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# *Petromyzon marinus* (Petromyzontidae), an unusual host for helminth parasites in western Europe

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ABSTRACT: The sea lamprey *Petromyzon marinus*, which is among the most phylogenetically ancient vertebrates, is a hematophagous ectoparasite that feeds on vertebrates and is considered vulnerable in Europe but is a pest in the North American Great Lakes. We conducted a literature review of helminth parasites of *P. marinus* and investigated postmetamorphic lampreys sampled in rivers and northeast Atlantic coastal waters (western France) during spawning migration. Based on the literature review, 16 helminth taxa have been recorded in *P. marinus*, among them 14 in North America but only 2 in Europe, with no species in common between these areas. Specific parasites are lacking, and helminth parasites recorded in P. marinus are mostly opportunistic and are trophically transmitted to fish hosts with both extremely low prevalence and mean intensity. Thus, P. marinus seems an unusual host that is probably infected through accidental ingestion of parasites by microphagous larvae (ammocoetes) and/or hematophagous postmetamorphs. Our field study supports this hypothesis, since only a single third-stage larva of Anisakis simplex sensu stricto was found in 2 postmetamorphic P. marinus among the 115 individuals dissected. This opportunistic, trophically transmitted, and cosmopolitan nematode species has never been recorded in North American sea lampreys and only once in Galician rivers (southern Europe). Infestation pathways of *P. marinus* by *A. simplex* are proposed vis-à-vis the feeding strategy of postmetamorphs and fish host species which potentially harbor anisakid larvae in their musculature. More generally, the complexity of biotic interactions is discussed considering *P. marinus* both as a host for helminth parasites and as a parasite for hosts such as fish and mammals, which are also potential predators of sea lamprey.

KEY WORDS: Sea lamprey  $\cdot$  Helminth parasites  $\cdot$  Unusual host  $\cdot$  Accidental ingestion  $\cdot$  Anisakis simplex sensu strict  $\cdot$  Nematode

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#### INTRODUCTION

The anadromous sea lamprey *Petromyzon marinus* is a fascinating biological model, as it is among the most phylogenetically ancient vertebrates and is a survivor of the jawless group Agnatha, whose life style and body structure have been highly conserved for at least 350 million years (see Hardisty 2006, Taverny & Elie 2010 for reviews). This species is also uncommon among vertebrates, as it is a parasitic hematophage after metamorphosis, feeding on various aquatic vertebrates (at least 36 fish species, and some marine mammals such as whales and dolphins) and involving a prolonged attachment (up to 83 d) to the body surface of its hosts (Farmer 1980, Hardisty 2006, Taverny & Elie 2010). Because of its feeding strategy, *P. marinus* is widely dispersed through transport by its diverse hosts, and does not home to its native river but rather exhibits regional panmixia (Waldman et al. 2008).

Native to the North Atlantic, *P. marinus* is distributed within the Atlantic drainages of North America from the coast of Labrador as far south as Florida, and throughout western Europe from Varanger Fjord in Norway to the western Mediterranean (Beamish 1980, Hardisty 2006, Taverny & Elie 2010). Despite their high migratory capacity which may extend over several hundred kilometers, North American and European populations of *P. marinus* are isolated, as shown by their genetic differences; the separation presumably occurred during the Cretaceous period, some 70 million years ago (Beamish 1980, Hardisty 2006, Taverny & Elie 2010).

In Europe, *P. marinus* is considered a vulnerable species (listed in Annex II of the EU Habitats Directive and Annex III of the Bern Convention), but has been classified as being of 'Least Concern' by the IUCN since 2008 (Taverny & Elie 2010). In contrast, *P. marinus* is considered a pest in the Great Lakes of North America, which it invaded in the 1930s (Hardisty 2006). In these lakes, the sea lamprey is landlocked in inland waters, and the hematophagous postmetamorphs cause tremendous damage to fish stocks, with a consequent yearly expenditure of millions of dollars for their control (e.g. Bence et al. 2003, Neeson et al. 2007).

