order reaction kinetic (inset of Fig. 2). The apparent kinetic rate constant values ( $k_{\text{app}}$ ) for the oxidation of ENO correspond to the slopes of the linear fits of the plots of  $\ln([\text{ENO}]_0/[\text{ENO}])$  versus time (Table 1). The decrease of  $k_{\text{app}}$  can most likely be attributed to the gradual acceleration of competitive reactions between hydroxyl radicals and the oxidation intermediates of ENO that were formed during the process (Flox et al., 2007 and Sirés et al., 2007a). Furthermore, even if the amount of ENO increased in solution, the number of OH produced remained constant. As a consequence, hydroxyl radicals were the limiting reagent of the degradation of ENO (Flox et al., 2006).

Table 1.

Apparent kinetic rate constants  $k_{\text{app}}$  obtained during the EF treatment at various initial ENO concentrations, Fe(II) concentrations and applied current intensity.

Initial ENO concentration (mg L <sup>-1</sup> )	Fe(II) concentration (mmol L <sup>-1</sup> )	Applied current intensity (mA)	Apparent kinetic rate constant k <sub>app</sub> (min <sup>-1</sup> )
50	0.2	200	0.082
75	0.2	200	0.075
100	0.2	200	0.075
200	0.2	200	0.032
50	0.1	200	0.055
50	0.2	200	0.082
50	0.5	200	0.049
50	1	200	0.039
50	0.2	50	0.045
50	0.2	100	0.048
50	0.2	200	0.082
50	0.2	300	0.094

### 3.2.2. Effect of the Fe(II) concentration

The effect of Fe(II) on the degradation of the target molecule was investigated in the presence of 50 mg L<sup>-1</sup> of ENO and for an applied constant current intensity of 200 mA. The ENO degradation yields increased with increasing Fe(II) concentration from 0.1 to 0.2 mmol L<sup>-1</sup>, while it decreased above 0.2 mmol L<sup>-1</sup> (Fig. 3). On the one hand, the increase of the degradation efficiency observed between 0.1 and 0.2 mmol L<sup>-1</sup> can be attributed to the higher production of hydroxyl radicals for increasing amount of Fe(II), according to the Fenton reaction (2) (Panizza and Cerisola, 2009). On the other hand, the negative effect of the Fe(II) amount beyond 0.2 mmol L<sup>-1</sup> should be most likely attributed to a competitive reaction occurring between the hydroxyl radicals generated and the excess of ferrous ions according to the reaction (5) (Panizza and Cerisola, 2009 and Sun and Pignatello, 1993):



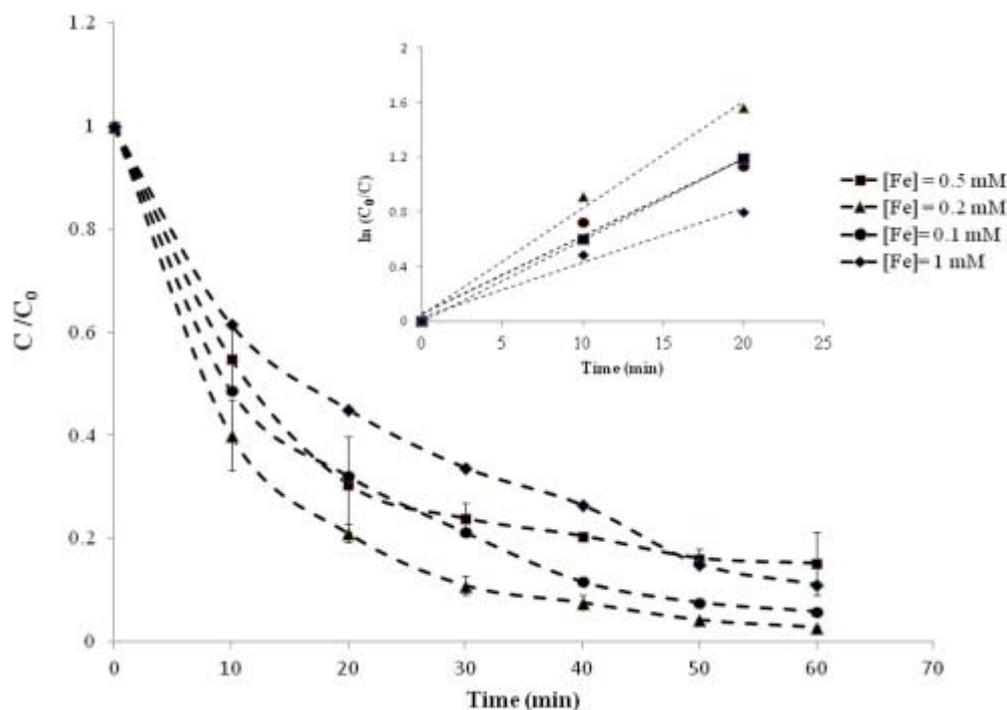


Fig. 3.

Effect of Fe (II) concentration on ENO degradation and kinetic analysis for the pseudo first-order reaction of ENO. Experimental conditions:  $C_0 = 50 \text{ mg L}^{-1}$ ,  $I = 200 \text{ mA}$ ,  $\text{pH} = 3$ ,  $[\text{Na}_2\text{SO}_4] = 50 \text{ mmol L}^{-1}$ ,  $V = 250 \text{ mL}$ .

The inset of the panel of Fig. 3 shows that experimental kinetics are well described by a pseudo-first order reaction rate. The values of the apparent rate constant  $k_{\text{app}}$  presented the same evolution than the degradation efficiency, they increased with increasing Fe(II) concentration until  $0.2 \text{ mmol L}^{-1}$  of catalyst and decreased beyond this Fe(II) amount (Table 1). As a consequence and in order to reach optimal results,  $0.2 \text{ mmol L}^{-1}$  of Fe(II) was employed as a catalyst concentration in the rest of the electrolyzes performed in this study.

### 3.2.3. Effect of the applied current intensity

The current intensity is a key parameter in the effectiveness of the EF process (Oturán et al., 2008 and Ozcan et al., 2009). To investigate its effect, experiments were carried out for  $50 \text{ mg L}^{-1}$  ENO and  $0.2 \text{ mmol L}^{-1}$  of Fe(II) and current intensities varying in the range 50–300 mA. The obtained results show that an increase in the current intensity improved the ENO degradation yields, from 74% after 30 min of electrolysis at 50 mA to 91% for 300 mA (Fig. 4). The improvement appeared more significant at the examination of  $k_{\text{app}}$  of the pseudo-first order kinetic model showing an increase from  $0.048 \text{ min}^{-1}$  to  $0.094 \text{ min}^{-1}$  for current intensities increasing from 50 to 300 mA (inset of Fig. 4 and Table 1). The increase of the degradation yield associated with the increase of the current intensity can be explained by an increase of hydrogen peroxide production (see Supplementary material 2) and an increase of the regeneration rate of Fe(II) at the cathode, leading to a higher amount of hydroxyl radicals produced (Oturán et al., 2008 and Ozcan et al., 2009).