To date, the parasitofauna of *P. marinus* has been mainly studied in the North American Great Lakes in the context of pest control research (Applegate 1950, McLain 1952, Guilford 1954, Bangham 1955, Wilson & Ronald 1967, Muzzall & Whelan 2011), whereas parasitological data are scarce in Europe, originating only from countries of the former USSR (Sobecka et al. 2009), or focusing on 1 parasite taxon (Bao et al. 2013).

In this study, we conducted a literature review of helminth parasites of *P. marinus* throughout its entire geographical distribution. We also investigated parasites in postmetamorphic lampreys sampled in French rivers and northeast Atlantic coastal waters (Bay of Biscay). Based on this literature review and our field investigation, *P. marinus* appears to be an unusual host that is likely infected through accidental ingestion of helminth parasites. The occurrence of these opportunistic and trophically transmitted helminths is discussed in terms of the complexity of the biotic interactions (parasitism, predation) involving the sea lamprey as a host for helminth parasites and as a parasite of vertebrate hosts (fish and mammals), some of which are also potential predators of the sea lamprey itself.

#### MATERIALS AND METHODS

The European 'Natura 2000' networking program, which aims to protect and preserve the biodiversity of lampreys, provided a total of 115 *Petromyzon marinus* postmetamorphs. These were caught by professional fishermen in 3 French rivers (Dordogne [n = 17], Garonne [n = 30], and Loire [n = 24]) and in the northeast Atlantic coastal waters (south Brittany in the Bay of Biscay [n = 44]) in May 2010 and February and March 2011.

All sea lampreys were frozen prior to being searched for helminth parasites in various organs (skin, gills, heart, body cavity, digestive system, liver, and gonad) and dissected using a binocular microscope. Parasites were morphologically identified and preserved in alcohol (70%) for molecular identification via DNA sequencing (cytochrome oxidase subunit 2, *cox2*).

Total DNA of parasites (i.e. nematodes of the family Anisakidae) was extracted from all individual nematode tissues using a ChargeSwitch<sup>®</sup> Forensic DNA Purification Kit (Invitrogen). A 530 bp fragment of the cox2 gene was amplified using the primers F-univnem 5'GGT GTT CTT TCT TTT GTT TCT G and R-univ-nem 5'GTG GGG CTA ACC ATA GTT TTA T specifically designed in the framework of the French National Research Agency (ANR) Fish-Parasites project (data not yet published) to amplify DNA of Anisakidae. PCR amplification was carried out in a total of 50 µl with the following composition: 0.5 to 2.5 µl DNA template was added to a PCR mix consisting of 0.2 mM of each dNTP (Interchim), 2 mM MgCl<sub>2</sub>, 10 µl 5× buffer, 1.25 units of GoTaq<sup>®</sup> polymerase (Promega), 0.2 µM of each primer and molecular biology grade water (Interchim) to adjust to the final volume. Conditions of cycling were as follows: an initial denaturing step at 95°C for 5 min, followed by 35 cycles of denaturing at 95°C for 30 s, annealing at 48°C for 30 s, and extension at 72°C for 40 s, and a final extension step at 72°C for 7 min. Automated DNA sequencing was performed by Genoscreen (Lille) using the primer R-univ-nem. Sequences were analyzed using BioEdit software and compared with nucleotide sequences from GenBank (National Center for Biotechnology Information) with the Basic Local Alignment Search Tool.

The parasite populations were described by prevalence (number of hosts infected with a particular parasite species / number of examined hosts) and mean intensity (total number of parasites of a particular species found in a sample / number of hosts infected with that parasite) (Bush et al. 1997). Mean values of data are reported  $\pm$  SE.

#### RESULTS

#### Literature review of helminth parasites of *Petromyzon marinus*

Helminth parasites have rarely been studied in *P. marinus*, particularly in Europe (2 references vs. 5 in North America; Table 1). In total, 16 helminth para-

site taxa have been recorded in *P. marinus* to date, among them 14 in North America but only 2 in Europe, with no species in common between these 2 areas (Table 1). All helminth parasites recorded in *P. marinus* are heteroxenous and, except for *Cystidicola stigmatura* (adult in salmonid swimbladder), they become adults in the digestive tract of a piscivorous vertebrate.