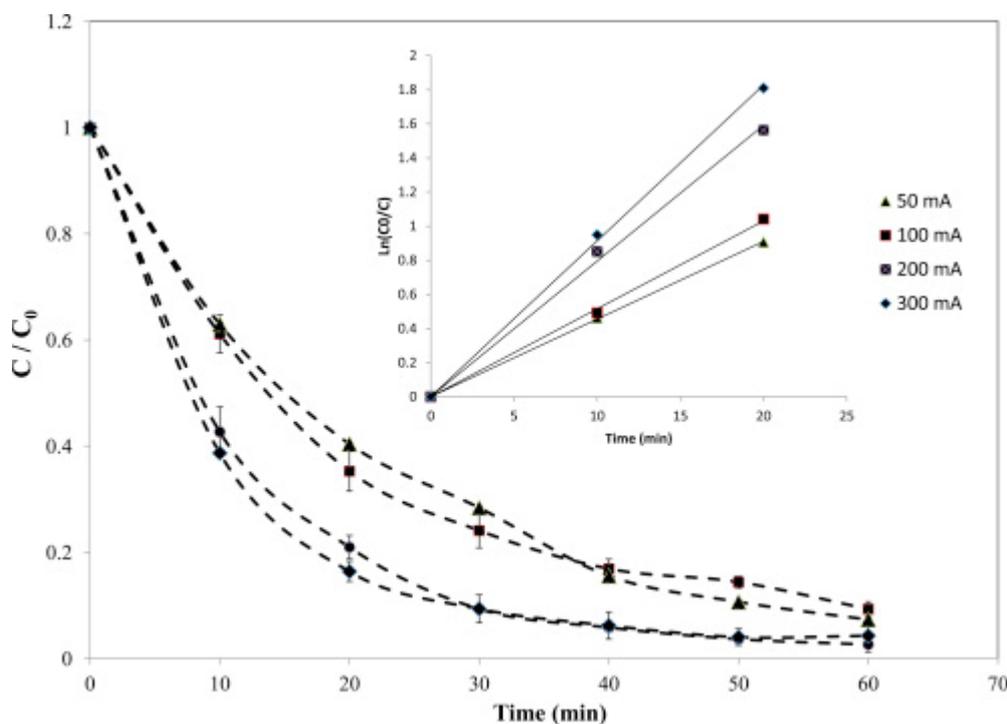


Fig. 4.

Effect of the applied current intensity on the ENO degradation and kinetic analysis for the pseudo first-order reaction of ENO. Experimental conditions:  $C_0 = 50 \text{ mg L}^{-1}$ ,  $[\text{Fe(II)}] = 0.2 \text{ mmol.L}^{-1}$ ,  $\text{pH} = 3$ ,  $[\text{Na}_2\text{SO}_4] = 50 \text{ mmol L}^{-1}$ ,  $V = 250 \text{ mL}$ .

Furthermore, the concentration of electrogenerated  $\text{H}_2\text{O}_2$  during electrolysis can decrease when the current intensity increase, since oxygen reduction can lead to water instead of  $\text{H}_2\text{O}_2$  at the cathode and owing to possible direct oxidation of  $\text{H}_2\text{O}_2$  in a monocompartment electrochemical cell at the anode (Brillas et al., 2009). These assumptions can account for the very close results obtained at 200 mA and 300 mA (Fig. 4 and Table 1).

Results obtained at 50 and 100 mA were similar; no significant effect on enoxacin degradation was observed. The apparent kinetic constants (Table 1) were then similar ( $0.045$  and  $0.048 \text{ min}^{-1}$  respectively).

### 3.3. Electro-Fenton efficiency on the mineralization of enoxacin

The above results have shown that the optimal conditions for the degradation of enoxacin by the electro-Fenton process were  $[\text{ENO}]_0 = 50 \text{ mg L}^{-1}$ ;  $I = 300 \text{ mA}$  and  $[\text{Fe(II)}] = 0.2 \text{ mmol L}^{-1}$ . To complete the analysis of the impact of this process on enoxacin removal, mineralization and oxidation were also monitored and are shown in Fig. 5. TOC and COD followed similar trends to that of enoxacin degradation, since a significant decrease occurred within 30 min of electrolysis, reaching 26 and 49% decrease respectively (Fig. 5). Refractory organic by-products were most likely responsible of the low mineralization yield (Sirés et al., 2007b). From 60 min of treatment, an almost total enoxacin abatement was

observed (97%), while mineralization and oxidation remained limited, 43 and 54% after 120 min, respectively (Fig. 5). The slight decrease of mineralization observed between 120 and 180 min of treatment has been previously attributed to the accumulation of organic recalcitrant aromatic rings (Babic et al., 2013). Furthermore, the amount of OH produced may be insufficient to mineralize the molecule.

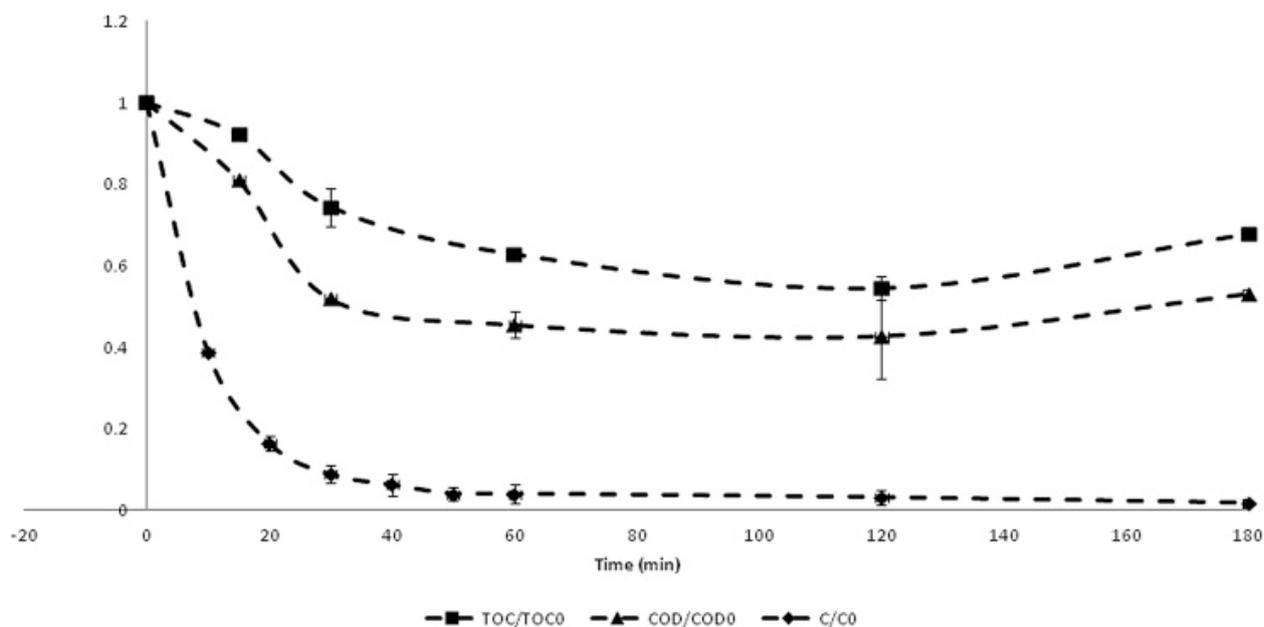


Fig. 5.

Time-course of ENO concentration, TOC and COD during ENO degradation by EF. Experimental conditions:  $C_0 = 50 \text{ mg L}^{-1}$ ,  $I = 300 \text{ mA}$ ,  $[\text{Fe(II)}] = 0.2 \text{ mM}$ ,  $\text{pH} = 3$ ,  $[\text{Na}_2\text{SO}_4] = 50 \text{ mmol L}^{-1}$ ,  $V = 250 \text{ mL}$ .

### 3.4. Enoxacin biodegradability study

The  $\text{BOD}_5$  on COD ratio is a recognized indicator of the biodegradability of a solution; this ratio must be higher or equal to 0.4 to consider that the solution is easily biodegradable (Salles et al., 2010). As expected, enoxacin was not biodegradable before electrolysis (Fig. 6). Biodegradability increased after 60 min electrolysis at 300 mA, with a  $\text{BOD}_5$  on COD ratio of 0.25, showing that primary by-products were relatively resistant to micro-organisms. However, a readily biodegradable solution, with a ratio of 0.51 was obtained after 180 min of electrolysis. It can be explained by the oxidation of the remaining recalcitrant by-products previously generated via the hydroxyl radicals (Mansour et al., 2012). However, for a longer electrolysis time, the obtained electrolyzed solution became less biodegradable since the  $\text{BOD}_5$  on COD ratio decreased from 0.51 to 0.30 after 360 min, in agreement with the related literature (Mousset et al., 2014), and which may be related to the accumulation of recalcitrant quinolone rings for increasing electrolysis time (Babic et al., 2013). From this, the relevance of an EF pretreatment was confirmed and 180 min electrolysis of the ENO solution is required for an effective biological treatment.