#### Helminth parasites in postmetamorphs from French rivers and coastal waters

Only 2 individuals (from the Dordogne in February 2011 and from the Garonne in May 2010) were parasitized among the 115 dissected *P. marinus* (prevalence =  $1.74 \pm 0.01\%$ ), and only by a single nematode

Table 1. Helminth parasites (16 taxa) recorded in sea lamprey *Petromyzon marinus* in its geographical range (Eu: Europe; Am: North American Great Lakes), with information on parasite development stages (L: larval; I: immature; A: adult), infected organs, and prevalence; na: not available

Parasite taxon	Stage	Infected organ	Range	Prevalence (%)	Reference
Trematoda					
Diplostomum sp.	L	Eye lens	Am	7.7	Bangham (1955)
Diplostomum huronensis	L	Eye lens	Am	10.6	Wilson & Ronald (1967)
Podocotyle lepomis	A?	Digestive tract	Am	0.1	Wilson & Ronald (1967)
Cestoda					
<i>Eubothrium</i> sp.	Ι	Digestive tract	Am	3.0	Applegate (1950)
				1.0	McLain (1952)
				2.6	Guilford (1954)
Proteocephalus exiguus	Ι	Digestive tract	Am	45.5	Guilford (1954)
Proteocephalus laruei	А	Digestive tract	Am	15.4	Bangham (1955)
Proteocephalus sp.	I or A	Digestive tract	Am	61.5	Bangham (1955)
				0.7	Wilson & Ronald (1967)
Triaenophorous crassus	L or I	Digestive tract	Am	3.0	Applegate (1950)
		0		1.0	McLain (1952)
				2.6	Guilford (1954)
				15.4	Bangham (1955)
				0.4	Wilson & Ronald (1967)
Acanthocephala					
Echinorhynchus leidyi	А	Digestive tract	Am	0.2	McLain (1952)
		-		5.2	Guilford (1954)
Echinorhynchus salmonis	А	Digestive tract	Am	17.0	Applegate (1950)
		5		9.5	McLain (1952)
				9.1	Guilford (1954)
				7.7	Bangham (1955)
				8.3	Wilson & Ronald (1967)
Neoechinorhynchus cylindratus	A?	Digestive tract	Am	1.3	Guilford (1954)
Nematoda		-			
Anisakis simplex sensu stricto	L	Body cavity	Eu	56.3	Bao et al. (2013)
		1 1		1.7	This study
<i>Camallanus</i> sp.	А	Liver	Am	1.0	McLain (1952)
Cystidicola stigmatura	Ι	Digestive tract	Am	1.3	Guilford (1954)
Hysterothylacium aduncum	na	na	Eu	na	Polyanskii (1955)
Truttaedacnitis stelmioides	L	Various organs	Am	1.8	Wilson & Ronald (1967)

(mean intensity =  $1.00 \pm 0.00$ ) at the third larval stage located on the gonadal mesorchium. According to the DNA sequence (*cox2*), the parasite was unambiguously identified as *Anisakis simplex* sensu stricto (Anisakidae). In fact, based on 460 DNA base pairs, the query sequence was 100 % covered and had 100 % identity with 4 *A. simplex* sequences (KC810002, KC480042, GQ338430, and GQ338433) present in the GenBank nucleotide database.

#### DISCUSSION

The helminth parasitofauna of *Petromyzon marinus* lacks specific forms that have co-evolved with its host and is mainly composed of opportunistic fish parasites (Table 1). Most of them are cosmopolitan and have been described in other host species both in North America and in Europe (Chappell et al. 1994, Khalil et al. 1994, Moravec 1994, Mattiucci & Nascetti 2008). Despite this, no common helminth parasites are found between *P. marinus* from North America and Europe, thus reinforcing the idea of anciently separated North American and European populations (Beamish 1980, Hardisty 2006, Taverny & Elie 2010).