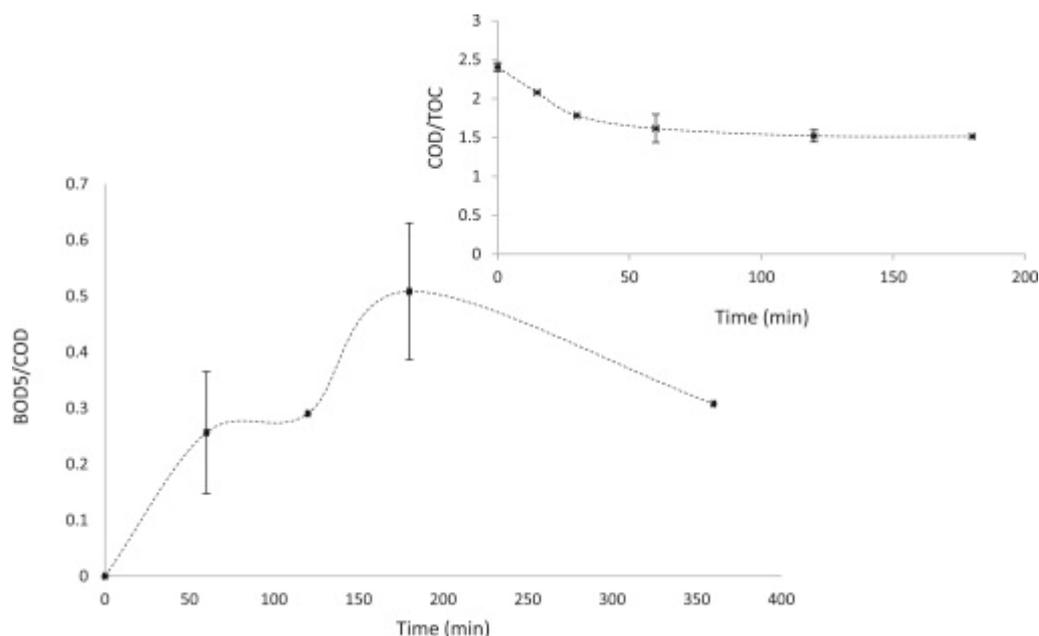


Fig. 6.

Evolution of the BOD<sub>5</sub> on COD ratio and the COD on TOC ratio during electrolysis. Experimental conditions:  $C_0 = 50 \text{ mg L}^{-1}$ ,  $I = 300 \text{ mA}$ ,  $[\text{Fe(II)}] = 0.2 \text{ mmol L}^{-1}$ ,  $\text{pH} = 3$ ,  $[\text{Na}_2\text{SO}_4] = 50 \text{ mmol L}^{-1}$ ,  $V = 250 \text{ mL}$ .

The relevance of the coupling of an EF pretreatment and a biological process can be also estimated from the examination of the evolution of the COD/TOC ratio, displayed in the inset of Fig. 6. A favorable trend is a decrease of this ratio (Chebli et al., 2010), which was experimentally observed after EF pretreatment. In accordance with the improvement of the solution's biodegradability, the COD/TOC ratio decreased from 2.40 for an untreated solution to 1.62 for 60 min of electrolysis. The ratio COD/TOC then slightly decreased to 1.51 after 180 min of treatment.

### 3.5. Evolution of the generated inorganic ions

The enoxacin molecule contains an atom of fluorine and three atoms of nitrogen. The progressive generation of inorganic ions such as fluorides  $\text{F}^-$ , nitrates  $\text{NO}_3^-$  and ammonium  $\text{NH}_4^+$  was therefore not surprising (Fig. 7 (a)). However, no nitrite  $\text{NO}_2^-$  anions were detected. The mineralization of enoxacin was therefore accompanied by the conversion of its heteroatoms into inorganic ions (El-Desoky et al., 2010). Ammonium concentration increased progressively until reaching a constant value of  $0.12 \text{ mmol L}^{-1}$  after 120 min of treatment. Nitrates remained constant throughout the electrolysis and did not exceed  $25 \text{ } \mu\text{mol L}^{-1}$ .

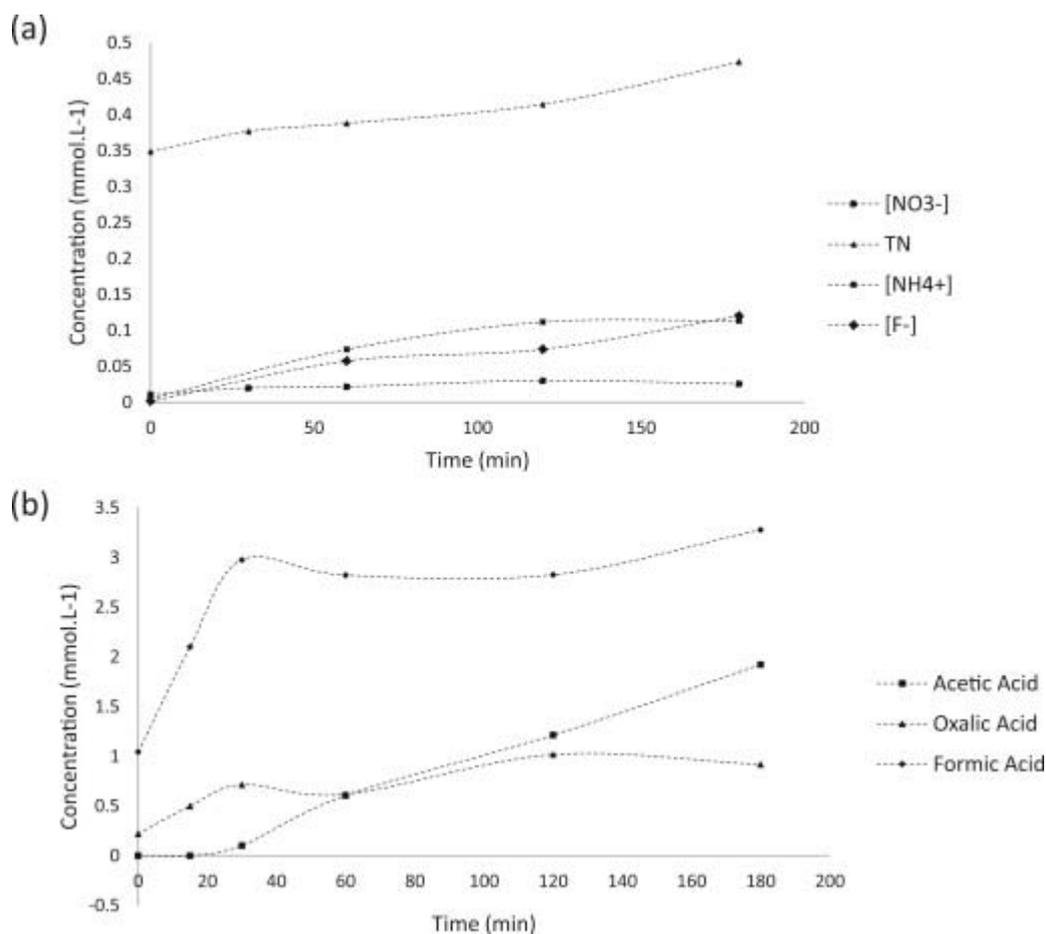


Fig. 7.

Time-course of the concentration of (a) inorganic species and (b) carboxylic acids during ENO degradation by EF. Experimental conditions:  $C_0 = 50 \text{ mg L}^{-1}$ ;  $I = 300 \text{ mA}$ ,  $[\text{Fe(II)}] = 0.2 \text{ mmol L}^{-1}$ ;  $[\text{Na}_2\text{SO}_4] = 50 \text{ mmol L}^{-1}$ ;  $t = 60 \text{ min}$ ,  $\text{pH} = 3$ ,  $V = 250 \text{ mL}$ .

24% of the nitrogen atoms of the target molecule were transformed into ammonium, while only 5% were transformed into nitrates after 120 min. The percentage of undetected nitrogen may be related to the presence of nitrogen in the degradation products, such as the stable N-derivatives found by LC-MS/MS (Garcia-Segura et al., 2012). The fluorides amount increased continuously until the end of electrolysis, reaching a value of  $0.12 \text{ mmol L}^{-1}$ , namely 77% of the fluorine likely to be found after a total conversion ( $0.16 \text{ mmol L}^{-1}$ ). The fluorine atoms unconverted into inorganic ions are likely to be included in the structure of some degradation by-products (Babic et al., 2013 and Garcia-Segura et al., 2012).

### 3.6. Study of the generated carboxylic acids

The degradation of aromatics and heteroatoms by hydroxyl radicals involves the generation of short-linear aliphatic carboxylic acids, produced from the cleavage of their aromatic moieties and lateral groups (Garcia-Segura et al., 2012). Based on this information, it can be noticed that the degradation of  $50 \text{ mg L}^{-1}$  of enoxacin by EF at 300 mA was also accompanied by the

generation of several carboxylic acids. Three of them were identified by Ion Chromatography, presenting well defined peaks: formic (6.22 min), acetic (5.44 min) and oxalic (21.08 min) acids. A progressive increase of the acids concentration was observed during the course of electrolysis (Fig. 7 (b)), with the higher concentration (more than 3 mg L<sup>-1</sup>) observed for formic acid (C-1 acid) compared to acetic and oxalic acids (C-2 acid). It can be interpreted by the transformation of the C-2 acids into a C-1 acid, due to the hydroxyl radicals' attack (Garcia-Segura et al., 2012). Note that oxalic and formic acids are ultimate carboxylic acids because they can be directly converted into CO<sub>2</sub> (Garcia-Segura and Brillas, 2011 and Oturan et al., 2008).