The occurrence of helminths in *P. marinus* is often sporadic or even accidental, with a prevalence of  $\leq 5\%$  (Table 1) and only 1 or 2 specimens found per host (McLain 1952, Sobecka et al. 2009). In some circumstances, the prevalence of some helminths can reach significant values, such as for Proteocephalus sp. (61.5%) and P. exiguus (45.5%) in North America, or Anisakis simplex sensu stricto (56.3%) in Europe (Table 1). Except for Diplostomum spp., most of the parasites listed in Table 1 are probably trophically transmitted to the sea lamprey. First, filter-feeding lamprey ammocoetes (larvae) may accidentally ingest infective helminths in diverse forms (eggs, free larvae, or parasitized intermediate hosts, such as crustaceans for Proteocephalus spp.), as suggested for Truttaedacnitis stelmioides by Moravec (1980). Second, hematophagous postmetamorphic lamprey can ingest helminth parasites encysted in fish flesh (e.g. plerocercoids of Triaenophorous crassus or thirdstage larvae of A. simplex; e.g. McLain 1952, Angelucci et al. 2011), because the anticoagulant lamphredin secreted by their buccal glands has cytolytic effects on host tissues (Farmer 1980).

Concerning *A. simplex* sensu stricto, the single helminth species recorded in our field study, its definitive hosts are marine cetaceans, such as dolphins, which are usually infected by eating intermediate hosts (zooplankton) or paratenic hosts (fish, cephalopods) containing third-stage larvae (see Mattiucci & Nascetti 2008 for a review). The first record of these third-stage larvae in *P. marinus* occurred in 2012 in 2 Galician rivers (southern Europe) with a much higher prevalence  $(60.0 \pm 0.5\%$  and  $42.9 \pm$ 1.9%) than in our study, but a comparable intensity  $(2.4 \pm 1.7 \text{ and } 1.0 \pm 0.0; \text{Bao et al. } 2013)$ . *P. marinus* is therefore a potential paratenic host which probably acquires anisakid larvae by feeding on infected fish acting as paratenic hosts for the nematode (Table 2). A. simplex larvae are never found in the blood of their fish hosts, but generally in the body cavity, and sometimes in the flesh, as shown for cod, herring, or mackerel (Petrie et al. 2005, Mattiucci & Nascetti 2008, Angelucci et al. 2011). Thus, when postmetamorphic sea lampreys feed on blood and, to a much lesser extent, on tissue cytolysis products, they can ingest anisakid larvae potentially encapsulated in fish muscle. To date, no studies have mentioned marine cetaceans among the fish, reptiles, birds, and mammals known to be predators of P. marinus (see Cochran 2009 for a review).

Based on the parasitism patterns of the 16 helminth taxa found in *P. marinus* to date, we can conclude that helminth parasites are neither a threat to vulnerable European populations nor a potential tool to control pest populations in the North American Great Lakes. Further research on the ecological foraging of *P. marinus* and trophic interactions is needed to determine whether *P. marinus* constitutes a dead end for these helminth parasites or is beneficial as an intermediate or paratenic host that is preyed on by the definitive host. Further investigations are also needed to better understand the lack of specific parasites in *P. marinus*, particularly regarding the immunological abilities of *P. marinus* to ward off parasitic infection.

Table 2. Fish species recorded as hosts of both sea lamprey *Petromyzon marinus* (Beamish 1980, Farmer 1980, Taverny & Elie 2010, Silva et al. 2013) and *Anisakis* third-stage larvae (Berland 1961, Petrie et al. 2005, Angelucci et al. 2011, Mladineo et al. 2011)

Order	Host species	
Clupeiformes	Clupea harengus (herring)	
Gadiformes	Gadus morhua (Atlantic cod)	
	Merluccius spp. (European hake)	
	Pollachius virens (pollock)	
Perciformes	Scomber scombrus (Atlantic mackerel)	
	Thunnus thynnus (Atlantic bluefin tuna)	
Salmoniformes	Salmo trutta (brown trout and sea trout) <sup>a</sup>	
<sup>a</sup> S. trutta is also known to prey upon sea lamprey (Apple- gate 1950)		

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