On the other hand, it has been reported that under EF conditions, carboxylic acids form complexes with Fe(III) present in the solution according to the Fenton reaction (reaction 3) (Brillas et al., 2009). This can explain the behavior of formic and oxalic acids (Fig. 7 (b)), which were accumulated within the first 30 min of electrolysis, reaching a plateau after this time. To account for this trend, it can be assumed that they probably reacted with Fe(III) and then their complexes rapidly reacted with hydroxyl radicals (Garcia-Segura and Brillas, 2011). In contrast, acetic acid was more recalcitrant and increased progressively, reaching 1.94 mg L<sup>-1</sup> after 180 min of treatment.

The carboxylic acids accumulated at the end of the electrolysis were involved in the residual TOC displayed in Fig. 5 (Garcia-Segura et al., 2012).

### 3.7. Elucidation of by-products and mechanism of degradation

Based on the results of the UPLC–MS/MS analysis, four by-products were detected after 30 and 60 min of treatment (Table 2), which allowed to suggest a general pathway for ENO degradation by EF (Fig. 8).

Table 2.

Intermediates products generated during the EF degradation of enoxacin.

Products	Retention time (min)	MH <sup>+</sup> (g mol <sup>-1</sup> )	Ion fragments (g mol <sup>-1</sup> )
Enoxacin	1.23	321	303
			257
			234
			206
			333
ENO I	1.68	351	305
			259
			232
			305
ENO II	1.8	323	259
			232
			262
			234
ENO III	1.8	280	234
			206

Products Retention time (min) MH<sup>+</sup> (g mol<sup>-1</sup>) Ion fragments (g mol<sup>-1</sup>)

Products	Retention time (min)	MH <sup>+</sup> (g mol <sup>-1</sup> )	Ion fragments (g mol <sup>-1</sup> )
			234
			206
ENO IV	2.02	252	178
			150
			123

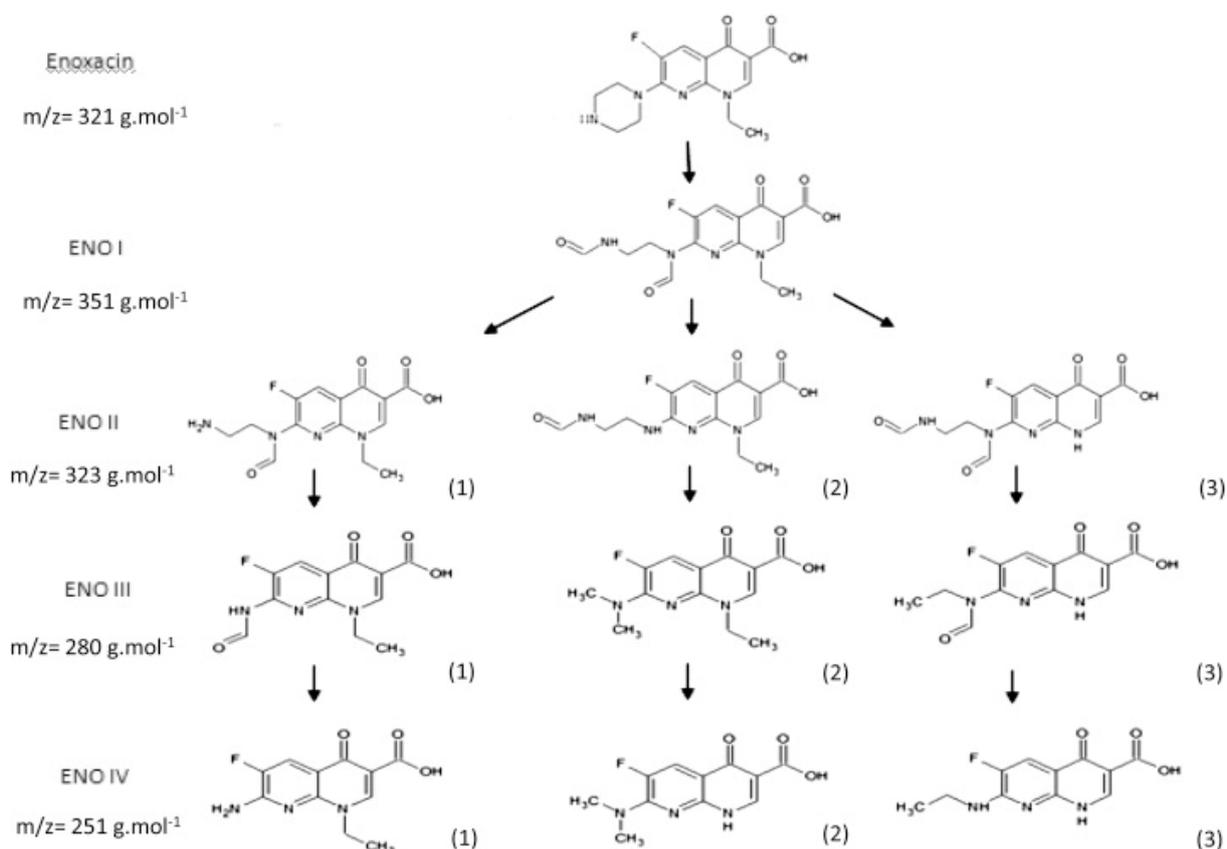


Fig. 8.

Proposed structures for by-products of enoxacin EF degradation.

There is no literature dealing with the degradation pathway of enoxacin. The proposed mechanism of ENO degradation is based on the mass spectra of intermediates, on the molecular weight of ion fragments derived from each degradation product and on previous studies done on other FQs (Babic et al., 2013, Dolar et al., 2013 and Sturini et al., 2012).

For each molecular mass found, various structures were suggested. Indeed, inorganic species time courses showed only a partial generation of fluorides, nitrates and ammonium, and hence the presence of by-products with or without heteroatoms is possible. Moreover, after 30 min of treatment, 87% of ENO was removed, so that the hydroxyl radicals generated can attack the by-products, accounting for the presence of a significant amount of short carboxylic acids after 30 min of degradation (Fig. 7 (b)).

According to the literature, ENO may be first attacked on its piperazine ring, seeing that it is the most oxidable part of the molecule (Babic et al., 2013). The structures of ENO I came principally from the opening of the piperazine ring and its transformation into two groups of amides (Babic et al., 2013). It was made up of oxidative rearrangements on the N-ethyl piperazine chain. The N-ethyl substituent of the benzo ring may also be oxidized on aldehyde group (not shown).

The second step consisted in the elimination of an ethyl group from the N-ethyl substituent or the elimination of a carbon monoxide (CO) molecule from one of the amide groups to obtain ENO II structures.

ENO III and ENO IV structures resulted from the successive elimination of an ethyl, a carbon monoxide and/or an ammonia molecule (NH<sub>3</sub>), in order to obtain diverse possible by-products, including a primary, a secondary or a tertiary amine in their structures (Babic et al., 2013 and Sturini et al., 2012).

In accordance with older studies, all the intermediates suggested showed the persistence of the quinolone ring in their structures, which explain the high residual TOC and COD in the treated solution.

Short carboxylic acids generated during electrolysis may derive from ENO IV by-products (Babic et al., 2013 and Vasconcelos et al., 2009).

The nomenclature of enoxacin intermediates suggested in this study is given in supplementary material (Supplementary material 3).

## 4. Conclusion

The present paper proves the effectiveness of the electro-Fenton treatment for the remediation of the fluoroquinolone enoxacin present in wastewaters and in surface water. Around 98% degradation yield was obtained after 60 min of electrolysis in the optimal experimental conditions, namely 50 mg L<sup>-1</sup> initial enoxacin amount, 300 mA and 0.2 mmol L<sup>-1</sup> Fe(II). Furthermore, biodegradability was enhanced after 180 min of treatment with a maximum BOD<sub>5</sub> on COD ratio of 0.51; while TOC removal yield did not exceed 32% after this electrolysis time. It can be explained by the generation of recalcitrant by-products during electrolysis and the persistence of the quinolone ring. This behavior was confirmed by the primary products found during UPLC–MS/MS analyses.

After 180 min electrolysis of 50 mg L<sup>-1</sup> ENO, 77% of the fluorine contained in the target compound was converted into fluorides. Moreover, the nitrogen content of ENO was partially converted to nitrate (5%) and ammonium (24%) ions. Oxalic, acetic and formic acids were also formed, which can be easily mineralized leading to CO<sub>2</sub> and H<sub>2</sub>O.

From the above results, EF is certainly suitable for the removal of ENO in wastewaters. It also presents a relevant solution and an effective pretreatment for the improvement of biodegradability. Further studies could be considered as a perspective in order to optimize the ideal electrolysis time for good biodegradability.

## Appendix A. Supplementary data

Attached

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